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eZKine[™] CD8 Activation 1 Whole Blood Intracellular Cytokine Kit

Catalog Number: 8822-6854

RUO: For Research Use Only. Not for use in diagnostic procedures.



Staining of human whole blood with the eZKine [™] CD8 Activation 1 Whole Blood Intracellular Cytokine Kit. Freshly isolated whole blood was stimulated for 5 hours with Cell Stimulation Cocktail (plus protein transport inhibitors) (500X) (cat. 00-4975). After stimulation, samples were fixed and lysed with eZKine Fix/Lyse buffer, washed with Permeabilization Buffer, and stained with the CD8 Activation 1 Cocktail. Total cells were gated for CD3 APC staining (top left) and then analyzed for staining of CD8 PerCP-eFluor® 710, IFN gamma FITC, and CD69 PE as indicated. Staining of the Isotype Control Cocktail D is indicated in pink (top right).

Product Information

Contents: eZKine[™] CD8 Activation 1 Whole Blood Intracellular Cytokine Kit

 REF
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 Handling Conditions:
 Use within 6 months of opening or by date indicated on the bottle

Temperature Limitation: Store at 2-8°C. Do not freeze. Light-sensitive material.
 Batch Code: Refer to vial
 Use By: Refer to vial

Contains sodium azide and formaldehyde

Description

This eZKine CD8 Activation 1 Whole Blood Intracellular Cytokine Kit is designed to rapidly identify cytokine producing T lymphocytes of the CD8+ T cell lineage after stimulation of whole peripheral blood samples. Stimulation of whole blood in the presence of Protein Transport Inhibitor Cocktail (cat. 00-4980, not included) is followed by red blood cell lysis and fixation in a single step and subsequent permeabilization. Samples are then ready to stain with the CD8 Activation 1 cocktail and concentration-matched Isotype Control cocktail.

CD8+ T cells, or cytotoxic T lymphocytes, are immune cells that are responsible for the detection and clearance of virally or bacterically infected host cells. Upon recognition of antigen displayed on the surface of infected cells, activated CD8+ T cells upregulate markers of early activation such as CD69 and FAS ligand, followed by the release of granules containing granzymes and perforin as well as secretion of cytokines such as IFN gamma and TNF alpha. Granzymes and perforin act in concert to directly lyse infected target cells, while cytokines have paracrine effects including the recruitment and activation of macrophages, lymphocytes, and PMNs. IFN gamma is important for the upregulation of MHC class I and II on nearby cells as well as promoting Th1 cell differentiation.

Components

eZKine CD8 Activation 1 Cocktail (cat. 22-7785-71): 25 tests. Store at 2-8°C. This cocktail contains the following antibodies:

Anti-Human CD3 (SK7) APC

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Anti-Human CD8 (SK1) PerCP-eFluor® 710 Anti-Human CD69 (FN50) PE Anti-Human IFN gamma (4S.B3) FITC **eZKine Isotype Control Cocktail D** (cat. 22-7790-71): 25 tests. Store at 2-8°C. This cocktail contains the following antibodies: Anti-Human CD3 (SK7) APC Anti-Human CD8 (SK1) PerCP-eFluor® 710 Mouse IgG1 K Isotype Control PE Mouse IgG1 K Isotype Control FITC **eZKine Fix/Lyse Concentrate (4X)**: 15 mL. Store at 2-8°C. Avoid agitation. **eZKine Fix/Lyse Diluent**: 50 mL. Store at 2-8°C. **Permeabilization Buffer (10X)**: 20 mL. Store at 2-8°C. *Note: The Permeabilization Buffer (10X) has a natural*

tendency to precipitate, however, its function is not affected by this. To clarify, the solution can be filtered after dilution to 1X working solution.

Applications Reported

eZKine[™] CD8 Activation 1 Whole Blood Intracellular Cytokine Kit has been reported for use in intracellular staining followed by flow cytometric analysis.

Applications Tested

The eZKine[™] CD8 Activation 1 Whole Blood Intracellular Cytokine Kit has been pre-titrated and tested by intracellular staining and flow cytometric analysis of stimulated human whole blood following the eZKine protocol. The CD8 Activation 1 Cocktail and Isotype Control Cocktail can be used at 20 µL per test. A test is defined as the amount of antibody that will stain a cell sample in a final volume of 100 µL. Cell number should be determined empirically but can range from 10⁵ to 10⁸ cells/test.

References

Riou C, Treurnicht F, Abrahams MR, Mlisana K, Liu MK, Goonetilleke N, Koup R, Roederer M, Abdool Karim S, de Bruyn G, Williamson C, Gray CM, Burgers WA; CAPRISA 002 Study Team. Increased memory differentiation is associated with decreased polyfunctionality for HIV but not for cytomegalovirus-specific CD8+ T cells. J Immunol. 2012 Oct 15;189(8):3838-47.

Zhang N, Bevan MJ. CD8(+) T cells: foot soldiers of the immune system. Immunity. Aug 26;35(2):161-8.

Precopio ML, Betts MR, Parrino J, Price DA, Gostick E, Ambrozak DR, Asher TE, Douek DC, Harari A, Pantaleo G, Bailer R, Graham BS, Roederer M, Koup RA. Immunization with vaccinia virus induces polyfunctional and phenotypically distinctive CD8(+) T cell responses. J Exp Med. 2007 Jun 11;204(6):1405-16.

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Russell JH, Ley TJ. Lymphocyte-mediated cytotoxicity. Annu Rev Immunol. 2002;20:323-70.

Related Products

00-4975 eBioscience[™] Cell Stimulation Cocktail (plus protein transport inhibitors) (500X) 00-4980 eBioscience[™] Protein Transport Inhibitor Cocktail (500X) 01-1111 OneComp[™] eBeads Compensation Beads 16-0289 eBioscience[™] Anti-Human CD28 Functional Grade Purified (CD28.2) 16-0499 eBioscience[™] Anti-Human CD49d (Integrin alpha 4) Functional Grade Purified (9F10) 8822-6856 eZKine[™] 4-Color Compensation Control Kit

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eZKine[™] Whole Blood Intracellular Cytokine Staining Kit

Protocol 1: eZKine™ Whole Blood Intracellular Cytokine Staining

Materials Provided

Refer to the components section of the datasheet.

Other Materials Needed

- 12 x 75 mm round bottom test tubes
- Flow Cytometry Staining Buffer (cat. 00-4222)
- [Optional] Cell Stimulation Cocktail (plus protein transport inhibitors) (500X) (cat. 00-4975)
- [Optional] Protein Transport Inhibitor Cocktail (500X) (cat. 00-4980)

Time Requirements

- Blood stimulation, 4-6 hours recommended
- Fixation/lysis, 25 minutes (or up to 3 days)
- Permeabilization, 10 minutes
- Staining, 30 minutes
- Final washing, 5-10 minutes

Experimental Procedure

Stimulation of Whole Blood

- 1. Use blood collected with sodium heparin anticoagulant. Other anticoagulants, such as sodium citrate and EDTA, may interfere with lymphocyte activation.
- 2. Split blood into 2 separate sterile conical tubes. Label one for the stimulation and the other for the unstimulated control.
 - \circ Generally, calculate the volume of blood to stimulate based on 100 μL of blood per stained sample specified in the experimental design.
 - \circ $\,$ Volumes of blood larger than 5 mL per 15 mL conical tube have not been tested.

3. Stimulation:

- Add Cell Stimulation Cocktail plus Protein Transport Inhibitors (cat. 00-4975) at 1:500 directly to the blood in the stimulation tube.
- Alternatively, add stimulant(s) as specified by a given experimental design. Be sure to add Protein Transport Inhibitor Cocktail (cat. 00-4980) to these stimulations as well.
- Add Protein Transport Inhibitor Cocktail (cat. 00-4980) at 1:500 directly to the blood in the unstimulated tube.
- 4. Incubate for 4-6 hours in a 37°C incubator with 5% CO₂. Loosen the cap on the tube to allow for gas exchange.
 - Alternative incubation times may be appropriate as specified by the experimental design.

Lysis, Fixation, Permeabilization and Staining of Blood

- 5. Prepare buffers:
 - Prepare fresh eZKine Fix/Lyse working solution by diluting eZKine Fix/Lyse Concentrate (1 part) with eZKine Fix/Lyse Diluent (3 parts). You will need 1 mL of the Fixation/Permeabilization working solution for each sample.
 - Prepare a 1X working solution of Permeabilization Buffer by diluting 10X Permeabilization Buffer with distilled water prior to use. You will need ~6 mL of 1X Permeabilization Buffer for each sample.
- 6. Aliquot 100 μL of stimulated or unstimulated blood per 12 x 75 mm round bottom tube.
- 7. Add 1 mL of freshly-prepared eZKine Fix/Lyse Buffer and incubate for 25 minutes to 2 hours at room temperature or up to 3 days at 2-8°C.
- 8. Add 2 mL of 1X Permeabilization Buffer and centrifuge at 600 x g at room temperature for 4 minutes. Discard the supernatant.
- 9. Add 2 mL of 1X Permeabilization Buffer and centrifuge at 600 x *g* at room temperature for 4 minutes. Discard the supernatant and pulse vortex the sample to completely dissociate the pellet.
- 10. Add 20 µL of the staining cocktail or the isotype control cocktail to the appropriate tubes and incubate at room temperature for 30 minutes while protecting from light.
- 11. Add 2 mL 1X Permeabilization Buffer and centrifuge at 600 x g at room temperature for 4 minutes. Discard the supernatant.

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- 12. Add 2 mL of Flow Cytometry Staining Buffer and centrifuge at 600 x g at room temperature for 4 minutes. Discard the supernatant.
- 13. Resuspend stained cells in an appropriate volume of Flow Cytometry Staining Buffer. Store at 2-8°C while protecting from light until ready to analyze on a flow cytometer.
 - For storage of up to 3 days prior to analysis, we recommend adding 100 µL of IC Fixation Buffer (cat. 00-8222) to the residual volume in the tube (typically ~100 µL of cells). Store at 2-8°C while protecting from light until ready to analyze on a flow cytometer.

Protocol 2: eZKine[™] Whole Blood Intracellular Cytokine Staining in 96-well plates

Materials Provided

Refer to the components section of the datasheet.

Other Materials Needed

- 96-well tissue culture plates
- Flow Cytometry Staining Buffer (cat. 00-4222)
- [Optional] Cell Stimulation Cocktail (plus protein transport inhibitors) (500X) (cat. 00-4975)
- [Optional] Protein Transport Inhibitor Cocktail (500X) (cat. 00-4980)

Time Requirements

- Blood stimulation, 4-6 hours recommended
- Fixation/lysis, 25 minutes (or up to 3 days)
- Permeabilization, 15 minutes
- Staining, 30 minutes
- Final washing, 5-10 minutes

Experimental Procedure

Stimulation of Whole Blood

1. Use blood collected with sodium heparin anticoagulant. Other anticoagulants, such as sodium citrate and EDTA, may interfere with lymphocyte activation.

- 2. Pipet 25 μ L of whole blood into the wells of a 96-well plate.
 - A 10-fold dilution of whole blood with eZKine Fix/Lyse buffer is necessary for complete lysis. If using a standard 96-well plate, 25 μL of blood is the maximum recommended to accommodate 250 μL of eZKine Fix/Lyse buffer used in Step 6.
- 3. Stimulation:
 - Add Cell Stimulation Cocktail plus Protein Transport Inhibitors (cat. 00-4975) at 1:500 directly to the blood in the appropriate well(s).
 - Alternatively, add stimulant(s) as specified by a given experimental design. Be sure to add Protein Transport Inhibitor Cocktail (cat. 00-4980) to these stimulations as well.
 - o Add Protein Transport Inhibitor Cocktail (cat. 00-4980) at 1:500 directly to the blood in the unstimulated control well(s).
- 4. Cover the plate and incubate for 4-6 hours in a 37°C incubator with 5% CO₂.
 - Alternative incubation times may be appropriate as specified by the experimental design.

Lysis, Fixation, Permeabilization and Staining of Blood

- 5. Prepare buffers:
 - Prepare fresh eZKine Fix/Lyse working solution by diluting eZKine Fix/Lyse Concentrate (1 part) with eZKine Fix/Lyse Diluent (3 parts). You will need 250 µL of the Fixation/Permeabilization working solution for each well (approximately 25 mL per plate).
 - Prepare a 1X working solution of Permeabilization Buffer by diluting 10X Permeabilization Buffer with distilled water prior to use. You will need approximately 1 mL of 1X Permeabilization Buffer for each well (approximately 100 mL per plate).
- 6. Add 250 μL of freshly-prepared eZKine Fix/Lyse Buffer to each well and incubate for 25 minutes to 2 hours at room temperature or up to 3 days at 2-8°C.
- 7. Centrifuge at 600 x g at room temperature for 4 minutes. Decant or aspirate the supernatant.
- 8. Add 250 μL of 1X Permeabilization Buffer to each well and centrifuge at 600 x *g* at room temperature for 4 minutes. Decant or aspirate the supernatant.
- 9. Add 250 µL of 1X Permeabilization Buffer to each well and centrifuge at 600 x *g* at room temperature for 4 minutes. Decant or aspirate the supernatant and pulse vortex the sample to completely dissociate the pellet.

- 10. Add 100 μL of 1X Permeabilization Buffer to each well.
- 11. Add 20 µL of the staining cocktail or the isotype control cocktail to the appropriate wells and incubate at room temperature for 30 minutes while protecting from light.
- 12. Add 250 μ L of 1X Permeabilization Buffer and centrifuge at 600 x *g* at room temperature for 4 minutes. Decant or aspirate the supernatant.
- 13. Add 250 µL of Flow Cytometry Staining Buffer and centrifuge at 600 x g for 4 minutes. Decant or aspirate the supernatant.
- 14. Resuspend stained cells in an appropriate volume (100-200 µL) of Flow Cytometry Staining Buffer. Store at 2-8°C while protecting from light until ready to analyze on a flow cytometer.
 - For storage of up to 3 days prior to analysis, we recommend adding 100 μL of IC Fixation Buffer (cat. 00-8222) to the residual volume in each well. Store at 2-8°C while protecting from light until ready to analyze on a flow cytometer.

Documentation and support

Customer and technical support

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 - Certificates of Analysis
 - Safety Data Sheets (SDSs; also known as MSDSs)

Note: For SDSs for reagents and chemicals from other manufacturers, contact the manufacturer.

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