Amplex[®] Red/UltraRed Stop Reagent

Introduction

Amplex® UltraRed and Amplex® Red assay kits provide sensitive biomolecular assays based on hydrogen peroxide-generating enzyme systems linked to peroxidase-mediated oxidation of the fluorogenic Amplex® UltraRed or Amplex® Red substrates.1,2,3 Typically, detection reactions are performed in microplate wells and are initiated by adding the fluorogenic Amplex® UltraRed or Amplex® Red substrate, resulting in a continuous fluorescence increase that proceeds for 30 minutes or more. Ultimately, unknown analyte concentrations are determined by referencing fluorescence intensities measured at a certain time point during the reaction to parallel measurements at the same time point on standard samples of known concentration. Clearly, it is quite critical to ensure that the timing of the standard and unknown sample measurements is the same. The Amplex® Red/UltraRed stop reagent provides convenience and control by allowing the fluorescence signal-generating reaction to be terminated at a user-determined time point (Figure 1). After addition of the stop reagent, the fluorescence signal remains stable for at least 3 hours. The Amplex® Red/UltraRed stop reagent is designed for use in conjunction with both the Amplex® Red and Amplex® UltraRed fluorogenic substrates and assay kits (Figure 2). It is designed to terminate reactions containing up to 0.1 units/mL of horseradish peroxidase (HRP) and 5 μ M H₂O₂. In principle, the Amplex[®] Red/UltraRed stop reagent should also be effective in other HRP-coupled nonradioactive detection systems, although we have not yet validated its performance in such systems.

Materials

• Amplex[®] Red/UltraRed stop reagent, 5 vials. Each vial contains sufficient material to terminate 100 detection reactions as defined in the protocol described below.

Materials required but not provided

- Ethanol and water for reconstitution
- Vial or pipettor reservoir for dilution of stop solution (note A).

Storage and Handling

Store refrigerated at $\leq 6^{\circ}$ C until required for use. If frozen, avoid freeze-thaw cycles. Desiccation is recommended but not essential.



Figure 1. Application of the Amplex[®] Red/UltraRed stop reagent to control H₂O₂/ peroxidase-coupled detection reactions. Two parallel reactions containing 0.5 mU/mL horseradish peroxidase in 50 mM sodium phosphate buffer, pH 7.4 were initiated by addition of 50 μ M Amplex[®] Red reagent + 1 mM H₂O₂. Reaction progress was monitored by detection of the fluorescent product resorufin at 37°C in a fluorescence microplate reader using excitation at 530 ± 12.5 nm and fluorescence detection at 590 ± 17.5 nm. After five minutes (\cdot), one of the reactions (\Box) was terminated by addition of Amplex[®] Red/UltraRed stop reagent. The fluorescence signal in the stopped reaction remained at the constant level shown for 3 hours (data not shown).

Protocol

1.1. Reconstitute one vial of Amplex[®] Red/UltraRed stop reagent by adding 1.45 mL of ethanol. Vortex or agitate briefly. Transfer 1.25 mL of this solution to user-supplied vial or pipettor reservoir (note A) and dilute with an equal volume (1.25 mL) of water, giving a total of 2.5 mL of stop solution. This amount is sufficient to stop 100 detection reactions of 100 μ L each (note B). After reconstitution, the stop reagent is stable for approximately one month stored at 2–6°C in the dark. The stop solution should be colorless. Appearance of amber coloration is indicative of decomposition.

1.2. At the desired stopping time point in the Amplex[®] Red or Amplex[®] UltraRed microplate assay, add 20 μ L of Amplex[®] Red/UltraRed stop reagent solution per 100 μ L sample in each microplate well (note B). The stop reagent should be added to all wells, including any reagent controls containing Amplex[®] Red or Amplex[®] UltraRed reagents but no HRP (note C). The time-dependent fluorescence signal increase will cease immediately and the fluorescence signal level should remain stable for at least 3 hours.



Figure 2. Application of the Amplex® Red/UltraRed stop reagent in the standard Amplex® Red Glucose/Glucose Oxidase Assay protocol. Reactions containing 50 μ M Amplex® Red reagent, 0.1 U/mL HRP, 1 U/mL glucose oxidase and the indicated amounts of glucose in 50 mM sodium phosphate buffer, pH 7.4, were analyzed in a fluorescence microplate reader (excitation at 530 ± 12.5 nm, fluorescence detection at 590 ± 17.5 nm) after 30 minutes incubation at room temperature (\blacksquare). A parallel set of reactions (\Box) contained 20 μ L of Amplex® Red/UltraRed stop reagent per 100 μ L sample added at the initiation of the reactions. The data demonstrate the complete inhibition of signal development by the stop reagent. Note that in coupled assay applications, the stop reagent would usually be added after allowing some time for signal development (user-defined, typically 10–60 minutes), not at the initiation point as in this example.

Notes

[A] Capacity of ≥ 3 mL is required.

[B] This protocol is based on a reaction volume of 100 μ L, as typically used in 96-well microplates. For other reaction volumes, adjust the addition of Amplex[®] Red/UltraRed stop reagent solution proportionally (e.g. add 8 μ L to a 40 μ L reaction volume).

[C] Addition to controls is required to take account of the ~17% dilution of the samples upon addition of the stop solution and also to compensate for the fluorescence quenching effect (typically <5%) of the Amplex[®] Red/UltraRed stop reagent stop reagent on the Amplex[®] Red and Amplex[®] UltraRed reagent oxidation products.

References

1. Anal Biochem 253, 162-168 (1997); 2. Anal Biochem 305, 118-119 (2002); 3. J Biochem Biophys Methods 38, 43-52 (1999).

Product List Current prices may be obtained from our website or from our Customer Service Department.		
Cat #	Product Name	Unit Size
A33855	Amplex® Red/UltraRed stop reagent *500 tests*	1 set

Contact Information

Further information on Molecular Probes products, including product bibliographies, is available from your local distributor or directly from Molecular Probes. Customers in Europe, Africa and the Middle East should contact our office in Paisley, United Kingdom. All others should contact our Technical Service Department in Eugene, Oregon.

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