SiteClick™ Antibody Azido Modification Kit *5 mg labeling*

Catalog No. S10901

Pub. No. MAN0018873 **Rev.** A.0

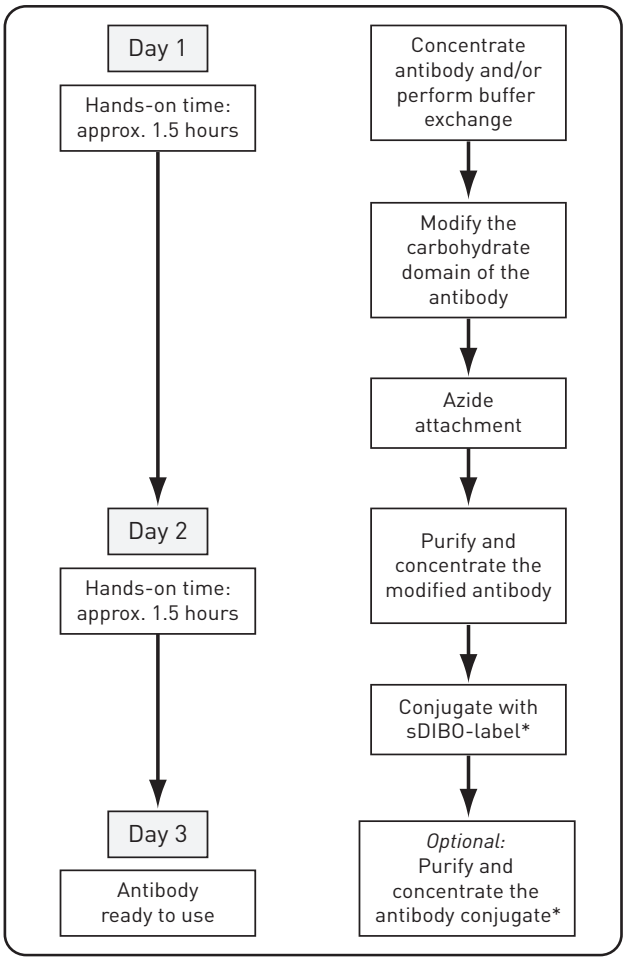
Product information

The SiteClick™ Antibody Azido Modification Kit allows you to specifically attach an azide moiety to the heavy chains of an unlabeled IgG antibody, ensuring that the antigen binding domains of the antibody remain unaltered for binding to your antigen target. The azide-modified antibody can then be covalently linked to SiteClick™ sDIBO Alkyne labels (available separately; see Table 2, page 3) in a copper-free click reaction without reducing the protein. This gives you the option to choose different fluorescent labels for your antibody, attach another molecule via streptavidin, or attach your own molecule via amine-reactive or amine-containing moieties depending on your assay.

Each SiteClick™ Antibody Azido Modification Kit contains sufficient reagents to perform one azido modification reaction starting with 5 mg of whole IgG produced in eukaryotic cells from any host species. The antibody concentrators provided in the kit are used to purify and concentrate the antibody at each step of the SiteClick™ antibody labeling workflow (Figure 1, page 2)

Table 1 Contents and storage

Material	Amount	Storage*
Antibody preparation buffer (Component A)	6 mL	• 2–8°C • DO NOT FREEZE
Antibody concentrator (small, 2 mL) (Component B)	each	
ß-Galactosidase (Component C)	110 µL	
UDP-GalNAz (Component D)	2.2 mg	
20X Tris pH 7.0 (1M) (Component E)	6 mL	
Buffer additive (Component F)	300 µL	
ß-1,4-galactosyltransferase (GalT) (Component G)	880 µL	
Antibody concentrator (large, 4 mL) (Component H)	each	
* When stored as directed, this kit is stable for at least 6 months.		



* Requires the use of the SiteClick™ sDIBO Alkyne for SiteClick™ Antibody Labeling kits (available separately).

Figure 1 SiteClick™ antibody azido modification and antibody labeling workflow. The SiteClick™ Antibody Azido Modification Kit is designed to be used with the SiteClick™ sDIBO Alkyne for SiteClick™ Antibody Labeling kits (available separately; see Table 2) for a complete antibody labeling workflow.

Table 2 SiteClick™ sDIBO Alkynes for SiteClick™ Antibody Labeling. The SiteClick™ sDIBO Alkyne labels (available separately) are used in conjunction with the SiteClick™ Antibody Azido Modification Kits (sufficient for 5 mg azide-modified antibody) or with engineered antibodies containing azido moieties to create high-quality antibody conjugates.

	Catalog No.[1]		
Product	**100 µg kit**	**1 mg kit**	**5 mg kit**
SiteClick™ Biotin sDIBO Alkyne	C20030	S10902	S10907
SiteClick™ pHrodo™ iFL Red sDIBO Alkyne	C20034	S10903	S10908
SiteClick™ Alexa Fluor™ 488 sDIBO Alkyne	C20027	S10904	S10909
SiteClick™ Alexa Fluor™ 555 sDIBO Alkyne	C20028	—	—
SiteClick™ Alexa Fluor™ 647 sDIBO Alkyne	C20029	S10906	S10911

[1] See Table 3 (page 9) for the amount of SiteClick™ sDIBO Alkyne label required to label 100 µg, 1 mg, and 5 mg azide-modified antibody with the SiteClick™ sDIBO Alkyne Kits available from Thermo Fisher Scientific.

Before you begin

Equipment required

- Centrifuge with swinging bucket rotor that can accommodate 17 mm × 100 mm centrifuge tubes

Required Materials not supplied

- 5 mg of whole IgG antibody produced in eukaryotic cells, preferably at a concentration of 2.5–20 mg/mL in a Tris-based buffer, free of carrier proteins and/or azide
- Centrifuge tubes: 1.5-mL, 15-mL, and 50-mL
- Distilled water (dH_2O)
- PBS or TBS
- SiteClick™ sDIBO Alkyne label (sDIBO-dye, sDIBO-biotin, or sDIBO-chelator) (Table 2, page 2).

Caution

- **IMPORTANT!** Avoid sodium azide throughout the protocol.
- β-Galactosidase (Component C) may cause an allergic skin reaction, and it may cause allergy or asthma symptoms or breathing difficulties, if inhaled. Read the Safety Data Sheet (SDS), available at **thermofisher.com**, before handling this reagent.
- Dispose of the reagents in compliance with all pertaining local regulations. In case of contact with eyes, rinse immediately with plenty of water and seek medical advice. Always wear suitable laboratory protective clothing and gloves when handling these reagents.

Step 1. Concentrate antibody and/or perform buffer exchange

Time required: 1 hour

This antibody concentration and buffer exchange step is required if:

- Your antibody concentration is less than 10 mg/mL, and/or
- Your antibody is in a phosphate-based buffer (e.g. PBS), and/or
- Your antibody is in a buffer containing azide.

Before you begin, briefly centrifuge the tubes containing enzymes, substrates, or dyes to ensure all material is at the bottom of the tubes.

Wash the antibody concentrator

1.1 Remove the conical collection tube from the large antibody concentrator (Component H).

1.2 Add 2 mL of dH_2O to the small antibody concentrator (Component B) and cap the device as shown in Figure 2 A (page 4).

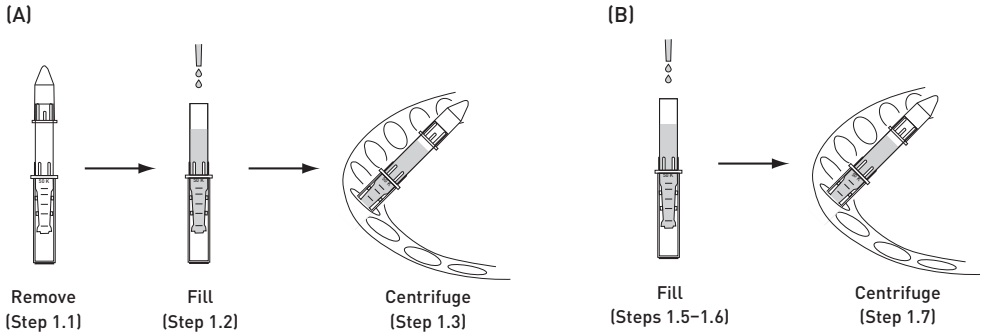


Figure 2 Antibody concentration and/or buffer exchange. (A) Wash the antibody concentrator; (B) Concentrate the antibody.

1.3 Ensure that one membrane panel of the concentrator faces the center of the rotor, then centrifuge at 1400 × *g* for 10 minutes.

IMPORTANT! To avoid damage to the antibody concentrator during centrifugation, ensure that it is properly assembled and seated at the bottom of the rotor. The rim of the concentrate collection tube should be inside the rotor well. Check clearance before centrifugation.

1.4 Discard the flow-through.

Note: After washing the antibody concentrator, do not let the membrane dry out.

Concentrate antibody and/or perform buffer exchange

1.5 Add a sufficient volume of antibody solution to contain 5 mg of antibody to the small antibody concentrator (Component B) as shown in Figure 2 B.

Note: If the antibody concentration is < 2.5 mg/mL, repeated additions to the concentrator are necessary.

1.6 Bring the total volume in the concentrator to 2 mL using the antibody preparation buffer (Component A).

1.7 Ensure that one membrane panel of the concentrator faces the center of the rotor, then centrifuge at 2000 × *g* for 12 minutes. Discard the flow-through.

1.8 Add antibody preparation buffer (Component A) to the small antibody concentrator (Component B) so that the total volume in the concentrator is 2 mL.

1.9 Ensure that one membrane panel of the concentrator faces the center of the rotor, then centrifuge at 2000 × *g* for 12 minutes. Discard the flow-through.

1.10 Repeat Steps 1.8 and 1.9.

Note: If the antibody volume in the concentrator is greater than 500 µL following Step 1.10, centrifuge at 2000 × *g* for an additional 3 minutes or until the appropriate volume is achieved.

1.11 Invert the small antibody concentrator (Component B) into the collection tube as shown in Figure 3.

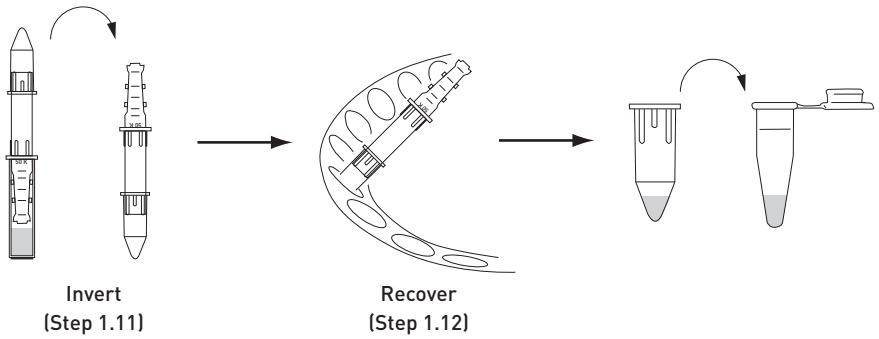


Figure 3 Collect concentrated antibody in preparation buffer.

1.12 Centrifuge for 3 minutes at 1000 × *g* to collect the concentrated antibody. Following collection, you should have approximately 500 µL of concentrated antibody in the collection tube.

1.13 Transfer the antibody from the conical collection tube to a 1.5-mL centrifuge tube, then bring volume to 500 µL with antibody preparation buffer (Component A).

Step 2. Modify the carbohydrate domain of the antibody

Time required: 5 minutes hands-on, then 6 hours to overnight incubation

Add ß-galactosidase

2.1 Add 100 µL of β-galactosidase (Component C) to the antibody collected in Step 1.13, as shown in Figure 4.

2.2 Wrap the tube cap with Parafilm™ laboratory film or similar, then incubate for 6 hours to overnight at 37°C.

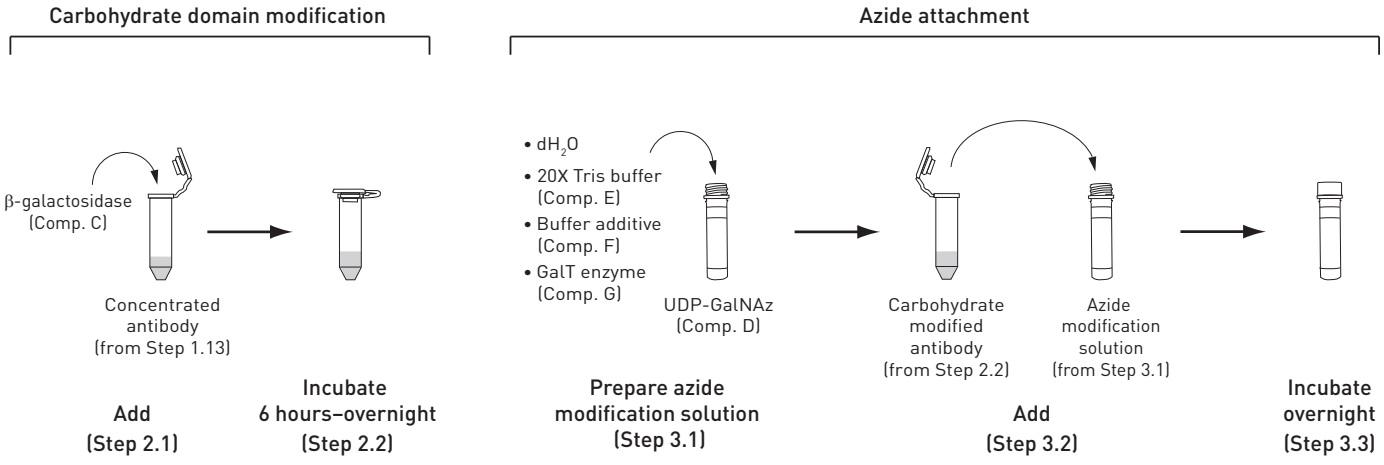


Figure 4 Modification of antibody carbohydrate domain and azide attachment

Step 3. Azide attachment

Time required: 5 minutes hands-on, then overnight incubation

Add GalT enzyme

3.1 Prepare the azide modification solution by adding the following components to the tube containing UDP-GalNAz (Component D), as shown in Figure 4:

- 60 µL of dH_2O
- 90 µL of 20X Tris buffer, pH 7.0 (Component E)
- 200 µL of buffer additive (Component F)
- 800 µL of GalT enzyme (Component G)

3.2 Vortex the reaction components, then add the modified antibody from Step 2.2 (600 µL) to the tube.

3.3 Briefly centrifuge the tube, wrap the tube cap with Parafilm™ laboratory film or similar, then incubate overnight at 30°C.

Step 4. Purify and concentrate the azide-modified antibody

Time required: 1 hour

- This step removes any excess substrate UDP-GalNAz.
- You can also use TBS or other phosphate-free buffers for the purification and collection of the modified antibody (Steps 4.2–4.12). 20X Tris, pH 7.0 is provided for your convenience.

4.1 Prepare 40 mL of 1X Tris, pH 7.0 by adding 2 mL of 20X Tris, pH 7.0 (Component E) to 38 mL of dH_2O in a 50-mL conical tube. Vortex briefly to mix.

Wash the antibody concentrator

4.2 Remove the cap from the large antibody concentrator (Component H).

4.3 Add 4 mL of 1X Tris, pH 7.0 (or TBS) to the large antibody concentrator (Component H) and replace the cap as shown in Figure 5.

4.4 Ensure that one membrane panel of the concentrator faces the center of the rotor, then centrifuge at 2000 × *g* for 5 minutes. Discard the flow-through.

Note: After washing the antibody concentrator, do not let the membrane dry out.

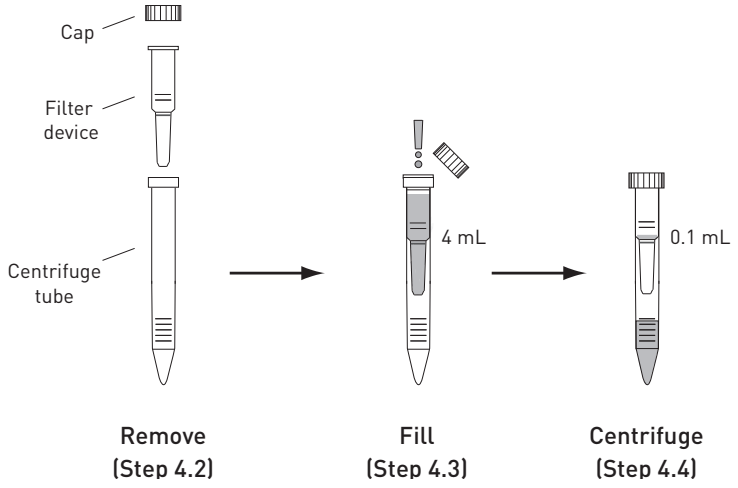


Figure 5 Wash the antibody concentrator

Purify the antibody

4.5 Add 2 mL of 1X Tris pH 7.0 (or TBS) and 1.75 mL of the azide-modified antibody from Step 3.3 to the filter device of large antibody concentrator (Component H) as shown in Figure 6.

4.6 Ensure that one membrane panel of the concentrator faces the center of the rotor, then centrifuge at 2000 × *g* for 8 minutes. Discard the flow-through.

4.7 Add 1X Tris pH 7.0 (or TBS) to a total volume of 4 mL to the large antibody concentrator (Component H).

4.8 Ensure that one membrane panel of the concentrator faces the center of the rotor, then centrifuge at 2000 × *g* for 12 minutes. Discard the flow-through.

4.9 Repeat Steps 4.7 and 4.8 two more times.

Note: If the antibody volume in the concentrator is greater than ~1 mL or if an antibody concentration of more than ~5.0 mg/mL is desired, you can reduce the volume in the concentrator by additional centrifugation (e.g., at 2000 × *g* for an additional 3 minutes or until the appropriate volume is achieved).

Note: If you intend to conjugate the azide-modified antibody to macromolecule-sDIBO, measure the OD_{260} in the flow-through (with ε_{260} = 9900 M^{-1} cm^{-1}) following the removal of UDP-GalNAz. In this case, you might need to perform Steps 4.7 and 4.8 again to remove free UDP-GalNAz sufficiently.

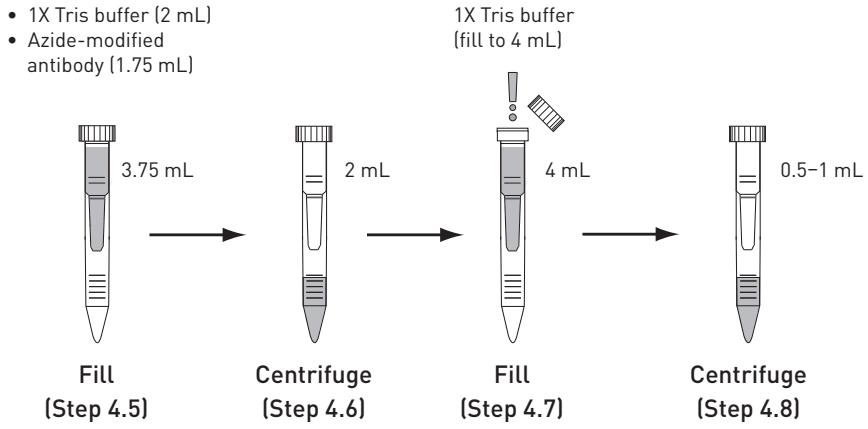


Figure 6 Purify and concentrate azide-modifed antibody

Collect the azide-modified antibody

4.10 Using a 200-µL pipettor, collect the concentrated antibody and transfer to a 1.5-mL centrifuge tube or another tube of choice.

4.11 Measure the OD_{280} (with OD_{280} at 1.4 = 1 mg/mL) to determine the antibody concentration. Expected concentration is ~4–5 mg/mL.

Store the azide-modified antibody

At this point, you can store the azide-modified antibody at 2–8°C until needed. Do not freeze the azide-modified antibody.

IMPORTANT! If you wish to perform a click reaction to conjugate your azide-modified antibody to a SiteClick™ sDIBO Alkyne label, do not add sodium azide to your modified antibody. Sodium azide must be avoided throughout the protocol.

Step 5. Attach sDIBO-modified label to azide-modified antibody

Time required: 5 minutes hands-on, then overnight incubation

This section provides instructions to covalently link the azide-modified antibody to a SiteClick™ sDIBO Alkyne label in a copper-free click reaction.

See Table 3 (page 9) for the amount of SiteClick™ sDIBO Alkyne label required to label 100 µg, 1 mg, and 5 mg azide-modified antibody with the SiteClick™ sDIBO Alkyne Kits available from Thermo Fisher Scientific.

Materials required but not provided

- Azide-modified antibody (from Step 4.11) in a Tris-based buffer, free of carrier proteins and/or azide
- SiteClick™ sDIBO Alkyne label (available separately; see Table 3, page 9)
- Anhydrous DMSO (only required for dissolving SiteClick™ pHrodo™ iFL Red sDIBO Alkyne; included in Cat. No. S10908)
- Distilled water (dH_2O)
- PBS or TBS
- 15-mL centrifuge tubes

Caution

- IMPORTANT! Sodium azide must be avoided throughout the protocol.
- DMSO is known to facilitate the entry of organic molecules into tissues. Handle reagents containing DMSO using equipment and practices appropriate for the hazards posed by such materials.
- Read the Safety Data Sheet (SDS), available at **thermofisher.com**, before handling the reagents.
- Dispose of the reagents in compliance with all pertaining local regulations. In case of contact with eyes, rinse immediately with plenty of water and seek medical advice. Always wear suitable laboratory protective clothing and gloves when handling these reagents.

Table 3 SiteClick™ sDIBO Alkynes for SiteClick™ Antibody Labeling. The SiteClick™ sDIBO Alkyne labels (available separately) are used in conjunction with the SiteClick™ Antibody Azido Modification Kits or with engineered antibodies containing azido moieties to create high-quality antibody conjugates.

	Catalog No.[4]			100 µg antibody		1 mg antibody		5 mg antibody	
Product	100 µg kit	1 mg kit	5 mg kit	in TBS[5]	sDIBO[6]	in TBS[5]	sDIBO[6]	in TBS[5]	sDIBO[6]
SiteClick™ Biotin sDIBO Alkyne	C20030	—	—	90 µL	10 µL	—	—	—	—
SiteClick™ pHrodo™ iFL Red sDIBO Alkyne[1]	C20034	—	—	90 µL	10 µL	—	—	—	—
SiteClick™ Alexa Fluor™ 488 sDIBO Alkyne	C20027	—	—	90 µL	10 µL	—	—	—	—
SiteClick™ Alexa Fluor™ 555 sDIBO Alkyne	C20028	—	—	90 µL	10 µL	—	—	—	—
SiteClick™ Alexa Fluor™ 647 sDIBO Alkyne	C20029	—	—	90 µL	10 µL	—	—	—	—
SiteClick™ Biotin sDIBO Alkyne	—	S10902	—	45 µL	5 µL	450 µL	50 µL	—	—
SiteClick™ pHrodo™ iFL Red sDIBO Alkyne[2]	—	S10903	—	45 µL	5 µL	450 µL	50 µL	—	—
SiteClick™ Alexa Fluor™ 488 sDIBO Alkyne	—	S10904	—	45 µL	5 µL	450 µL	50 µL	—	—
SiteClick™ Alexa Fluor™ 647 sDIBO Alkyne	—	S10906	—	45 µL	5 µL	450 µL	50 µL	—	—
SiteClick™ Biotin sDIBO Alkyne	—	—	S10907	45 µL	5 µL	450 µL	50 µL	2.25 mL	250 µL
SiteClick™ pHrodo™ iFL Red sDIBO Alkyne[3]	—	—	S10908	45 µL	5 µL	450 µL	50 µL	2.25 mL	250 µL
SiteClick™ Alexa Fluor™ 488 sDIBO Alkyne	—	—	S10909	45 µL	5 µL	450 µL	50 µL	2.25 mL	250 µL
SiteClick™ Alexa Fluor™ 647 sDIBO Alkyne	—	—	S10911	45 µL	5 µL	450 µL	50 µL	2.25 mL	250 µL

[1–3] Dissolve SiteClick™ pHrodo™ iFL Red sDIBO Alkyne in 25 µL[1], 50 µL[2], and 250 µL[3] DMSO, respectively.
[4] 100 µg kits, 1 mg kits, and 5 mg kits contain sDIBO in 25 µL, 50 µL, and 250 µL DMSO, respectively, enough to label 2.5 × 100 µg, 10 × 100 µg, and 50 × 100 µg azide tagged antibody.
[5] Antibody can be in 50 mM Tris (pH 7.0), TBS, PBS, or other thiol-free and sodium azide-free buffer.
[6] sDIBO-derivatives are dissolved in DMSO.

Add SiteClick™ sDIBO Alkyne to azide-modified antibody

5.1 Add 250 µL of the SiteClick™ sDIBO Alkyne label to the 5 mg azide-modified antibody in 2.25 mL of 1X Tris pH 7.0 (or TBS) in the 15-mL centrifuge tube.

Note: The SiteClick™ pHrodo™ iFL Red sDIBO Alkyne for SiteClick™ Antibody Labeling (Cat. No. S10908) is supplied lyophilized as a solid powder. Before use, dissolve the SiteClick™ pHrodo™ iFL Red sDIBO Alkyne in 250 µL of anhydrous DMSO, which is included in the kit.

Other SiteClick™ sDIBO Alkynes for SiteClick™ Antibody Labeling are supplied as 250-µL solutions in DMSO and do not need to be dissolved.

5.2 Vortex the reaction mixture, briefly centrifuge, and incubate overnight at 25°C.

Note: Following incubation, you can store the antibody conjugate at 2–8°C until needed (see "Store the antibody conjugate", page 11) or purify it of the excess SiteClick™ sDIBO Alkyne label (Step 6, optional).

Step 6. Purify and concentrate the antibody conjugate (optional)

Time required: 2 hours

- The purification step removes any excess SiteClick™ sDIBO alkyne label that has not been conjugated with antibody. This removal can be achieved by size exclusion chromatography or centrifugal filtration. For your convenience, centrifugal filters have been included in kits S10907 to S10911.
- You may use TBS or PBS for the purification and collection of the modified antibody (Steps 6.2–6.7)

Materials required but not provided

- Antibody conjugate (from Step 5.2)
- Antibody concentrator, large (included in the 5 mg SiteClick™ sDIBO Alkyne for SiteClick™ Antibody Labeling kits)

  Note: The antibody concentrator included in the 5 mg SiteClick™ sDIBO Alkyne for SiteClick™ Antibody Labeling kits (Component B; Component C in Cat. No. S10908) is identical to the large antibody concentrator (Component H) supplied with the 5 mg SiteClick™ Antibody Azido Modification Kit.
- Distilled water (dH_2O)
- PBS or TBS
- 15-mL centrifuge tubes

Wash the antibody concentrator

6.1 Remove the cap from the large antibody concentrator.

6.2 Add 4 mL of 1X Tris, pH 7.0 (or TBS) to the large antibody concentrator and replace the cap as shown in Figure 7.

6.3 Ensure that one membrane panel of the concentrator faces the center of the rotor, then centrifuge at 2000 × *g* for 5 minutes. Discard the flow-through.

Note: After washing the antibody concentrator, do not let the membrane dry out.

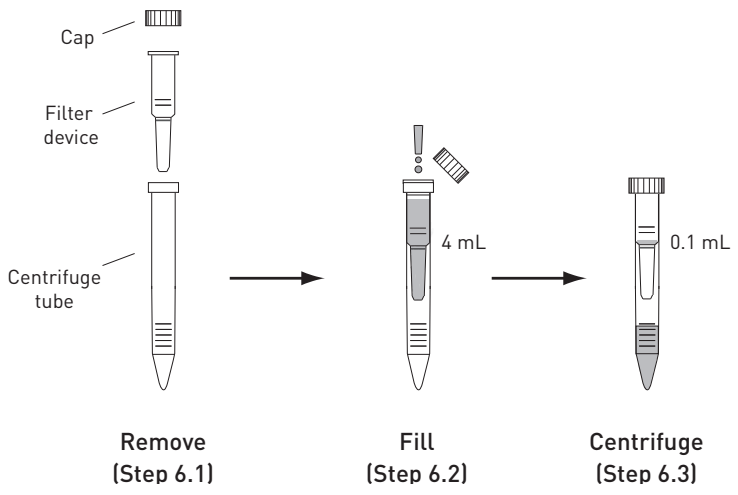


Figure 7 Wash the antibody concentrator

Purify the antibody conjugate

6.4 Add 1.0 mL of 1X Tris, TBS, or PBS and 2.5 mL of the sDIBO-modified antibody (from Step 5.2) to the large antibody concentrator as shown in Figure 8.

6.5 Ensure that one membrane panel of the concentrator faces the center of the rotor, then centrifuge at 2000 × *g* for 12 minutes. Discard the flow-through.

6.6 Add 1X Tris, TBS, or PBS to a total volume of 4 mL to the large antibody concentrator. Ensure that one membrane panel of the concentrator faces the center of the rotor, then centrifuge at 2000 × *g* for 20 minutes. Discard the flow-through.

6.7 Repeat Step 6.6 at least three more times.

Note: If an antibody concentration of more than ~5 mg/mL is desired, you can reduce the volume in the concentrator by additional centrifugation (e.g., at 2000 × *g* for an additional 3 minutes or until the appropriate volume is achieved).

Collect the purified antibody conjugate

6.8 Using a 200-µL pipettor, collect the concentrated antibody and transfer to a 1.5-mL centrifuge tube or another tube of choice.

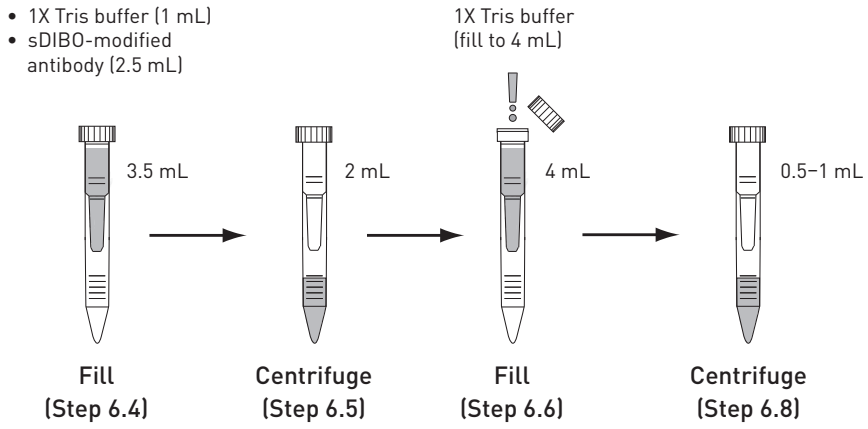


Figure 8 Collect the purified and concentrated antibody conjugate

Store the antibody conjugate

Store the antibody conjugate at 2–8°C until needed. DO NOT FREEZE.

You can add sodium azide or thimerosal at this stage to a final concentration of 0.02% (w/v) for long term storage, if preferred.

Step 7. Determine the Degree of Labeling (DOL) of sDIBO alkyne-labeled antibody (optional)

7.1 Determine the DOL from the A_{dye}/A_{280} ratio. Use Correction Factor (CF_{280}) of the label at A_{280} to calculate (see Table 4).

$$(Moles/L)_{dye} = A_{dye}/\varepsilon_{dye}$$

$$(Moles/L)_{IgG} = [A_{280} - (CF_{280} \times A_{dye})]/203{,}000$$

$$DOL = (Moles)_{dye}/(Moles)_{IgG}$$

Table 4 Molecular weight (MW), emission maxima (λ_{max}), molar extinction coefficient (ε_{dye}), and Correction Factor (CF_{280}) for the SiteClick™ sDIBO Alkynes for SiteClick™ Antibody Labeling.

Product	~MW	λ_{max}	ε_{dye} [1]	CF_{280} [2]
SiteClick™ pHrodo™ iFL Red sDIBO Alkyne	~1800	560 nm	65,000	0.221
SiteClick™ Alexa Fluor™ 488 sDIBO Alkyne	~1450	495 nm	73,000	0.134
SiteClick™ Alexa Fluor™ 555 sDIBO Alkyne	~1850	555 nm	145,000	0.091
SiteClick™ Alexa Fluor™ 647 sDIBO Alkyne	~1900	655 nm	234,000	0.037

[1] Extinction coefficient at λ_{max} in $M^{-1}cm^{-1}$.
[2] Correction factor for absorption readings (A_{280}) at 280 nm; e.g. $A_{280,actual} = A_{280,observed} - (CF_{280} \times A_{dye})$.

Example calculation with SiteClick™ Alexa Fluor™ 488 sDIBO Alkyne

Determine the DOS from the A_{495}/A_{280} ratio, using $CF_{280} = 0.134$ for Alexa Fluor™ 488 at A_{280} (see Table 4, page 12) for the calculation:

$$(Moles/L)_{dye} = A_{495}/73{,}000$$

$$(Moles/L)_{IgG} = [A_{280} - (0.134 \times A_{495})]/203{,}000$$

$$DOL = (Moles)_{dye}/(Moles)_{IgG}$$

Appendix: Assay principle

In the first step of SiteClick™ conjugation, terminal galactose residues on the N-linked sugars in the Fc region of the antibody are removed by β-Galactosidase. The azide-containing sugar, GalNAz, is then added to the modified carbohydrate domain of the antibody via the β-1,4-galactosyltransferase (Gal-T)-catalyzed reaction targeting the terminal GlcNAc residues. This specific targeting maintains the integrity of the antigen binding site on the antibody. Finally, the antibody (now containing an azide moiety) is conjugated to the sDIBO-modified label in a copper-free click reaction with simple overnight incubation (Figure 9).

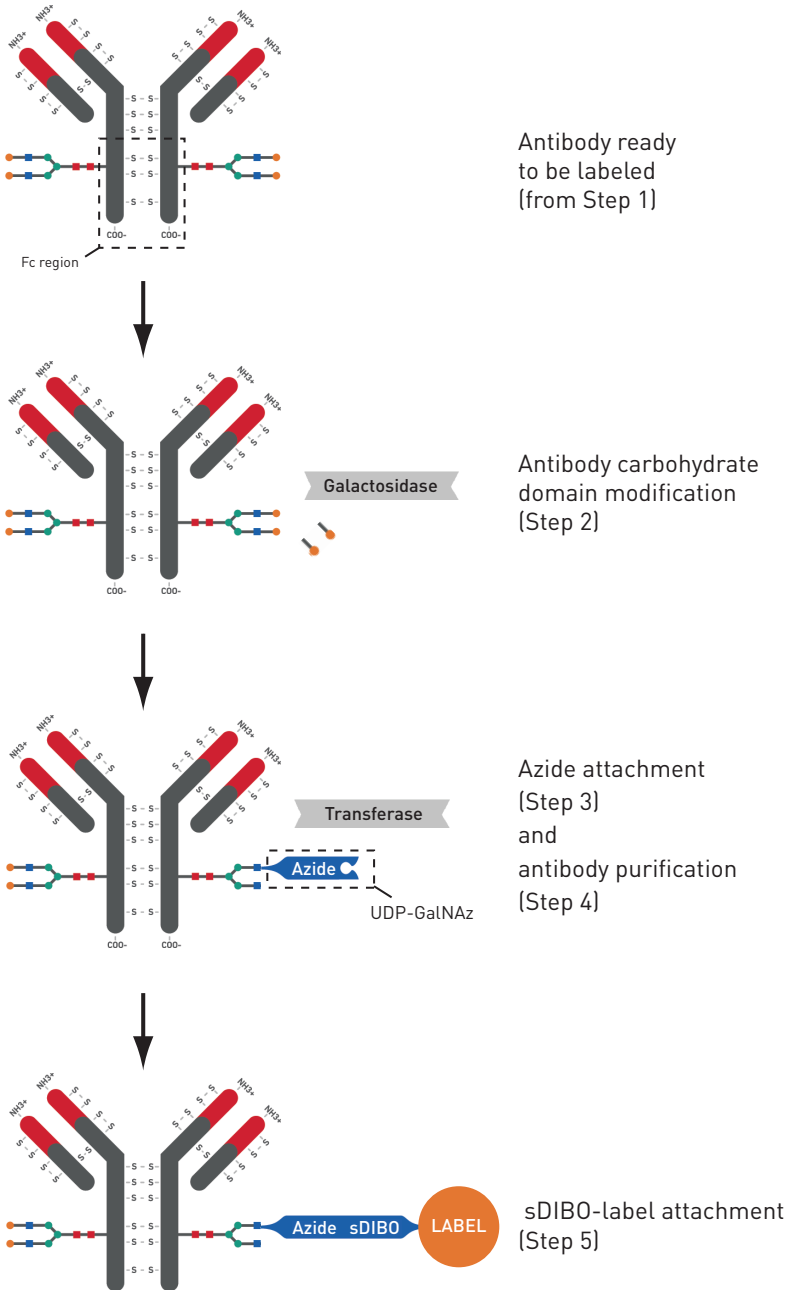


Figure 9 SiteClick™ conjugation reaction

Ordering information

Cat. No.	Product	Unit size
S10901	SiteClick™ Antibody Azido Modification Kit *5 mg labeling*	1 kit

Related products

5 mg SiteClick™ sDIBO labels for azido-modified antibodies:

S10907	SiteClick™ Biotin sDIBO Alkyne for SiteClick™ Antibody Labeling	1 kit
S10908	SiteClick™ pHrodo™ iFL Red sDIBO Alkyne for SiteClick™ Antibody Labeling	1 kit
S10909	SiteClick™ Alexa Fluor™ 488 sDIBO Alkyne for SiteClick™ Antibody Labeling	1 kit
S10911	SiteClick™ Alexa Fluor™ 647 sDIBO Alkyne for SiteClick™ Antibody Labeling	1 kit

1 mg SiteClick™ sDIBO labels for azido-modified antibodies:

S10902	SiteClick™ Biotin sDIBO Alkyne for SiteClick™ Antibody Labeling	1 kit
S10903	SiteClick™ pHrodo™ iFL Red sDIBO Alkyne for SiteClick™ Antibody Labeling	1 kit
S10904	SiteClick™ Alexa Fluor™ 488 sDIBO Alkyne for SiteClick™ Antibody Labeling	1 kit
S10906	SiteClick™ Alexa Fluor™ 647 sDIBO Alkyne for SiteClick™ Antibody Labeling	1 kit

100 µg SiteClick™ sDIBO labels for azido-modified antibodies:

C20027	SiteClick™ Alexa Fluor™ 488 sDIBO Alkyne for SiteClick™ Antibody Labeling	1 kit
C20028	SiteClick™ Alexa Fluor™ 555 sDIBO Alkyne for SiteClick™ Antibody Labeling	1 kit
C20029	SiteClick™ Alexa Fluor™ 647 sDIBO Alkyne for SiteClick™ Antibody Labeling	1 kit
C20030	SiteClick™ Biotin sDIBO Alkyne for SiteClick™ Antibody Labeling	1 kit
C20031	SiteClick™ Amine sDIBO Alkyne for SiteClick™ Antibody Labeling	1 kit
C20032	SiteClick™ SDP Ester sDIBO Alkyne for SiteClick™ Antibody Labeling	1 kit
C20034	SiteClick™ pHrodo™ iFL Red sDIBO Alkyne for SiteClick™ Antibody Labeling	1 kit
S20033	SiteClick™ Biotin Antibody Labeling Kit	1 kit
S20026	SiteClick™ Antibody Azido Modification Kit	1 kit

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  Note: For SDSs for reagents and chemicals from other manufacturers, contact the manufacturer.

Limited Product Warranty

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Revision history: Pub. No. MAN0018873

Revision	Date	Description
A.0	11 September 2019	New User Guide

Manufacturer: Life Technologies Corporation | 29851 Willow Creek Road | Eugene, OR 97402