

# Human EGFR (Full length) ELISA Kit

Catalog Number KHR9061 (96 tests)

Pub. No. MAN0014927 Rev. 2.0 (30)

**CAUTION!** This kit contains materials with small quantities of sodium azide. Sodium azide reacts with lead and copper plumbing to form explosive metal azides. Upon disposal, flush drains with a large volume of water to prevent azide accumulation. Avoid ingestion and contact with eyes, skin and mucous membranes. In case of contact, rinse affected area with plenty of water. Observe all federal, state, and local regulations for disposal.

**Note:** For safety and biohazard guidelines, see the “Safety” appendix in the *ELISA Technical Guide* (Pub. no. MAN0006706). Read the Safety Data Sheets (SDSs) and follow the handling instructions. Wear appropriate protective eyewear, clothing, and gloves.

## Product description

The Invitrogen™ Human EGFR (Full length) ELISA Kit is a solid-phase sandwich Enzyme-Linked Immunosorbent Assay (ELISA) designed to detect and quantify the level of full length EGFR (non-truncated) in fresh or frozen human cell lysates. The assay recognizes both natural and recombinant full length human EGFR, independent of its phosphorylation state.

The epidermal growth factor receptor (EGFR) belongs to the family of receptor tyrosine kinases (RTKs), which regulate cell growth, survival, proliferation and differentiation. Full length EGFR is a 170 kDa type I transmembrane glycoprotein consisting of an extracellular ligand-binding domain, a single hydrophobic transmembrane region, and an intracellular segment with a highly conserved, tyrosine kinase domain. Alternative EGFR transcripts encode truncated 60 kDa and 110 kDa isoforms. This kit can be used to normalize the phosphorylated full length EGFR content of the samples when using the Human EGFR [pY1173] ELISA Kit (Cat. No. KHR9071) or the Human EGFR [pY1068] ELISA Kit (Cat. No. KHR9081).

## Contents and storage

Upon receipt, store the kit at 2°C to 8°C.

Contents	Cat. No. KHR9061 (96 tests)
Hu EGFR Standard, lyophilized; contains 0.1% sodium azide. Refer to vial label for quantity and reconstitution volume.	2 vials
Standard Diluent Buffer <sup>[1]</sup> ; contains 0.1% sodium azide	25 mL
Antibody Coated Plate, 96-well strip-well plate	1 plate
Hu EGFR Detection Antibody; contains 0.1% sodium azide	11 mL
Anti-Rabbit IgG HRP (100X); contains 3.3 mM thymol	0.125 mL
HRP Diluent; contains 3.3 mM thymol	25 mL
Wash Buffer Concentrate (25X)	100 mL
Stabilized Chromogen, Tetramethylbenzidine (TMB)	25 mL
Stop Solution	25 mL
Plate Covers, adhesive strips	3

<sup>[1]</sup> If precipitates are found in standard diluent buffer, they should be completely dissolved by warming to room temperature before use.

## Required materials not supplied

- Distilled or deionized water
- Microtiter plate reader with software capable of measurement at or near 450 nm
- Plate washer—automated or manual (squirt bottle, manifold dispenser, or equivalent)
- Calibrated adjustable precision pipettes and glass or plastic tubes for diluting solutions; beakers, flask and cylinders for preparation of reagents
- Cell Extraction Buffer (Cat. No. FNN0011, or see “Prepare Cell Extraction Buffer”)

## Before you begin

**IMPORTANT!** Reagents are lot-specific. Do not mix or interchange different reagent lots from various kit lots.

- Review the **Procedural guidelines** and **Plate washing directions** in the *ELISA Technical Guide* available at [thermofisher.com](http://thermofisher.com).
- Allow reagents to reach room temperature before use. Mix to redissolve any precipitated salts.

## Prepare 1X Wash Buffer

1. Dilute 16 mL of Wash Buffer Concentrate (25X) with 384 mL of deionized or distilled water. Label as 1X Wash Buffer.
2. Store the concentrate and 1X Wash Buffer in the refrigerator. Use the diluted buffer within 14 days.

## Prepare Cell Extraction Buffer

**Note:** See the *ELISA Technical Guide* for detailed information on preparing Cell Extraction Buffer.

1. Prepare Cell Extraction Buffer.  
Cell Extraction Buffer consists of 10 mM Tris (pH 7.4), 100 mM NaCl, 1 mM EDTA, 1 mM EGTA, 1 mM NaF, 20 mM Na<sub>4</sub>P<sub>2</sub>O<sub>7</sub>, 2 mM Na<sub>3</sub>VO<sub>4</sub>, 1% Triton™ X-100, 10% glycerol, 0.1% SDS, and 0.5% deoxycholate.
2. Immediately before use, add PMSF (0.3 M stock in DMSO) to 1 mM and 50 µL protease inhibitor cocktail (e.g., Sigma Cat. No. P-2714) for each 1 mL of Cell Extraction Buffer.

## Prepare cell lysate

1. Collect cells by centrifugation (non-adherent cells) or scraping from culture flasks (adherent cells), then wash cells twice with cold PBS.
2. Remove and discard the supernatant and collect the cell pellet. The pellet can be stored at  $-80^{\circ}\text{C}$  and lysed at a later date if desired.
3. Lyse the cell pellet in Cell Extraction Buffer for 30 minutes, on ice. Vortex at 10-minute intervals.

**Note:** The volume of Cell Extraction Buffer used depends on the number of cells in the cell pellet, and expression levels of full length human EGFR. [FOR EXAMPLE,  $10^8$  A431 cells can be extracted in 1 mL of Cell Extraction Buffer. Researchers must optimize the extraction procedures for their own applications].

4. Transfer the lysate into microcentrifuge tubes and centrifuge at 13,000 rpm for 10 minutes at  $4^{\circ}\text{C}$ .
5. Transfer the supernatant into clean microcentrifuge tubes. Samples can be stored at  $-80^{\circ}\text{C}$  (avoid multiple freeze-thaw cycles).

## Pre-dilute samples

Sample concentrations should be within the range of the standard curve. Because conditions may vary, each investigator should determine the optimal dilution for each application.

- Perform sample dilutions with Standard Diluent Buffer.
- Dilute samples prepared in Cell Extraction Buffer 1:10 or greater in Standard Diluent Buffer (e.g., 10  $\mu\text{L}$  sample into 90  $\mu\text{L}$  buffer).

For  $10^8$  A431 cells, use 0.1–1  $\mu\text{L}$  of the clarified lysate diluted to 100  $\mu\text{L}$  in Standard Diluent Buffer for each well.

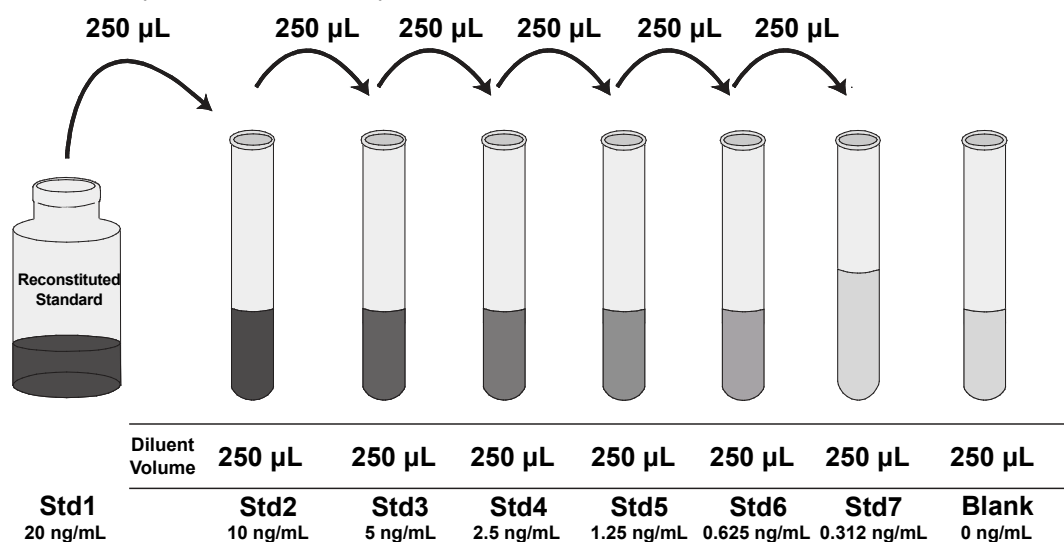
While a 1:10 sample dilution has been found to be satisfactory, higher dilutions such as 1:50 or 1:100 may be optimal.

## Dilute standards

**Note:** Use glass or plastic tubes for diluting standards.

**Note:** The Hu EGFR Standard is prepared using human EGFR (p170) from A431 cells and is calibrated against the mass of affinity purified EGFR.

1. Reconstitute Hu EGFR Standard to 20 ng/mL with Standard Diluent Buffer. Refer to the standard vial label for instructions. Swirl or mix gently and allow the contents to sit for 10 minutes to ensure complete reconstitution. Label as 20 ng/mL human EGFR. **Use the standard within 1 hour of reconstitution.**
2. Add 250  $\mu\text{L}$  Standard Diluent Buffer to each of 7 tubes labeled as follows: 10, 5, 2.5, 1.25, 0.625, and 0.312 ng/mL human EGFR.
3. Make serial dilutions of the standard as shown in the following dilution diagram. Mix thoroughly between steps.
4. Remaining reconstituted standard should be discarded or frozen in aliquots at  $-80^{\circ}\text{C}$  for further use. Standard can be frozen and thawed one time only without loss of activity.



## Prepare 1X Anti-Rabbit IgG HRP solution

**Note:** Prepare 1X Anti-Rabbit IgG HRP solution within 15 minutes of usage.

The Anti-Rabbit IgG HRP (100X) is in 50% glycerol, which is viscous. To ensure accurate dilution:






1. For each 8-well strip used in the assay, pipet 10  $\mu\text{L}$  Anti-Rabbit IgG HRP (100X) solution, wipe the pipette tip with clean absorbent paper to remove any excess solution, and dispense the solution into a tube containing 1 mL of HRP Diluent. Mix thoroughly.
2. Return the unused Anti-Rabbit IgG HRP (100X) solution to the refrigerator.

## Perform ELISA (Total assay time: 4 hours)

**IMPORTANT!** Perform a standard curve with each assay.

- Allow all components to reach room temperature before use. Mix all liquid reagents prior to use.
- Determine the number of 8-well strips required for the assay. Insert the strips in the frames for use. Re-bag any unused strips and frames, and store at 2°C to 8°C for future use.



<b>1</b>	<b>Bind antigen</b> 	<ol style="list-style-type: none"> <li>Add 100 <math>\mu</math>L of standards, controls, or samples (see "Pre-dilute samples" on page 2) to the appropriate wells. Leave the wells for chromogen blanks empty.</li> <li>Cover the plate with a plate cover and incubate 2 hours at room temperature.</li> <li>Thoroughly aspirate the solution and wash wells 4 times with 1X Wash Buffer.</li> </ol>
<b>2</b>	<b>Add detector antibody</b> 	<ol style="list-style-type: none"> <li>Add 100 <math>\mu</math>L of Hu EGFR Detection Antibody solution into each well except the chromogen blanks.</li> <li>Cover the plate with a plate cover and incubate 1 hour at room temperature.</li> <li>Thoroughly aspirate the solution and wash wells 4 times with 1X Wash Buffer.</li> </ol>
<b>3</b>	<b>Add IgG HRP</b> 	<ol style="list-style-type: none"> <li>Add 100 <math>\mu</math>L 1X Anti-Rabbit IgG HRP solution into each well except the chromogen blanks.</li> <li>Cover the plate with plate cover and incubate for 30 minutes at room temperature.</li> <li>Thoroughly aspirate the solution and wash wells 4 times with 1X Wash Buffer.</li> </ol>
<b>4</b>	<b>Add Stabilized Chromogen</b> 	<ol style="list-style-type: none"> <li>Add 100 <math>\mu</math>L Stabilized Chromogen to each well. The substrate solution begins to turn blue.</li> <li>Incubate for 30 minutes at room temperature in the dark.</li> </ol> <p><b>Note:</b> TMB should not touch aluminum foil or other metals.</p>
<b>5</b>	<b>Add Stop Solution</b> 	Add 100 $\mu$ L Stop Solution to each well. Tap the side of the plate to mix. The solution in the wells changes from blue to yellow.

## Read the plate and generate the standard curve

1. Read the absorbance at 450 nm. Read the plate within 2 hours after adding the Stop Solution.
2. Use curve-fitting software to generate the standard curve. A four parameter algorithm provides the best standard curve fit. Optimally, the background absorbance may be subtracted from all data points, including standards, unknowns and controls, prior to plotting.
3. Read the concentrations for unknown samples and controls from the standard curve. Multiply value(s) obtained for sample(s) by the appropriate factor to correct for the sample dilution.

**Note:** Dilute samples producing signals greater than the upper limit of the standard curve in Standard Diluent Buffer and reanalyze. Multiply the concentration by the appropriate dilution factor.

## Performance characteristics

### Standard curve example

The following data were obtained for the various standards over the range of 0 to 20 ng/mL human EGFR.

Standard Hu EGFR (ng/mL)	Optical Density (450 nm)
20	2.88
10	1.78
5	1.00
2.5	0.61
1.25	0.33
0.63	0.23
0.32	0.15
0	0.10

### Recovery

The recovery of full length EGFR added to an EGFR-negative cell lysate (3T3-L1 cells), adjusted to 200  $\mu$ g/mL, averaged 106% when diluted in Standard Diluent Buffer.

### Inter-assay precision

Samples were assayed 36 times in multiple assays to determine precision between assays.

Parameters	Sample 1	Sample 2	Sample 3
Mean (ng/mL)	9.61	4.88	2.16
Standard Deviation	0.79	0.52	0.16
% Coefficient of Variation	6.66	10.71	7.21

### Intra-assay precision

Samples of known full length human EGFR concentration were assayed in replicates of 12 to determine precision within an assay.

Parameters	Sample 1	Sample 2	Sample 3
Mean (ng/mL)	9.58	4.88	2.19
Standard Deviation	0.56	0.43	0.15
% Coefficient of Variation	5.83	8.79	6.68

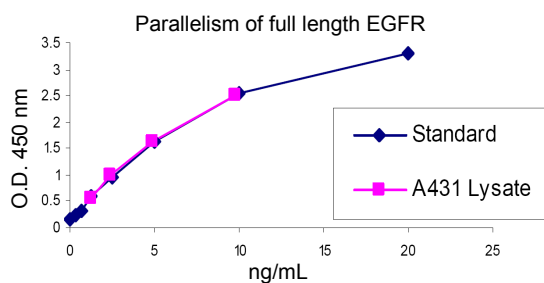
## Linearity of dilution

A431 lysate in Cell Extraction Buffer was adjusted to 10 ng/mL EGFR and serially diluted in Standard Diluent Buffer over the range of the assay. Linear regression analysis of samples versus the expected concentration yielded a correlation coefficient of 0.99 in both cases.

Dilution	Measured (ng/mL)	Expected	
		(ng/mL)	%
1/2	9.74	9.74	100
1/4	4.92	4.87	101
1/8	2.79	2.43	106
1/16	1.32	1.22	109
1/32	0.65	0.61	107

## Parallelism

Natural EGFR from A431 cell lysate was serially diluted in Standard Diluent Buffer. The optical density of each dilution was plotted against the EGFR standard curve. The standard accurately reflects full length EGFR content in samples.



## Sensitivity

The analytical sensitivity of the assay is <0.300 ng/mL full length human EGFR. This was determined by adding two standard deviations to the mean O.D. obtained from 30 assays of the zero standard.

The sensitivity of the ELISA is ~10-fold greater than that of western blot.

### Detection of full length EGFR by ELISA vs western blot

Western blot (170 kDa)



ELISA: OD 450 nm

0.10	0.21	0.29	0.40	0.73	1.16	1.96	2.89
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EGFR (ng/test)

0	0.02	0.03	0.06	0.13	0.25	0.5	1
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## Limited product warranty

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### Product label explanation of symbols and warnings

<b>REF</b>	Catalog Number	<b>LOT</b>	Batch code		Temperature limitation		Use by		Manufacturer		Consult instructions for use		Caution, consult accompanying documents
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**Manufacturer's address:** Bender MedSystems GmbH | Campus Vienna Biocenter 2 | 1030 Vienna, Austria

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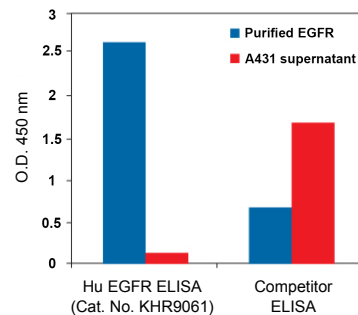
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## Specificity

The Human EGFR (Full length) ELISA Kit is specific for full length human EGFR protein and detects cellular levels of EGFR, but not soluble EGFR in the supernatant of A431 cell culture.



The kit recognizes full length EGFR regardless of phosphorylation state. The results obtained when purified EGFR was autophosphorylated *in vitro* indicate that the assay detects both phosphorylated and non-phosphorylated forms of EGFR, whereas the phosphorylation site specific EGFR ELISAs only react with phosphorylated protein.

