

Tricine Mini Gels

	Package Contents	<table border="1"> <thead> <tr> <th>Product</th> <th>Quantity</th> </tr> </thead> <tbody> <tr> <td>10% Tricine Gels</td> <td>Box of 10 gels</td> </tr> <tr> <td>16% Tricine Gels</td> <td>Box of 10 gels</td> </tr> <tr> <td>10–20% Tricine Gels</td> <td>Box of 10 gels</td> </tr> </tbody> </table>	Product	Quantity	10% Tricine Gels	Box of 10 gels	16% Tricine Gels	Box of 10 gels	10–20% Tricine Gels	Box of 10 gels
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10% Tricine Gels	Box of 10 gels									
16% Tricine Gels	Box of 10 gels									
10–20% Tricine Gels	Box of 10 gels									
	Storage Conditions	<ul style="list-style-type: none"> Store at 2–8°C for a 4 to 8-week shelf life, depending on gel type. Do not freeze. 								
	Required Materials	<ul style="list-style-type: none"> Protein sample and standard Tricine SDS Running Buffer (10X) Tricine SDS Sample Buffer (2X) NuPAGE® Sample Reducing Agent (10X) Novex® Power Supply Adapters (Cat. no. ZA10001) if not using a Life Technologies™ power supply XCell <i>SureLock</i>™ Mini-Cell gel running tank 								
	Timing	<p>Run Time: ~90 minutes (depending on gel percentage)</p> <p>Voltage: 125 V constant</p>								
	Selection Guide	<p>Specialized Protein Gels Go online to view related products.</p>								
	Product Description	<p>Tricine Gels are precast polyacrylamide gels designed for optimal separation and resolution of low molecular weight proteins and peptides (2–200 kDa) under denaturing gel electrophoresis conditions.</p> <p>Tricine Mini Gels are available in the following variations:</p> <ul style="list-style-type: none"> Polyacrylamide percentages: 10%, 16%, and 10–20% Well formats: 5, 10, 12, 15, and 1D wells Thickness: 1.0 mm 								
	Important Guidelines	<ul style="list-style-type: none"> This system is designed for use in the XCell <i>SureLock</i>® Mini-Cell gel running tank. 								
	Online Resources	<p>Visit our product page for additional information and protocols. For support, visit www.lifetechnologies.com/support.</p>								



Protocol Outline

- Prepare samples, buffers, and gels.
- Assemble the gel apparatus.
- Load buffer, samples, and standards.
- Perform electrophoresis.

Electrophoresis Protocol

- i** See page 2 to view a procedure for preparing and running your electrophoresis experiment.

Choosing the Right Gel Type for Your Application

- i** Review the table in the pop-up to determine the best gel type for your experiment.

Choosing the Right Gel Percentage and Buffer

- i** Refer to the migration chart in the pop-up to find the gel best suited for your application. As a general rule, your proteins of interest should migrate through ~70% of the length of the gel for the best resolution. When protein molecular weights are wide ranging or unknown, gradient gels are usually the best choice.

Choosing a Well Format and Gel Thickness

- i** We offer polyacrylamide gels in a choice of nine well formats and two thicknesses, depending on the gel type. When loading large samples (>30 µL), a thicker gel with fewer wells is more appropriate; Bolt™ Bis-Tris Plus gels are the best choice when loading large samples. When blotting, however, proteins will transfer more easily from a thinner gel.

Choosing a Protein Standard for your Application

Choose a Life Technologies™ standard based on your experiment:

Pre-stained: SeeBlue® Plus2 Pre-Stained Standard or Novex® Sharp Pre-Stained Standard

Unstained: Novex® Sharp Unstained Protein Standard or Mark12™ Unstained Standard

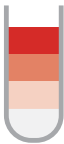

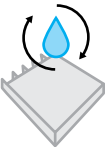
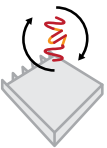
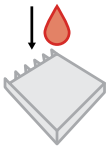

Western: MagicMark™ XP Western Protein Standard

For all other specialty standards, please view further information [here](#).

i Limited Product Warranty and Disclaimer Details

Tricine Mini Gel Electrophoresis Protocol

Follow the procedure below to prepare for and perform SDS polyacrylamide gel electrophoresis using Tricine Mini Gels.

Timeline	Steps	Procedure Details																		
1 	Prepare samples	<table border="1"> <thead> <tr> <th>Components</th> <th>Reduced Sample</th> <th>Non-Reduced Sample</th> </tr> </thead> <tbody> <tr> <td>Sample</td> <td>x μL</td> <td>x μL</td> </tr> <tr> <td>Tricine SDS Sample Buffer (2X)</td> <td>5 μL</td> <td>5 μL</td> </tr> <tr> <td>NuPAGE® Reducing Agent (10X)</td> <td>1 μL</td> <td>--</td> </tr> <tr> <td>Deionized Water</td> <td>to 4 μL</td> <td>to 5 μL</td> </tr> <tr> <td>Total Volume</td> <td>10 μL</td> <td>10 μL</td> </tr> </tbody> </table> <p>Heat at 85°C for 2 minutes. Prepare 1X Sample Buffer for dilutions of samples, if needed.</p>	Components	Reduced Sample	Non-Reduced Sample	Sample	x μL	x μL	Tricine SDS Sample Buffer (2X)	5 μL	5 μL	NuPAGE® Reducing Agent (10X)	1 μL	--	Deionized Water	to 4 μL	to 5 μL	Total Volume	10 μL	10 μL
Components	Reduced Sample	Non-Reduced Sample																		
Sample	x μL	x μL																		
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NuPAGE® Reducing Agent (10X)	1 μL	--																		
Deionized Water	to 4 μL	to 5 μL																		
Total Volume	10 μL	10 μL																		
2 	Prepare buffers	Add 100 mL of 10X Tricine SDS Running Buffer to 900 mL of deionized water to prepare 1X Tricine SDS Running Buffer.																		
3 	Prepare gels	<ol style="list-style-type: none"> Remove the comb, and rinse the gel wells three times using 1X Running Buffer. Remove the white tape near the bottom of the gel cassettes. Place the gels in the XCell SureLock® Mini-Cell gel running tank. Fill the gel wells with 1X Running Buffer. 																		
4 	Load samples and standards	Load the appropriate volume and protein mass of your sample on the gel. Then, load your standards.																		
5 	Load buffers	Fill the Upper (200 mL) and Lower (600 mL) Buffer Chambers with 1X Running Buffer.																		
6 	Run	<p>Note: If you are not using a Life Technologies™ power supply, install the Novex® Power Supply Adapters (Catalog number ZA10001).</p> <p>Run for ~90 minutes (depending on gel percentage and electrophoresis device) at 125 V constant.</p>																		