

Human FGF basic ELISA Kit

Catalog Number KHG0021 (96 tests), KHG0022 (2 × 96 tests)

Pub. No. MAN0004008 Rev. 3.0 (31)

CAUTION! This kit contains materials with small quantities of sodium azide. Sodium azide reacts with lead and copper plumbing to form explosive metal azides. Upon disposal, flush drains with a large volume of water to prevent azide accumulation. Avoid ingestion and contact with eyes, skin and mucous membranes. In case of contact, rinse affected area with plenty of water. Observe all federal, state, and local regulations for disposal.

Note: For safety and biohazard guidelines, see the “Safety” appendix in the *ELISA Technical Guide* (Pub. no. MAN0006706). Read the Safety Data Sheets (SDSs) and follow the handling instructions. Wear appropriate protective eyewear, clothing, and gloves.

Product description

The Invitrogen™ Human FGF basic ELISA Kit (Hu FGF-b) is a solid-phase sandwich Enzyme-Linked Immunosorbent Assay (ELISA). This assay is designed to detect and quantify the level of human FGF-b in human serum, plasma, buffered solution, or cell culture medium. The assay will recognize both the natural 18 kDa isoform and recombinant FGF-b-146 and FGF-b-157.

Contents and storage

Upon receipt, store the kit at 2°C to 8°C.

Contents	Cat. No. KHG0021 (96 tests)
Hu FGF-b Standard, recombinant Hu FGF-b-146 expressed in <i>E. coli</i> ; contains 0.1% sodium azide	2 vials
Standard Diluent Buffer; contains 0.1% sodium azide	25 mL
Antibody Coated Wells, 96-well plate	1 plate
Hu FGF-b Biotin Conjugate; contains 0.1% sodium azide	11 mL
Streptavidin-HRP (100X); contains 3.3 mM thymol	0.125 mL
Streptavidin-HRP Diluent; contains 3.3 mM thymol	25 mL
Wash Buffer Concentrate (25X)	100 mL
Stabilized Chromogen, Tetramethylbenzidine (TMB)	25 mL
Stop Solution	25 mL
Adhesive Plate Covers	3

Materials required but not supplied

- Distilled or deionized water
- Calibrated adjustable precision pipettes and glass or plastic tubes for diluting solutions; beakers, flask and cylinders for preparation of reagents
- Microtiter plate reader with software capable of measurement at or near 450 nm
- Plate washer—automated or manual (squirt bottle, manifold dispenser, or equivalent)

Before you begin

IMPORTANT! Reagents are lot-specific. Do not mix or interchange different reagent lots from various kit lots.

- Review the **Procedural guidelines** and **Plate washing directions** in the *ELISA Technical Guide* available at thermofisher.com.
- Allow reagents to reach room temperature before use. Mix to redissolve any precipitated salts.

Prepare 1X Wash Buffer

1. Dilute 16 mL of Wash Buffer Concentrate (25X) with 384 mL of deionized or distilled water. Label as 1X Wash Buffer.
2. Store the concentrate and 1X Wash Buffer in the refrigerator. Use the diluted buffer within 14 days.

Sample preparation guidelines

- Refer to the *ELISA Technical Guide* at thermofisher.com for detailed sample preparation procedures.
- Collect samples in pyrogen/endotoxin-free tubes.
- Freeze samples after collection if samples will not be tested immediately. Avoid multiple freeze-thaw cycles of frozen samples. Thaw completely and mix well (do not vortex) prior to analysis.
- Avoid the use of hemolyzed or lipemic sera. If large amounts of particulate matter are present in the sample, centrifuge or filter sample prior to analysis.

Pre-dilute samples

Sample concentrations should be within the range of the standard curve. Because conditions may vary, each investigator should determine the optimal dilution for each application.

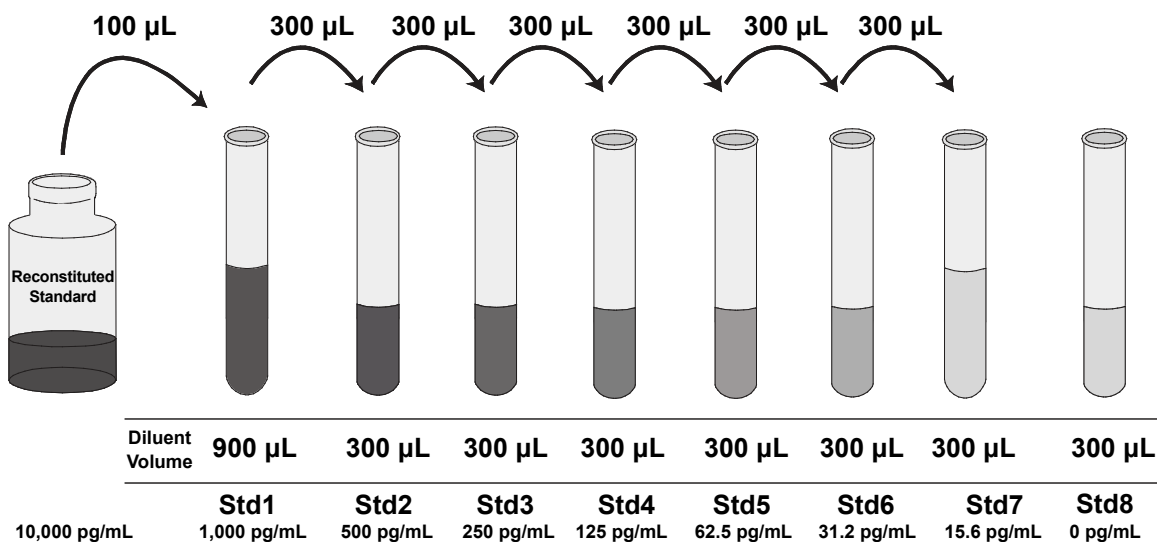
- Perform sample dilutions with Standard Diluent Buffer.
- Dilute serum, plasma, buffered solution, and cell culture medium two-fold in Standard Diluent Buffer.

Dilute standards

Note: Use glass or plastic tubes for diluting standards.

Note: This assay has been calibrated against WHO reference preparation 90/712 (version 2) (NIBSC, Hertfordshire, UK, EN6 3QG). One microgram equals 5450 International Units.

1. Reconstitute Hu FGF-b Standard to 10,000 pg/mL with Standard Dilution Buffer. Refer to the standard vial label for instructions. Swirl or mix gently and allow the contents to sit for 10 minutes to ensure complete reconstitution. Label as 10,000 pg/mL human FGF-b. **Use the standard within 1 hour of reconstitution.**
2. Add 100 μ L Reconstituted Standard to one tube containing 900 μ L Standard Diluent Buffer and mix. Label as 1,000 pg/mL human FGF-b.
3. Add 300 μ L Standard Diluent Buffer to each of 7 tubes labeled as follows: 500, 250, 125, 62.5, 31.2, 15.6, and 0 pg/mL human FGF-b.
4. Make serial dilutions of the standard as shown in the following dilution diagram. Mix thoroughly between steps.
5. Discard any remaining reconstituted standard. Return the Standard Diluent Buffer to the refrigerator.



Prepare 1X Streptavidin-HRP solution

Note: Prepare 1X Streptavidin-HRP within 15 minutes of usage.

The Streptavidin-HRP (100X) is in 50% glycerol, which is viscous. To ensure accurate dilution:






1. For each 8-well strip used in the assay, pipet 10 μ L Streptavidin-HRP (100X) solution, wipe the pipette tip with clean absorbent paper to remove any excess solution, and dispense the solution into a tube containing 1 mL of Streptavidin-HRP Diluent. Mix thoroughly.
2. Return the unused Streptavidin-HRP (100X) solution to the refrigerator.

Perform ELISA (Total assay time: 4 hours)

IMPORTANT! Perform a standard curve with each assay.

- Allow all components to reach room temperature before use. Mix all liquid reagents prior to use.
- Determine the number of 8-well strips required for the assay. Insert the strips in the frames for use. Re-bag any unused strips and frames, and store at 2°C to 8°C for future use.



1	Bind antigen 	<ol style="list-style-type: none"> Add 100 μL of Standard Diluent Buffer to zero wells. Leave the wells for chromogen blanks empty. Add 100 μL of standards to the appropriate wells. For all samples (serum, plasma, buffered solution, or cell culture medium), add 50 μL of Standard Diluent Buffer followed by 50 μL of sample (see “Pre-dilute samples” on page 2) to the appropriate wells. Leave the wells for chromogen blanks empty. Tap the side of the plate to mix. Cover the plate with a plate cover and incubate for 2 hours at room temperature. Thoroughly aspirate the solution and wash wells 4 times with 1X Wash Buffer.
2	Add Biotin Conjugate 	<ol style="list-style-type: none"> Add 100 μL Hu FGF-b Biotin Conjugate solution into each well except the chromogen blanks. Cover the plate with plate cover and incubate for 1 hour at room temperature . Thoroughly aspirate the solution and wash wells 4 times with 1X Wash Buffer.
3	Add Streptavidin-HRP 	<ol style="list-style-type: none"> Add 100 μL 1X Streptavidin-HRP solution (see page 2) into each well except the chromogen blanks. Cover the plate with a plate cover and incubate for 30 minutes at room temperature. Thoroughly aspirate the solution from the wells and wash wells 4 times with 1X Wash Buffer.
4	Add Stabilized Chromogen 	<ol style="list-style-type: none"> Add 100 μL Stabilized Chromogen to each well. The substrate solution begins to turn blue. Incubate for 30 minutes at room temperature in the dark. <p>Note: TMB should not touch aluminum foil or other metals.</p>
5	Add Stop Solution 	Add 100 μ L Stop Solution to each well. Tap the side of the plate to mix. The solution in the wells changes from blue to yellow.

Read the plate and generate the standard curve

1. Read the absorbance at 450 nm. Read the plate within 2 hours after adding the Stop Solution.
2. Use curve-fitting software to generate the standard curve. A 4 parameter algorithm provides the best standard curve fit. Optimally, the background absorbance may be subtracted from all data points, including standards, unknowns and controls, prior to plotting.
3. Read the concentrations for unknown samples and controls from the standard curve. Multiply value(s) obtained for sample(s) by the appropriate factor to correct for the sample dilution.

Note: Dilute samples producing signals greater than the upper limit of the standard curve in Standard Diluent Buffer and reanalyze. Multiply the concentration by the appropriate dilution factor.

Performance characteristics

Standard curve example

The following data were obtained for the various standards over the range of 0 to 1,000 pg/mL human FGF-b.

Standard Human FGF basic (pg/mL)	Optical Density (450 nm)
1,000	2.41
500	1.36
250	0.73
125	0.38
62.5	0.22
31.2	0.13
15.6	0.08
0	0.04

Inter-assay precision

Samples were assayed 42 times in multiple assays to determine precision between assays.

Parameters	Sample 1	Sample 2	Sample 3
Mean (pg/mL)	89.7	214.0	847.0
Standard Deviation	4.8	9.5	56.9
% Coefficient of Variation	5.4	4.4	6.7

Intra-assay precision

Samples of known human FGF-b concentration were assayed in replicates of 14 to determine precision within an assay.

Parameters	Sample 1	Sample 2	Sample 3
Mean (pg/mL)	86.0	213.0	800.0
Standard Deviation	3.5	8.0	39.9
% Coefficient of Variation	4.1	3.8	5.0

Cross-reactivity

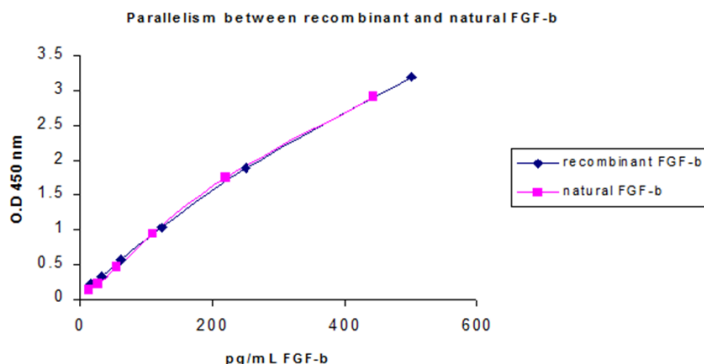
Bovine FGF basic showed 10% cross-reactivity.

Linearity of dilution

Human serum and tissue culture medium containing 10% fetal bovine serum were spiked with human FGF-b and serially diluted in *Standard Diluent Buffer* over the range of the assay. Linear regression analysis of samples versus the expected concentration yielded a correlation coefficient of 0.99 in both cases.

Parallelism

Natural human FGF-b derived from bronchial smooth muscle cells was serially diluted in Standard Diluent Buffer. The optical density of each dilution was plotted against the standard curve. The standard accurately reflects natural human FGF-b content in samples.



Limited product warranty

Life Technologies Corporation and/or its affiliate(s) warrant their products as set forth in the Life Technologies' General Terms and Conditions of Sale found on Life Technologies' website at www.thermofisher.com/us/en/home/global/terms-and-conditions.html. If you have any questions, please contact Life Technologies at www.thermofisher.com/support.

Product label explanation of symbols and warnings

REF	Catalog Number	LOT	Batch code	Temperature limitation	Use by	Manufacturer	Consult instructions for use	Caution, consult accompanying documents
-----	----------------	-----	------------	------------------------	--------	--------------	------------------------------	---

Manufacturer's address: Bender MedSystems GmbH | Campus Vienna Biocenter 2 | 1030 Vienna, Austria

The information in this guide is subject to change without notice.

DISCLAIMER: TO THE EXTENT ALLOWED BY LAW, LIFE TECHNOLOGIES AND/OR ITS AFFILIATE(S) WILL NOT BE LIABLE FOR SPECIAL, INCIDENTAL, INDIRECT, PUNITIVE, MULTIPLE, OR CONSEQUENTIAL DAMAGES IN CONNECTION WITH OR ARISING FROM THIS DOCUMENT, INCLUDING YOUR USE OF IT.

Important Licensing Information: These products may be covered by one or more Limited Use Label Licenses. By use of these products, you accept the terms and conditions of all applicable Limited Use Label Licenses.

©2017 Thermo Fisher Scientific Inc. All rights reserved. All trademarks are the property of Thermo Fisher Scientific and its subsidiaries unless otherwise specified.

Recovery

The recovery of human FGF-b added to EDTA plasma averaged 89%. The recovery of human FGF-b added to tissue culture medium containing 10% fetal bovine serum averaged 90%. Heparin and citrate plasma are not recommended.

Sensitivity

The analytical sensitivity of the assay is <15.6 pg/mL human FGF-b. This was determined by adding two standard deviations to the mean O.D. obtained when the zero standard was assayed 30 times, and calculating the corresponding concentration.

Specificity

Buffered solutions of a panel of substances at 10,000 pg/mL were assayed with the Human FGF basic ELISA Kit. The following substances were tested and found to have no cross-reactivity: **Human** IL-1 b, IL-2, IL-4, IL-5, IL-6, IL-8, IL-10, IL-12, IL-13, IL-15, IL-17, DR5, Eotaxin, EGF, FGF acidic, G-CSF, GM-CSF, IFN- g, MIP-1 a, MIP-1 b, TNF- a; **bovine** FGF acidic.