# CTS<sup>™</sup> RevitaCell<sup>™</sup> Supplement (100X)

#### Catalog Number A4238401

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**WARNING!** Read the Safety Data Sheets (SDSs) and follow the handling instructions. Wear appropriate protective eyewear, clothing, and gloves. Safety Data Sheets (SDSs) are available from **thermofisher.com/support**.

# **Product description**

Gibco<sup>™</sup> CTS<sup>™</sup> RevitaCell<sup>™</sup> Supplement (100X) is a chemically defined recovery supplement containing a specific ROCK inhibitor coupled with molecules that have antioxidant and free radical scavenger properties. Use CTS<sup>™</sup> RevitaCell<sup>™</sup> Supplement to achieve efficient and optimal post-thaw recovery of cryopreserved pluripotent stem cells (PSCs). CTS<sup>™</sup> RevitaCell<sup>™</sup> Supplement can also be used for the recovery of PSCs from single-cell passaging procedures using dissociation reagents such as CTS<sup>™</sup> TrypLE<sup>™</sup> Select Enzyme.

### Contents and storage

Contents	Amount	Storage	Shelf Life <sup>[1]</sup>
CTS™ RevitaCell™ Supplement (100X)	5 mL	–20°C to –5°C. Protect from light	12 months

<sup>[1]</sup> Shelf Life duration is determined from Date of Manufacture.

# Procedural guidelines

Divide thawed CTS<sup>™</sup> RevitaCell<sup>™</sup> Supplement (100X) into usagesize aliquots and store in a non-frost-free freezer at −20°C to −5°C.

# **Recover cryopreserved PSCs**

- 1. Coat a culture vessel with the appropriate substrate on which to culture your PSCs.
- **2.** Quickly thaw cryopreserved PSCs in a 37°C waterbath until only a small ice crystal remains.
- **3.** Gently pipet the thawed cells up and down to create a cell suspension and transfer to a 50-mL conical tube.
- 4. Dilute the cell suspension with 3 mL of growth medium, adding it dropwise while gently rocking the tube back and forth to avoid osmotic shock to the cells.
- **5.** Centrifuge the cell suspension at  $200 \times g$  for 4 minutes.
- 6. Aspirate the medium.

Note: Be careful not to disturb the cell pellet.

 Gently resuspend the cells in growth medium supplemented with CTS<sup>™</sup> RevitaCell<sup>™</sup> Supplement at a 1X final concentration (i.e., 100 µL of CTS<sup>™</sup> RevitaCell<sup>™</sup> Supplement in 10 mL of growth medium).

**Note:** Do not add any additional ROCK inhibitors to the growth medium.

**8.** Transfer the cell suspension to an appropriate pre-coated culture vessel.

**Note:** See "Recommended reagent volumes and seeding densities".

- **9.** Move the vessel in several quick back-and-forth and side-to-side motions to disperse the cells across its surface.
- **10.** Incubate the cells for 18–24 hours in the recommended cell culture environment.
- 11. Following incubation, aspirate the growth medium supplemented with CTS<sup>™</sup> RevitaCell<sup>™</sup> Supplement and replace it with unsupplemented growth medium (i.e., without the addition of CTS<sup>™</sup> RevitaCell<sup>™</sup> Supplement) for the remainder of the culture.

# Recover single-cell passaged PSCs in CTS<sup>™</sup> Essential 8<sup>™</sup> Medium

1. Coat the culture vessels with the appropriate substrate on which to culture your PSCs.

The recommended substrate is CTS<sup>™</sup> Vitronectin (VTN-N) Recombinant Human Protein, Truncated (Cat. No. A27940).

- 2. Aspirate the spent medium from the culture vessel.
- Rinse the vessel once with recommended volume of CTS<sup>™</sup> DPBS without calcium chloride, without magnesium chloride (see Table 1).



- 4. Add recommended volume of CTS<sup>™</sup> TrypLE<sup>™</sup> Select Enzyme to the vessel (see Table 1).
- 5. Incubate the vessel at 37°C, 5% CO<sub>2</sub> for 5 minutes, continually observing the wells for cell detachment.

**Note:** Avoid extended incubation of cells with dissociation reagents to minimize damage to cells.

- **6.** Gently pipet the cells up and down 5–10 times to generate single cell suspension.
- Transfer the cell suspension to a conical tube containing the recommended volume of CTS<sup>™</sup> Essential 8<sup>™</sup> Medium to dilute the dissociation reagent (see Table 1).
- **8.** Centrifuge the cells at  $200 \times g$  for 4 minutes.
- 9. Discard the supernatant, flick the tube 3–5 times to loosen the pellet, and resuspend the cells by pipetting them up and down 5–10 times in the recommended volume of CTS<sup>™</sup> Essential 8<sup>™</sup> Medium supplemented with CTS<sup>™</sup> RevitaCell<sup>™</sup> Supplement at a 1X final concentration (see Table 1).
- Determine viable cell density and percent viability using a Countess<sup>™</sup> II Automated Cell Counter or a similar automated or manual method.

# Recommended reagent volumes and seeding densities

 Table 1
 Reagent volumes (per well or per dish)

 Adjust the concentration of the cell suspension using CTS<sup>™</sup> Essential 8<sup>™</sup> Medium supplemented with CTS<sup>™</sup> RevitaCell<sup>™</sup> Supplement at a 1X final concentration to achieve the cell seeding density recommended for your culture vessel (see Table 2).

**Note:** Do not add any additional ROCK inhibitors to the growth medium.

12. Transfer the cell suspension to the pre-coated culture vessel.

Move the vessel in several quick back-and-forth and side-toside motions to disperse the cells across its surface.

- **13.** Incubate the cells for 18–24 hours in the recommended cell culture environment.
- 14. Following incubation, aspirate the CTS<sup>™</sup> Essential 8<sup>™</sup> Medium supplemented with CTS<sup>™</sup> RevitaCell<sup>™</sup> Supplement and replace it with unsupplemented CTS<sup>™</sup> Essential 8<sup>™</sup> Medium (i.e., without the addition of CTS<sup>™</sup> RevitaCell<sup>™</sup> Supplement) for the remainder of the culture.

Culture vessel (surface area)	CTS™ DPBS for wash	CTS <sup>™</sup> TrypLE <sup>™</sup> Select Enzyme	CTS <sup>™</sup> Essential 8 <sup>™</sup> Medium <sup>[1]</sup>	CTS <sup>™</sup> Essential 8 <sup>™</sup> Medium + 1X CTS <sup>™</sup> RevitaCell <sup>™</sup> Supplement <sup>[2]</sup>
6-well (10 cm <sup>2</sup> )	2 mL	1 mL	3 mL	2 mL
12-well (4 cm <sup>2</sup> )	1 mL	0.4 mL	1.2 mL	1 mL
24-well (2 cm <sup>2</sup> )	0.5 mL	0. 2 mL	0.6 mL	0.5 mL
35-mm (10 cm <sup>2</sup> )	2 mL	1 mL	3 mL	2 mL
60-mm (20 cm <sup>2</sup> )	4 mL	2 mL	6 mL	4 mL
100-mm (60 cm²)	12 mL	6 mL	18 mL	12 mL

<sup>[1]</sup> For dilution

<sup>[2]</sup> For resuspension

#### Table 2 Recommended cell seeding densities and volumes of medium for plating (per well or per dish)

	Number of viab	CTS <sup>™</sup> Essential 8 <sup>™</sup> Medium + 1X CTS <sup>™</sup>		
Culture vessel (surface area)	12,500 cells/cm <sup>2</sup> 25,000 cells/c		RevitaCell <sup>™</sup> Supplement <sup>[2]</sup>	
6-well (10 cm <sup>2</sup> )	125,000	250,000	2 mL	
12-well (4 cm <sup>2</sup> )	50,000	100,000	1 mL	
24-well (2 cm <sup>2</sup> )	25,000	50,000	0.5 mL	
35-mm (10 cm²)	125,000	250,000	2 mL	
60-mm (20 cm <sup>2</sup> )	250,000	500,000	4 mL	
100-mm (60 cm <sup>2</sup> )	750,000	1,500,000	12 mL	

<sup>[1]</sup> Time to confluency is 4–5 days for a seeding density of 12,500 cells/cm<sup>2</sup> and 3–4 days for a seeding density of 25,000 cells/cm<sup>2</sup>

<sup>[2]</sup> For resuspension

# **Related products**

Unless otherwise indicated, all materials are available through thermofisher.com.

Item	Source
CTS™ Essential 8™ Medium	A2656101
CTS™ Vitronectin (VTN-N) Recombinant Human Protein, Truncated	A27940
CTS <sup>™</sup> Versene Solution	A42391
CTS™ TrypLE™ Select Enzyme	A12859
CTS <sup>™</sup> DPBS without calcium chloride, without magnesium chloride	A12856
CTS <sup>™</sup> PSC Cryomedium	A42388
CTS <sup>™</sup> PSC Cryopreservation Kit	A42393

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