


CTS™ AIM V™ Medium

Serum-free medium without Gentamicin sulfate, Phenol red, and Streptomycin sulfate

Catalog Number A4672701

Pub. No. MAN0018999 Rev. 1.0

 **WARNING!** Read the Safety Data Sheets (SDSs) and follow the handling instructions. Wear appropriate protective eyewear, clothing, and gloves. Safety Data Sheets (SDSs) are available from thermofisher.com/support.

Product description

Gibco™ CTS™ AIM V™ Medium is the first commercially available defined serum-free medium for proliferation and/or manipulation of T-cells and dendritic cells, manufactured in compliance with cGMP (21 CFR part 820). CTS™ AIM V™ Medium contains L-glutamine at 0.3358 mg/mL. CTS™ AIM V™ Medium does not contain gentamicin sulfate, phenol red, nor streptomycin sulfate. Each container is a sterile filtered single-use container.

Contents and storage

Contents	Cat. No.	Amount	Storage	Shelf life ^[1]
CTS™ AIM V™ Medium	A4672701	2 L	2°C to 8°C; Protect from light	12 months

^[1] Shelf Life duration is determined from Date of Manufacture.

Safety information

Human origin raw materials are non-reactive (donor level) for anti-HIV 1 & 2, anti-HCV and HBsAg. Handle in accordance with established bio-safety practices.

Culture conditions

Media: CTS™ AIM V™ Medium

Cells: Human peripheral blood mononuclear cells (PBMCs)

Culture type: Static suspension

Culture vessels: T-Flasks or cell culture bag

Temperature range: 36°C to 38°C

Incubator atmosphere: Humidified atmosphere of 4–6% CO₂ in air. Ensure proper gas exchange and minimize exposure of cultures to light.

Procedural guidelines

- CTS™ AIM V™ Medium comes supplemented with L-glutamine.
- Additional supplementation with cytokines or growth factors may be required per specific investigator's procedures and should be aseptically added immediately prior to use.
- The following protocol serves as a general guideline for static T cell and dendritic cell culture, regardless of vessel.
- Feed and maintain cells at desired concentrations while cells are in log phase growth. Dilute cells to a viable cell density of 5×10^5 cells/mL whenever the viable cell density reaches $\geq 1 \times 10^6$ cells/mL.
- For optimal gas exchange in static plate cultures, it is recommended that medium depth not exceed 1–1.2 cm.
- For high-density culture in bioreactors, optimal procedures should be determined empirically by the investigator.

T-cell culture

1. Prepare fresh peripheral blood mononuclear cells (PBMCs) or rapidly thaw (<1 minute) a vial of frozen PBMCs in a 37°C water bath according to standard PBMC thawing protocols.
2. Wash cells with CTS™ DPBS without calcium chloride, without magnesium chloride, supplemented with 2–5% heat-inactivated human pooled Type AB serum or 5–10% CTS™ Immune Cell SR.

The optimal concentration should be determined based on your application.
3. Determine viable cell density using a Countess™ II Automated Cell Counter.
4. Centrifuge cells at 200 × g for 5–10 minutes and aspirate wash buffer supernatant.
5. Resuspend PBMC pellet at approximately 0.5 × 10⁶–1 × 10⁶ cells/mL in medium supplemented with cytokines (e.g., IL-2), if used at culture initiation.
6. Transfer the desired number of cells to the desired tissue culture vessel.

Note: A variety of protocols may be used for activating T-cells for subsequent expansion, including adding stimulatory antibodies or antigen presenting cells. Similarly, for either small or large scale T-cell expansion, cells can be isolated, activated and expanded using CTS™ Dynabeads™ CD3/CD28 according to instructions in the product insert.

Prepare monocyte derived dendritic cell culture

1. Prepare fresh PBMCs and seed into a culture flask with 25 mL RPMI 1640 or CTS™ AIM-V™ Medium (Therapeutic Grade) at cell density of 1–2 × 10⁵ cells/cm².
2. Incubate for 2–3 hours at 37°C in a humidified atmosphere of 5% CO₂ in air.
3. Aspirate and discard medium containing non-adherent cells.
4. Wash the adherent cells (mainly CD14+ monocytes) three times with CTS™ DPBS without calcium chloride, without magnesium chloride.
5. Add medium supplemented with 50–100 ng/mL recombinant human IL-4 and 50 ng/mL recombinant human GM-CSF.
6. Incubate cells at 37°C in a humidified atmosphere of 5% CO₂ in air for 5 days.
7. On day 3, transfer spent media to a sterile conical tube and centrifuge at 200 × g for 5–10 minutes to collect all non-adherent or loosely adherent cells.

8. Aspirate supernatant and gently resuspend cell pellet in an equal volume of fresh pre-warmed medium containing IL-4 and GM-CSF.
9. Transfer cell suspension to the original culture flask containing adherent cells.

After 6 days, the loosely adherent or non-adherent cells should display typical dendritic cell morphology and surface markers (e.g., CD1a, CD80, CD86, and HLA-DR).
10. Induce maturation of dendritic cells by the addition of either 1 µg/mL lipopolysaccharide (LPS) or 50 µL/mL CTS™ TNF-α to the medium.

Note: As an alternative to plastic adherence, monocytes can be isolated by magnetic separation.

Related products

Unless otherwise indicated, all materials are available through thermofisher.com.

Item	Source
CTS™ DPBS without calcium chloride, without magnesium chloride	A12856
CTS™ IL-2 Recombinant Human	CTP0021
CTS™ IL-4 Recombinant Human	CTP0041
CTS™ IL-7 Recombinant Human	CTP0071
CTS™ GM-CSF Recombinant Human	CTP2011
CTS™ TNF Recombinant Human	CTP3011
CTS™ Immune Cell SR	A2596101
CTS™ Dynabeads™ CD3/CD28	40203D
CTS™ DynaMag™ Magnet	12102
Dynabeads™ Human Treg Expander	11129D
RPMI 1640 Medium, GlutaMAX™ Supplement, HEPES	72400
CTS™ GlutaMAX™-I Supplement	A1286001
L-Glutamine (200 mM) (100X)	25030
Trypan Blue Solution, 0.4%	15250
Countess™ II Automated Cell Counter	AMQAX1000

Limited product warranty

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For descriptions of symbols on product labels or product documents, go to [thermofisher.com/symbols-definition](https://www.thermofisher.com/symbols-definition).

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