Human c-Met (Total) ELISA Kit

Catalog Number KH00251 (96 tests)

Pub. No. MAN0014647 **Rev.** 2.0 (30)

CAUTION! This kit contains materials with small quantities of sodium azide. Sodium azide reacts with lead and copper plumbing to form explosive metal azides. Upon disposal, flush drains with a large volume of water to prevent azide accumulation. Avoid ingestion and contact with eyes, skin and mucous membranes. In case of contact, rinse affected area with plenty of water. Observe all federal, state, and local regulations for disposal.

Note: For safety and biohazard guidelines, see the "Safety" appendix in the *ELISA Technical Guide* (Pub. no. MAN0006706). Read the Safety Data Sheets (SDSs) and follow the handling instructions. Wear appropriate protective eyewear, clothing, and gloves.

Product description

The Invitrogen™ Human c-Met (Total) ELISA Kit is a solid-phase sandwich Enzyme-Linked Immunosorbent Assay (ELISA). This assay is designed to detect and quantify the level of human c-Met (Total) in cell lysates. The assay recognizes both natural and recombinant human c-Met (Total).

c-Met, a member of the tyrosine kinase superfamily, is the receptor for hepatocyte growth factor, also known as scatter factor (HGF/SF). Alternative splicing yields several c-Met isoforms, including proteins that remain in the uncleaved, monomeric state or that lack various portions of the c-Met cytoplasmic domain. Cells expressing c-Met include epithelial cells, endothelial cells, blood cells of various types and glomerular mesenchymal cells.

Contents and storage

Upon receipt, store the kit at 2°C to 8°C.

Contents	Cat. No. KH00251 (96 tests)
Hu c-Met (Total) Standard, lyophilized; contains 0.1% sodium azide.	2 vials
Standard Diluent Buffer[1]; contains 0.1% sodium azide	25 mL
Antibody Coated Wells; 96-well plate	1 plate
Hu c-Met (Total) Detection Antibody; contains 0.1% sodium azide	11 mL
Anti-Rabbit IgG HRP (100X)	0.125 mL
HRP Diluent; contains 3.3 mM thymol	25 mL
Wash Buffer Concentrate (25X)	100 mL
Stabilized Chromogen, Tetramethylbenzidine (TMB)	25 mL
Stop Solution	25 mL
Plate Covers, adhesive strips	3

[1] If precipitates are found in Standard Diluent Buffer, they should be completely dissolved by warming to room temperature before use.

Required materials not supplied

- Distilled or deionized water
- Microtiter plate reader with software capable of measurement at or near 450 nm
- Plate washer–automated or manual (squirt bottle, manifold dispenser, or equivalent)
- Calibrated adjustable precision pipettes and glass or plastic tubes for diluting solutions; beakers, flask and cylinders for preparation of reagents
- Cell Extraction Buffer (Cat. No. FNN0011, or see "Prepare Cell Extraction Buffer")

Before you begin

IMPORTANT! Reagents are lot-specific. Do not mix or interchange different reagent lots from various kit lots.

- Review the **Procedural guidelines** and **Plate washing directions** in the *ELISA Technical Guide* available at **thermofisher.com**.
- Allow reagents to reach room temperature before use. Mix to redissolve any precipitated salts.

Prepare 1X Wash Buffer

1. Dilute 16 mL of Wash Buffer Concentrate (25X) with 384 mL of deionized or distilled water. Label as 1X Wash Buffer.
2. Store the concentrate and 1X Wash Buffer in the refrigerator. Use the diluted buffer within 14 days.

Prepare Cell Extraction Buffer

Note: See the *ELISA Technical Guide* for detailed information on preparing Cell Extraction Buffer.

1. Prepare Cell Extraction Buffer.

   Cell Extraction Buffer consists of 10 mM Tris (pH 7.4), 100 mM NaCl, 1 mM EDTA, 1 mM EGTA, 1 mM NaF, 20 mM $Na_4P_2O_7$, 2 mM Na_3VO_4, 1% Triton™ X-100, 10% glycerol, 0.1% SDS, and 0.5% deoxycholate.
2. Immediately before use, add PMSF (0.3 M stock in DMSO) to 1 mM and 50 µL protease inhibitor cocktail (e.g., Sigma Cat. No. P-2714) for each 1 mL of Cell Extraction Buffer.

Prepare cell lysate

1. Collect cells by centrifugation (non-adherent cells) or scraping from culture flasks (adherent cells), then wash cells twice with cold PBS.
2. Remove and discard the supernatant and collect the cell pellet. The pellet can be stored at −80°C and lysed at a later date if desired.
3. Lyse the cell pellet in Cell Extraction Buffer for 30 minutes, on ice. Vortex at 10-minute intervals.

   Note: The volume of Cell Extraction Buffer used depends on the number of cells in the cell pellet, and expression levels of human c-Met (Total). For example, 10^8 HeLa cells grown in RPMI plus 10% FBS can be extracted in 1 ml of Cell Extraction Buffer. Researchers must optimize the extraction procedures for their own applications.
4. Transfer the lysate into microcentrifuge tubes and centrifuge at 13,000 rpm for 10 minutes at 4°C.
5. Transfer the supernatant into clean microcentrifuge tubes. Samples can be stored at −80°C (avoid multiple freeze-thaw cycles).

Pre-dilute samples

Sample concentrations should be within the range of the standard curve. Because conditions may vary, each investigator should determine the optimal dilution for each application.

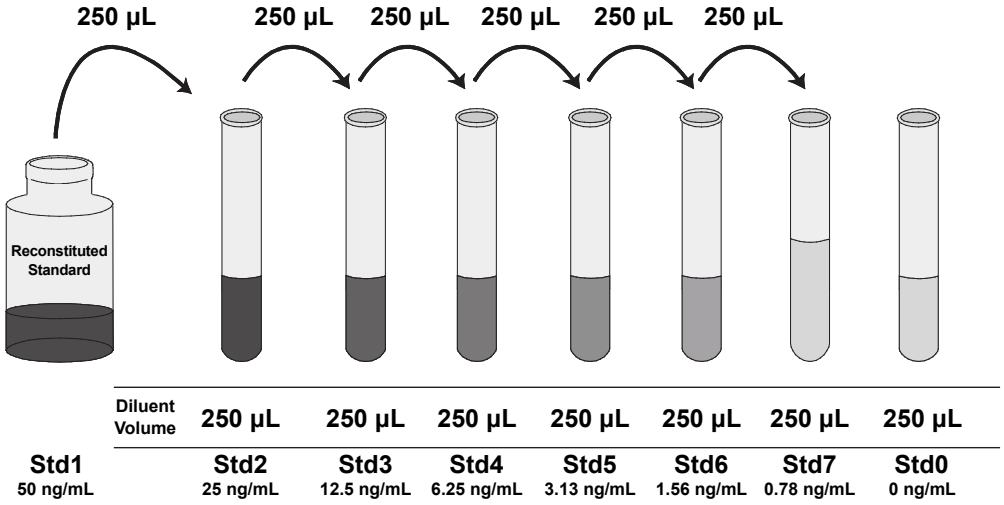
- Dilute samples prepared in Cell Extraction Buffer 1:10 or greater in Standard Diluent Buffer (e.g., 10 µL sample into 90 µL buffer). This dilution is necessary to reduce the matrix effect of the Cell Extraction Buffer. SDS concentration should be less than 0.01% before adding to the plate. While a 1:10 sample dilution has been found to be satisfactory, higher dilutions such as 1:25 or 1:50 may be optimal.
- Dilute samples >50 ng/mL with Standard Diluent Buffer.

Dilute standards

Note: Use glass or plastic tubes for diluting standards.

Note: The Hu c-Met (Total) Standard (lyophilized cell extract from GTL-16 cells) was calibrated against the mass of highly purified, truncated, recombinant human c-Met protein expressed in a mouse myeloma cell line, N50.

1. Reconstitute Hu c-Met (Total) Standard to 50 ng/mL with Standard Diluent Buffer. Refer to the standard vial label for instructions. Swirl or mix gently and allow the contents to sit for 10 minutes to ensure complete reconstitution. Label as 50 ng/mL human c-Met (Total). **Use the standard within 1 hour of reconstitution.**
2. Add 250 µL Standard Diluent Buffer to each of 7 tubes labeled as follows: 25, 12.50, 6.25, 3.13, 1.56, 0.78 and 0 ng/mL human c-Met (Total).
3. Make serial dilutions of the standard as shown in the following dilution diagram. Mix thoroughly between steps.
4. Remaining reconstituted standard should be discarded or frozen in aliquots at −80°C for further use. Standard can be frozen and thawed one time only without loss of immunoreactivity.



Prepare 1X Anti-Rabbit IgG HRP solution

Note: Prepare 1X Anti-Rabbit IgG HRP solution within 15 minutes of usage.

1. For each 8-well strip used in the assay, pipet 10 µL Anti-Rabbit IgG HRP (100X) solution, and dispense the solution into a tube containing 1 mL of HRP Diluent. Mix thoroughly.
2. Return the unused Anti-Rabbit IgG HRP (100X) solution to the refrigerator.

Perform ELISA (Total assay time: 4 hours)

IMPORTANT! Perform a standard curve with each assay.

- Allow all components to reach room temperature before use. Mix all liquid reagents prior to use.
- Determine the number of 8-well strips required for the assay. Insert the strips in the frames for use. Re-bag any unused strips and frames, and store at 2°C to 8°C for future use.

1 Bind antigen

a. Add 100 µL of standards, controls, or samples (see "Pre-dilute samples" on page 2) to the appropriate wells. Leave the wells for chromogen blanks empty. Tap gently on side of the plate to mix
b. Cover the plate with a plate cover and incubate 2 hours at room temperature.
c. Thoroughly aspirate the solution and wash wells 4 times with 1X Wash Buffer.

2 Add detector antibody

a. Add 100 µL of Hu c-Met (Total) Detection Antibody solution into each well except the chromogen blanks. Tap gently on side of plate to mix.
b. Cover the plate with a plate cover and incubate 1 hours at room tempurature.
c. Thoroughly aspirate the solution and wash wells 4 times with 1X Wash Buffer.

3 Add IgG HRP

a. Add 100 µL 1X Anti-Rabbit IgG HRP Solution into each well except the chromogen blanks.
b. Cover the plate with plate cover and incubate for 30 minutes at room temperature.
c. Thoroughly aspirate the solution and wash wells 4 times with 1X Wash Buffer.

4 Add Stabilized Chromogen

a. Add 100 µL Stabilized Chromogen to each well. The substrate solution begins to turn blue.
b. Incubate for 30 minutes at room temperature in the dark.

Note: TMB should not touch aluminum foil or other metals.

5 Add Stop Solution

Add 100 µL Stop Solution to each well. Tap the side of the plate to mix. The solution in the wells changes from blue to yellow.

Read the plate and generate the standard curve

1. Read the absorbance at 450 nm. Read the plate within 2 hours after adding the Stop Solution.
2. Use curve-fitting software to generate the standard curve. A four parameter algorithm provides the best standard curve fit. Optimally, the background absorbance may be subtracted from all data points, including standards, unknowns and controls, prior to plotting.
3. Read the concentrations for unknown samples and controls from the standard curve. Multiply value(s) obtained for sample(s) by the appropriate factor to correct for the sample dilution.

   Note: Dilute samples producing signals greater than the upper limit of the standard curve in Standard Diluent Buffer and reanalyze. Multiply the concentration by the appropriate dilution factor.

Performance characteristics

Standard curve example

The following data were obtained for the various standards over the range of 0 to 50 ng/mL human c-Met (Total).

Standard Human c-Met (Total) (ng/mL)	Optical Density (450 nm)
50	2.76
25	1.87
12.5	1.25
6.25	0.95
3.13	0.72
1.57	0.49
0.78	0.39
0	0.17

Inter-assay precision

Samples were assayed 48 times in multiple assays to determine precision between assays

Parameters	Sample 1	Sample 2	Sample 3
Mean (ng/mL)	37.1	16.6	8.0
Standard Deviation	3.9	1.8	0.9
% Coefficient of Variation	10.4	10.6	11.4

Intra-assay precision

Samples of known human c-Met (Total) concentration were assayed in replicates of 16 to determine precision within an assay.

Parameters	Sample 1	Sample 2	Sample 3
Mean (ng/mL)	37.8	16.3	8.4
Standard Deviation	3.4	1.4	0.7
% Coefficient of Variation	9.1	8.4	8.4

Linearity of dilution

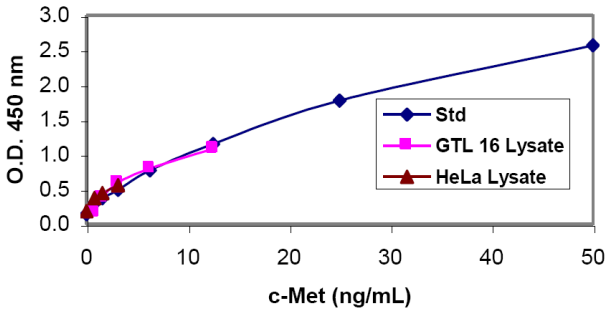
GTL-16 cells grown in cell culture medium containing 10% fetal calf serum were treated with sodium orthovanadate at 1 mM for 16 hours and lysed with cell extraction buffer. This lysate was diluted in Standard Diluent Buffer over the range of the assay and measured for human c-Met (Total) content. Linear regression analysis of samples versus the expected concentration yielded a correlation coefficient of 0.99.

Dilution	Cell Lysate		
	Measured (ng/mL)	Expected	
		(ng/mL)	%
Neat	50.0	50.0	100
1/2	29.6	25.0	118
1/4	15.8	12.5	126

Parallelism

Natural human c-Met (Total) from GTL-16 and HeLa cell lysates was serially diluted in Standard Diluent Buffer. The optical density of each dilution was plotted against the human c-Met (Total) standard curve. The standard accurately reflects c-Met content in samples.

c-Met (Total) ELISA: Parallelism



Recovery

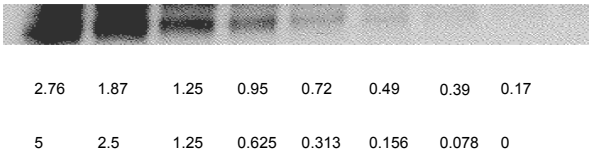
To evaluate recovery, cell extraction buffer was diluted 1:10 with Standard Diluent Buffer to bring the SDS concentration to <0.01%. human c-Met (Total) Standard was spiked into this. The average recovery was 99.9%.

Sensitivity

The analytical sensitivity of this assay is <0.4 ng/ml of human c-Met (Total). This was determined by adding two standard deviations to the mean O.D. obtained when the zero standard was assayed 30 times.

The sensitivity of this ELISA was compared to western blotting using known quantities of human c-Met (Total). The data presented below show that the sensitivity of the ELISA is approximately 2X greater than that of western blotting. The bands shown in the western blot data were developed using rabbit anti-c-Met [pYpYpY1230/1234/1235] and an alkaline phosphatase conjugated anti-rabbit IgG followed by a chemiluminescent substrate and autoradiography.

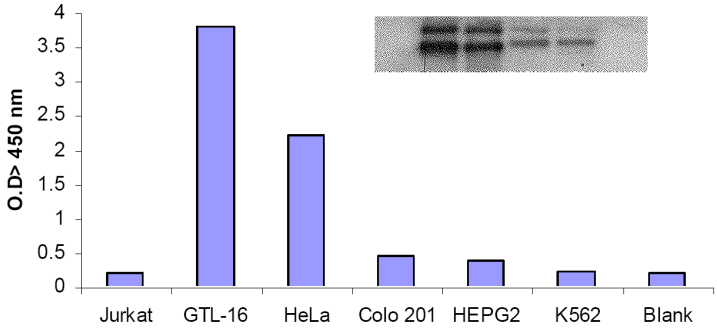
Detection of c-Met (Total) by ELISA vs Western Blot:



Specificity

The Human c-Met (Total) ELISA Kit is specific for the measurement of total c-Met. To determine the specificity of this ELISA kit, cell extracts from different cell lines, each at a concentration of 200 mg/mL total protein, were analyzed. The data presented in the figure below show that the kit detects c-Met protein in cell lysates from human Jurkat, GTL-16, HeLa, Colo 201, HEPG2, and K562 cells. The levels of c-Met protein detected with this ELISA kit are consistent with results obtained by western blot analysis (insert).

Detection of total c-Met in several cell lines



Limited product warranty

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Product label explanation of symbols and warnings

REF	Catalog Number	LOT	Batch code		Temperature limitation		Use by		Manufacturer		Consult instructions for use		Caution, consult accompanying documents

Manufacturer's address: Bender MedSystems GmbH | Campus Vienna Biocenter 2 | 1030 Vienna, Austria

The information in this guide is subject to change without notice.