



## Contents and storage

| Product   | Cat. No. 34577<br>(for 2,000 cm <sup>2</sup> ) | Cat. No. 34580<br>(for 5,000 cm <sup>2</sup> ) | Cat. No. 34578<br>(for 10,000 cm <sup>2</sup> ) |
|---|--|--|---|
| SuperSignal™ West Pico PLUS Luminol/Enhancer Solution | 100 mL   | 250 mL   | 500 mL  |
| SuperSignal™ West Pico PLUS Stable Peroxide Solution  | 100 mL   | 250 mL   | 500 mL  |
| <b>Storage: Store at room temperature.</b>            |  |  |   |



## Product description

Thermo Scientific™ SuperSignal™ West Pico PLUS Chemiluminescent Substrate is a sensitive, luminol-based enhanced chemiluminescent substrate for detecting horseradish peroxidase (HRP) on immunoblots. SuperSignal™ West Pico PLUS Chemiluminescent Substrate enables low picogram or high femtogram detection of antigen by oxidizing luminol in the presence of HRP and peroxide. This reaction produces a prolonged chemiluminescence which can be visualized on X-ray film or an imaging system. Optimal signal intensity and duration can be attained with appropriate primary and secondary antibody dilutions (see Table 1).



## Required materials

- Western blot membrane
- X-ray film or CCD imaging system (e.g., Invitrogen™ iBright™ western blot imaging systems)
- Rotary or rocking platform shaker



## Online resources

- Visit [thermofisher.com/chemisubstrates](https://thermofisher.com/chemisubstrates) for additional information and protocols.
- For support, visit [thermofisher.com/support](https://thermofisher.com/support).

**Table 1. Recommended antibody concentration**

| Primary antibody dilution<br>(from a 1 mg/mL stock) | Secondary antibody dilution<br>(from a 1 mg/mL stock) |
|---|---|
| 1:1,000–1:5,000 or<br>0.2–1 µg/mL                   | 1:20,000–1:100,000 or<br>10–50 ng/mL                  |



## Important guidelines

- Optimize your western blot procedure for best results. Variables include sample amount, gel type, transfer method, membrane type, blocking reagent, wash buffer, primary and secondary antibody concentrations, and incubation times.
- Use a sufficient volume of all solutions to ensure that membranes do not dry out.
- Use a shaking or rocking platform during incubation steps for optimal results.
- Do not use sodium azide in buffers, because it inhibits HRP.
- Always wear gloves or use clean, plastic forceps. Metallic devices (e.g., scissors) must have no visible signs of rust, which may cause speckling and/or high background.
- The substrate Working Solution is stable for 8 hours at room temperature. Avoid exposure to the sun or other intense light. Short-term exposure to lab lighting is okay.

## Important licensing information


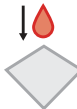







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## Perform western detection using SuperSignal™ West Pico PLUS Substrate

| Step     |   | Action   |
|----------|---|--|
| <b>1</b> |    | <b>Wash membrane</b><br>After protein transfer, remove the blot from the transfer apparatus and wash the membrane in deionized water for 5 minutes using agitation to remove all transfer buffer.  |
| <b>2</b> |    | <b>Block membrane</b><br>Block nonspecific sites with Blocking Reagent (e.g., StartingBlock™ (PBS) Blocking Buffer, Cat. No. 37538) for 30–60 minutes at room temperature with shaking. Alternatively, block overnight at 2–8°C without shaking.   |
| <b>3</b> |    | <b>Add primary antibody</b><br>Incubate the membrane with primary antibody solution (0.2–1 µg/mL or follow manufacturer's recommended dilution) containing 10% blocking solution with continuous rocking for 1 hour. If desired, incubate the blot overnight at 2–8°C.   |
| <b>4</b> |    | <b>Wash membrane</b><br>a. Wash the membrane for 10 minutes using agitation with Tris-buffered saline (TBS), phosphate-buffered saline (PBS), or other physiological wash buffer containing 0.05% Tween 20 detergent.<br>b. Repeat wash step 2 more times.<br>c. Proceed to next step, or if using an enzyme-conjugated HRP primary antibody, proceed to Step 6.   |
| <b>5</b> |    | <b>Add secondary antibody</b><br>Incubate blot with the secondary antibody HRP-conjugate working dilution (10–50 ng/mL or 1:20,000–1:100,000, from a 1 mg/mL stock solution) for 30 minutes to 1 hour at room temperature using shaking.   |
| <b>6</b> |    | <b>Wash membrane</b><br>Wash the membrane 6 times for 5 minutes each in wash buffer to remove any unbound secondary antibody conjugate. It is crucial to thoroughly wash the membrane after incubation with the HRP enzyme conjugate.  |
| <b>7</b> |  | <b>Prepare substrate</b><br>Prepare the substrate working solution by mixing equal parts of the Substrate and Stable Peroxide components (e.g., for a mini blot mix 5 mL Substrate with 5 mL Stable Peroxide). Use a sufficient volume to ensure that the blot is completely wetted with the substrate and the blot does not become dry (0.1 mL/cm <sup>2</sup> ).<br><b>Note:</b> The working solution is stable for up to 8 hours at room temperature. |
| <b>8</b> |  | <b>Develop substrate</b><br>Incubate the membrane with the substrate working solution for 5 minutes.   |
| <b>9</b> |  | <b>Image membrane</b><br>a. Remove blot from working solution and place it in a plastic sheet protector or clear plastic wrap.<br>b. Use an absorbent tissue to remove excess liquid and carefully press out any bubbles from between the blot and the membrane protector.<br>c. Image the blot using an imaging system or X-ray film.   |

## Troubleshooting

| Observation   | Cause  | Solution  |
|---|--|---|
| Reverse image on film (i.e., white bands with a black background) | Too much HRP in the system.  | Further dilute the HRP-conjugate (see guidelines in Table 1).               |
| Membrane has brown or yellow bands                                |  |   |
| Blot glows in the darkroom  |  |   |
| Signal duration is less than 8 hours                              |  |   |
| Weak or no signal   | Too much HRP in the system depleted the substrate and caused the signal to fade quickly. | Further dilute the HRP-conjugate (see guidelines in Table 1).               |
|   | Insufficient quantities of antigen or antibody.  | Increase amount of antibody or antigen.                                     |
|   |  | Use SuperSignal™ Western Blot Enhancer (Cat. No. 46640).                    |
|   | Inefficient protein transfer.  | Optimize transfer conditions.   |
| Reduction of HRP or substrate activity.                           | Test the activity of the system. <sup>[1]</sup>  |   |
| High background   | Too much HRP in the system.  | Further dilute the HRP-conjugate (see guidelines in Table 1).               |
|   | Inadequate blocking.   | Optimize blocking conditions.   |
|   | Inappropriate blocking reagent.  | Try a different blocking reagent.   |
|   | Inadequate washing.  | Increase length, number or volume of washes.                                |
|   | Film has been overexposed.   | Decrease exposure time or use Pierce Background Eliminator (Cat No. 21065). |
|   | Concentration of antigen or antibody is too high.  | Decrease amount of antigen or antibody.                                     |
|   | Poor antibody specificity.   | Use SuperSignal™ Western Blot Enhancer (Cat. No. 46640).                    |
| Spots within the protein bands                                    | Inefficient protein transfer.  | Optimize transfer conditions.   |
|   | Unevenly hydrated membrane.  | Perform manufacturer's recommendations for hydrating membrane properly.     |
|   | Bubble between the film and the membrane.  | Remove bubbles before exposing blot to film.                                |

[1] To test the activity of the system in the darkroom, prepare 1–2 mL of the SuperSignal™ Substrate Working Solution in a clear test tube. With the lights turned off, add 1 µL undiluted HRP-conjugate to the Working Solution. The solution should immediately emit a blue light that will fade over the next several minutes. If no light emission is evident, test another source of HRP to determine the root cause.

## Troubleshooting

| Observation                 | Cause  | Solution  |
|-----------------------------|--|---|
| Speckled background on film | Aggregate formation in the HRP-conjugate.        | Filter conjugate through a 0.2 µm filter.                     |
| Nonspecific bands           | Too much HRP in the system.                      | Further dilute the HRP-conjugate (see guidelines in Table 1). |
|                             | SDS caused nonspecific binding to protein bands. | Do not use SDS during the Western blot procedure.             |
|                             | Poor antibody specificity.                       | Use SuperSignal™ Western Blot Enhancer (Cat. No. 46640).      |
|                             | Insufficient blocking.                           | Increase blocking time or use different blocking reagent.     |

## Choosing the right substrate for your application

| Thermo Scientific™ Substrate | Choose when...  | Sensitivity                    | Signal Duration (hours) | Recommended Antibody Dilutions (From 1 mg/mL stock) |
|------------------------------|---|--------------------------------|-------------------------|---|
| Pierce™ ECL                  | Detecting protein targets that are in high abundance and the sample is abundant.            | Low picogram                   | 1 to 2                  | 1°: 1:1,000<br>2°: 1:1,000-1:15,000                 |
| SuperSignal™ West Pico PLUS  | Performing routine western blot or setting up new procedures that need to be optimized.     | Low picogram to high femtogram | Up to 24                | 1°: 1:1,000<br>2°: 1:20,000-1:100,000               |
| SuperSignal™ West Dura       | Performing quantitative western blots or if extended signal duration is necessary.          | Mid-femtogram                  | 24                      | 1°: 1:5,000<br>2°: 1:50,000-1:250,000               |
| SuperSignal™ West Atto       | Detecting very low abundance protein targets, or if antibodies or sample volume is limited. | Low femtogram to high attogram | 6                       | 1°: 1:5,000<br>2°: 1:100,000-1:250,000              |

## Related products

| Product  | URL   |
|--|---|
| Western blot transfer devices and membranes          | <a href="https://thermofisher.com/transfer">thermofisher.com/transfer</a>               |
| Blocking buffers, wash buffers and stripping buffers | <a href="https://thermofisher.com/westernbuffers">thermofisher.com/westernbuffers</a>   |
| Enhanced chemiluminescence (ECL) kits                | <a href="https://thermofisher.com/chemisubstrates">thermofisher.com/chemisubstrates</a> |
| Western blot antibodies                              | <a href="https://thermofisher.com/antibodies">thermofisher.com/antibodies</a>           |
| Western blot imaging and analysis                    | <a href="https://thermofisher.com/westernimaging">thermofisher.com/westernimaging</a>   |