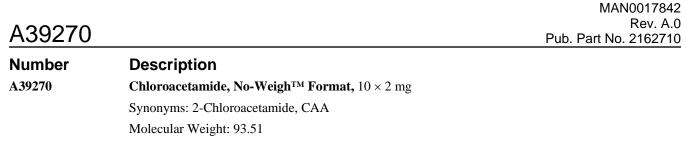
## **INSTRUCTIONS**

# Chloroacetamide, No-Weigh Format



Formula: C<sub>2</sub>H<sub>4</sub>ClNO

**Storage:** Upon receipt store product at 4°C.

**Note:** Product labels have been provided for your convenience. Please label the vials using one of the labels provided in the Al foil pouch to avoid any confusion as you work with this No-Weigh reagent.

#### Introduction

Chloroacetamide is a sulfhydryl-reactive alkylating reagent used to block reduced cysteine residues for protein characterization and peptide mapping. Like iodoacetamide, chloroacetamide modification results in the covalent addition of a carbamidomethyl group (57.07 Da) to reduced cysteine sulfhydryls and prevents the formation of disulfide bonds.<sup>1</sup> However, chloroacetamide is less reactive and more stable in solution than iodoacetamide which results in more specific cysteine modification with less off-target labeling of other amino acids and side reactions.<sup>2</sup>

### Traditional Sequential Reduction/Alkylation Procedure

**Note:** Chloroacetamide is mildly light-sensitive. Prepare solutions immediately before use and minimize exposure to light during alkylation. Although cysteine modification is the primary alkylation reaction, using too much chloroacetamide or incorrect buffer pH can result in alkylation of amines (lysine, N-termini), thioethers (methionine), imidazoles (histidine) and carboxylates (aspartate, glutamate). Dithiothreitol (DTT) can be used as an alternative reducing agent instead of TCEP for traditional reduction/alkylation but is not recommended for rapid "one-pot" reduction/alkylation.

- 1. Dissolve 5-100 µg of protein sample in 100 µL of 100 mM ammonium bicarbonate (pH 8.0) or 100 mM triethylammonium bicarbonate (pH 8.5) buffer.
- Add 1 µL of 500 mM neutral Bond-Breaker<sup>™</sup> TCEP Solution (Product No. 77720) to a final concentration of 5 mM TCEP and incubate sample at 37°C for 30 minutes.
- 3. Immediately before use, dissolve one tube of chloroacetamide (2 mg) with 107 μL of 100 mM ammonium bicarbonate (pH 8.0) or 100 mM triethylammonium bicarbonate (pH 8.5) buffer to make 200 mM chloroacetamide stock solution.
- 4. Add 11 μL of the 200 mM chloroacetamide to the sample for a final concentration of 20 mM chloroacetamide and incubate for 30 minutes at room temperature protected from light.
- 5. Perform proteolytic digestion before MS analysis or other application.

### Rapid "One-Pot" Reduction/Alkylation Procedure

- 1. Dissolve 5-100 µg of protein sample in 100 µL of 100 mM ammonium bicarbonate (pH 8.0) or 100 mM triethylammonium bicarbonate (pH 8.5) buffer.
- Add 1 µL of 500 mM neutral Bond-Breaker<sup>™</sup> TCEP Solution (Product No. 77720) to a final concentration of 5 mM TCEP.
- 3. Immediately before use, dissolve one tube of chloroacetamide (2 mg) with 107 μL of 100 mM ammonium bicarbonate (pH 8.0) or 100 mM triethylammonium bicarbonate (pH 8.5) buffer to make 200 mM chloroacetamide stock solution.



- 4. Add 11 μL of the 200 mM chloroacetamide to the sample for a final concentration of 20 mM chloroacetamide and incubate for 10 minutes at 95°C protected from light.
- 5. Perform proteolytic digestion before MS analysis or other application.

#### Troubleshooting

Problem	Possible Cause	Solution	
Sulfhydryls not blocked	Proteins not reduced	Reduce protein samples by adding DTT or TCEP before addition of chloroacetamide	
	Chloroacetamide hydrolysis	Make chloroacetamide solutions immediately before each use and dispose of excess reconstituted reagent	
Sulfhydryls partially blocked	Protein not denatured	Add 2% SDS or 8 M urea to protein sample to denature protein	
	Insufficient chloroacetamide used	Use at least a 10-fold excess of chloroacetamide to sulfhydryls	
	Incorrect reaction buffer	Avoid buffers that contain sulfhydryls or that are not at a slightly alkaline pH	
	Insufficient reaction time or temperature	Allow reaction to proceed for 10 minutes at 95°C or 30-60 minutes at room temperature	
Amines or other functional groups labeled	Incorrect pH of the reaction buffer	Maintain the reaction buffer pH at 7.5-8.5	
	Excessive chloroacetamide or incubation time	Reduce the amount of reagent or incubation time	
		Quench excess reagent by addition of excess reducing agent after alkylation	

#### **Related Thermo Scientific Products**

A39255	DTT, No-Weigh™ Format, 48 × 7.7 mg	
A39271	Iodoacetamide, No-Weigh <sup>TM</sup> Format, $30 \times 9.3$ mg	
A35349	TCEP-HCl, No-Weigh Format, 10 × 1mg	
20490	TCEP•HCl [Tris(2-carboxyethyl) phosphine hydrochloride], $1 \text{ g}$	
23011	Methyl Methanethiosulfonate (MMTS), 200 mg	
23030	<i>N</i> -Ethylmaleimide (NEM), 25 g	
77720	Bond-Breaker <sup>TM</sup> TCEP Solution, Neutral pH, 5 mL	
77712	Immobilized TCEP Disulfide Reducing Gel, 5 mL	
90055	Trypsin Endoproteinase, modified, TPCK treated, MS Grade, $5\times20~\mu g$	

#### **General References**

- 1. Gurd, F.R.N. (1967). Carboxymethylation. In Methods in Enzymology, (C.H.W. Hirs, ed.), Vol. 11, p 532. Academic Press, New York.
- 2. Müller T. and Winter D. (2017). Systematic Evaluation of Protein Reduction and Alkylation Reveals Massive Unspecific Side Effects by Iodinecontaining Reagents. *Mol Cell Proteomics* 16(7):1173-1187.

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