

## Dehydroepiandrosterone Sulfate (DHEA-S) Competitive ELISA Kit

Catalog Number EIADHEA [96 tests]

Rev 1.0

**Note:** For safety and biohazard guidelines, see the “Safety” appendix in the *ELISA Technical Guide* (Pub. no. MAN0006706). Read the Safety Data Sheets (SDSs) and follow the handling instructions. Wear appropriate protective eyewear, clothing, and gloves.

### Product description

The Dehydroepiandrosterone Sulfate (DHEA-S) ELISA Kit is a solid-phase competitive Enzyme-Linked Immunosorbent Assay (ELISA). This assay is designed to detect and quantify the level of dehydroepiandrosterone sulfate (DHEA-S) in serum, plasma, urine, saliva dried fecal extracts, and tissue culture media. The assay recognizes dehydroepiandrosterone sulfate (DHEA-S) independent of species.

### Contents and storage

Kit and components are shipped at –20°C. Upon receipt, store the kit at –20°C. Once open, store the kit at 4°C and use within 2 weeks.

Components	Quantity
Dehydroepiandrosterone Sulfate (DHEA-S) Standard; 1,200 ng/mL dehydroepiandrosterone sulfate (DHEA-S) in a special stabilizing solution	70 µL
Assay Buffer Concentrate (5X)	28 mL
Antibody Coated Wells, 96-well strip-well plate coated with donkey anti-sheep IgG	1 plate
Dehydroepiandrosterone Sulfate (DHEA-S) Antibody	3 mL
Dehydroepiandrosterone Sulfate (DHEA-S) Conjugate	3 mL
Wash Buffer Concentrate (20X)	30 mL
TMB (Tetramethylbenzidine) Substrate	11 mL
Stop Solution; contains 1 M HCl, <b>CAUSTIC</b>	5 mL
Plate Sealer	1

### Materials required but not supplied

- Distilled or deionized water
- Microtiter plate reader with software capable of measurement at or near 450 nm (preferably with correction between 570 nm and 590 nm).
- Plate washer—automated or manual (squirt bottle, manifold dispenser, or equivalent)
- Plate shaker
- Calibrated adjustable precision pipettes and glass or plastic tubes for diluting solution

### Procedural guidelines

**IMPORTANT!** Reagents are lot-specific. Do not mix or interchange different reagent lots from various kit lots.

- Review the **Procedural guidelines** and **Plate washing directions** in the *ELISA Technical Guide* available at [thermofisher.com](http://thermofisher.com).
- Allow reagents to reach room temperature before use. Mix to redissolve any precipitated salts.
- Solutions containing sodium azide will inhibit the activity of the peroxidase conjugate. Ensure that there is no contamination of labware or the plate washer with azide containing solutions.

### Prepare 1X Wash Buffer

1. Dilute 15 mL of Wash Solution Concentrate (20X) with 285 mL of deionized or distilled water. Label as 1X Wash Buffer.
2. Store the concentrate and 1X Wash Buffer in the refrigerator. Use the diluted buffer within 3 months.

### Prepare 1X Assay Buffer

1. Dilute 14 mL of Assay Buffer (5X) with 56 mL of deionized or distilled water. Label as 1X Assay Buffer.
2. Store the concentrate and 1X Assay Buffer in the refrigerator. 1X Assay Buffer is stable at 4°C for 3 months.

For research use only. Not for use in diagnostic procedures.

## Sample preparation guidelines

- Refer to the *ELISA Technical Guide* at [thermofisher.com](http://thermofisher.com) for detailed sample preparation procedures.
- Collect samples in pyrogen/endotoxin-free tubes.
- Freeze samples after collection if samples will not be tested immediately. Avoid multiple freeze-thaw cycles of frozen samples. Thaw completely and mix well (do not vortex) prior to analysis.
- Avoid the use of hemolyzed or lipemic sera.
- If large amounts of particulate matter are present in the sample, centrifuge or filter sample prior to analysis.

## Prepare samples

Sample concentrations should be within the range of the standard curve. Because conditions may vary, each investigator should determine the optimal dilution for each application.

Use all samples within **2 hours** of dilution, or store at  $-20^{\circ}\text{C}$  or lower until ready to perform assay.

Sample type	Procedure
Serum and plasma	Minimum dilution for human serum and plasma samples is 1:2, but due to the high sample concentration, most samples will have to be diluted at least 1:100 with diluted 1X Assay Buffer. For measurement of DHEA-S in non-human samples, it is recommended to carry out a preliminary dilution series to determine the correct dilution for their samples.
Urine	Dilute samples $\geq 1:2$ with 1X Assay Buffer. <b>Note:</b> A Urinary Creatinine Detection Kit (Cat. no. EIACUN) is available for measuring urine creatinine for normalization of dehydroepiandrosterone sulfate (DHEA-S) in random urine specimens.
Saliva	Dilute saliva samples at least 1:2 with diluted Assay Buffer. A saliva collection and clarification protocol is available on the product page at <a href="http://thermofisher.com">thermofisher.com</a> .
Dried feces	See detailed extraction protocol on the product page at <a href="http://thermofisher.com">thermofisher.com</a> . <b>Note:</b> The ethanol concentration in the final diluted Assay Buffer dilution added to the well should be $<5\%$ .
Tissue culture media	Perform sample dilutions with the corresponding tissue culture medium.

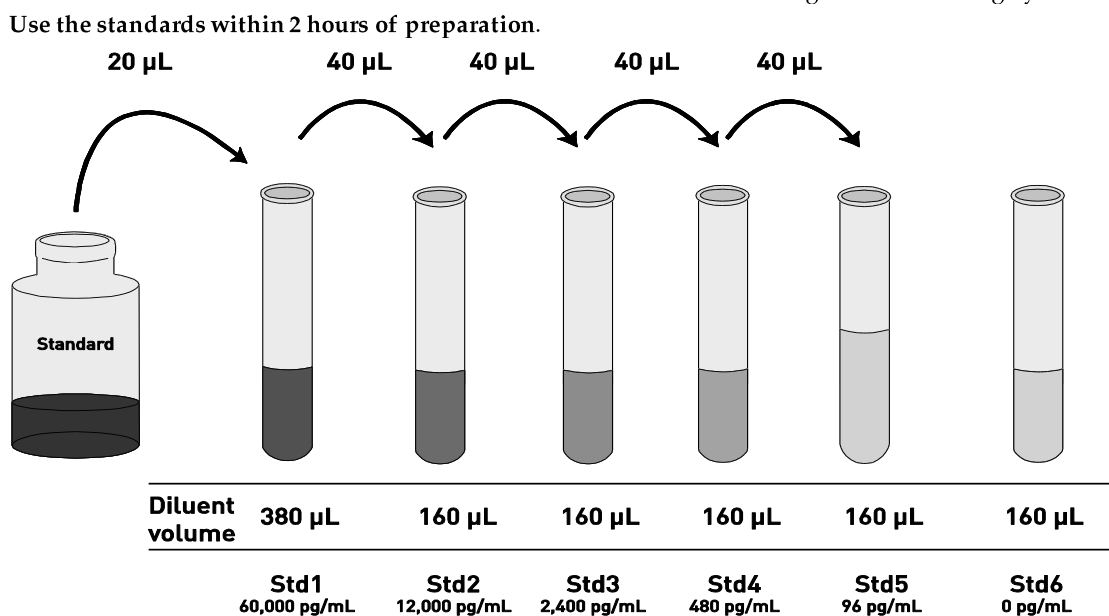
## Dilute standards

**Note:** Use glass or plastic tubes for diluting standards.

Instructions are for diluting standards from 60,000 to 96 pg/mL. Choose the range that fits your sample concentrations most appropriately.

The Dehydroepiandrosterone Sulfate (DHEA-S) Standard contains an organic solvent. Pipette the standard up and down several times to wet the pipet tip before transfer to ensure that volumes are accurate.

1. Add 20  $\mu\text{L}$  Dehydroepiandrosterone Sulfate (DHEA-S) Standard to one tube containing 380  $\mu\text{L}$  1X Assay Buffer and label as 60,000 pg/mL dehydroepiandrosterone sulfate (DHEA-S).
2. Add 160  $\mu\text{L}$  1X Assay Buffer to each of 5 tubes labeled as follows 12,000; 2,400; 480; 96; and 0 pg/mL dehydroepiandrosterone sulfate (DHEA-S).
3. Make serial dilutions of the standard as described below in the dilution diagram. Mix thoroughly between steps.



## Perform ELISA (Total assay time: 2.5 hours)

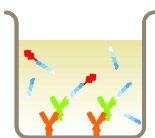
**IMPORTANT!** Perform a standard curve with each assay.

Allow all components to reach room temperature before use. Mix all liquid reagents prior to use.

Determine the number of 8-well strips required for the assay. Insert the strips in the frames for use. Re-bag any unused strips and frames, and store desiccated at 2°C to 8°C for future use. The silica pack in the bag keeps the plate dry, and turns from blue to pink if the bag is not properly sealed.

### Bind antigen

- Add 50  $\mu$ L of standards or samples (see "Prepare samples" on page 2) to the appropriate wells.
- Add 75  $\mu$ L of 1X Assay Buffer into wells for detecting non-specific binding (NSB).
- Add 25  $\mu$ L of Dehydroepiandrosterone Sulfate (DHEA-S) Conjugate to each well.
- Add 25  $\mu$ L of Dehydroepiandrosterone Sulfate (DHEA-S) Antibody to each well except NSB wells.
- Tap the side of the plate to mix. Cover the plate with plate sealer and incubate for 2 hours at room temperature with shaking.

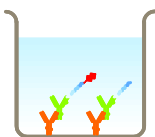


**Note:** If the plate is not shaken the bound of the signals will be ~20% lower.

- Thoroughly aspirate the solution and wash wells 4 times with 300  $\mu$ L of 1X Wash Buffer.

### Add chromogen

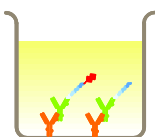
- Add 100  $\mu$ L TMB Substrate to each well. The substrate solution will begin to turn blue.
- Incubate for 30 minutes at room temperature without shaking.



**Note:** TMB should not touch aluminum foil or other metals.

### Add stop solution

Add 50  $\mu$ L Stop Solution to each well. Tap side of the plate gently to mix. The solution in the wells changes from blue to yellow.



## Read the plate and generate the standard curve

- Read the absorbance at 450 nm. Read the plate within 10 minutes after adding the Stop Solution.
- Use curve-fitting software to generate the standard curve. A four parameter algorithm provides the best standard curve fit. Optimally, the background absorbance may be subtracted from all data points, including standards, unknowns and controls, prior to plotting.
- Read the concentrations for unknown samples and controls from the standard curve. Multiply value(s) obtained for sample(s) by the appropriate factor to correct for the sample dilution.

**Note:** Dilute samples producing signals lower than that of the highest standard and reanalyze. Multiply the concentration by the appropriate dilution factor.

## Performance characteristics

### Standard curve (example)

The following data were obtained for the various standards over the range of 0–60,000 pg/mL dehydroepiandrosterone sulfate (DHEA-S).

Standard Dehydroepiandrosterone Sulfate (DHEA-S) (pg/mL)	Optical Density (450 nm)*
60,000	0.141
12,000	0.266
2,400	0.463
480	0.641
96	0.725
0	0.765

**Note:** The NSB gave a Mean OD value of 0.065.

### Intra-assay precision

Samples were assayed in replicates of 20 to determine precision within an assay.

Parameters	Sample 1	Sample 2	Sample 3
Mean (pg/mL)	8,148	3,692	920.9
%CV	4.6	6.1	10.3

CV = Coefficient of Variation

### Inter-assay precision

Samples were assayed in duplicates in 18 assay runs by four operators to determine precision between assays.

Parameters	Sample 1	Sample 2	Sample 3
Mean (pg/mL)	8,567	3,794	932.2
%CV	5.2	8.4	11.5

CV = Coefficient of Variation

## Performance characteristics, continued

### Expected values

Nineteen serum samples were tested in the assay and adjusted neat concentrations of DHEA-S ranged from 438.5 to 5,879 ng/mL with a mean of 3,177 ng/mL.

Seven plasma samples were tested in the assay and adjusted neat concentrations of DHEA-S ranged from 299.9 to 5,847 ng/mL with a mean of 2,105 ng/mL.

Nine urine samples were tested in the assay and adjusted neat concentrations of DHEA-S ranged from 371.2 to 2,555 ng/mL with a mean of 942.4 ng/mL.

Five saliva samples were tested in the assay and adjusted neat concentrations of DHEA-S ranged from 0.170 to 8.24 ng/mL with a mean of 3.93 ng/mL.

Three fecal extracts were tested in the assay and adjusted neat concentrations of DHEA-S ranged from 5.40 to 429 pg/mg with a mean of 278 pg/mg.

### Recovery

Recovery was determined using human serum and urine samples, by taking samples with a high known DHEA-S concentration and a lower DHEA-S concentration and mixing them in the ratios given below. The measured DHEA-S concentrations were compared to the expected values based on the ratios used.

High Sample %	Low Sample %	Expected Conc. (ng/mL)		Observed Conc. (ng/mL)		% Recovery	
		Serum	Urine	Serum	Urine	Serum	Urine
80	20	30.20	23.96	30.02	24.04	99.4	100.3
60	40	23.62	19.29	28.80	19.34	121.9	100.3
40	60	17.05	14.62	19.82	15.70	116.2	107.4
20	80	10.47	9.95	11.70	10.46	111.8	105.2
Mean Recovery						112.3%	103.3%

### Specificity

The following cross reactants were tested in the assay and calculated at the 50% binding point.

Steroid	Cross-reactivity %
DHEA-S	100
DHEA	162.0
Epiandrosterone	44.5
Androsterone	28.4
Androstenedione	15.2
DHT	0.5
Adrenosterone	0.4
Testosterone	0.4
Desoxycorticosterone	0.2
Progesterone	0.2
Estrone	0.1
17OH-Progesterone	0.1
17OH-Pregnenolone	<0.1
Aldosterone	<0.1
Corticosterone	<0.1
Cholesterol	<0.1
Estradiol	<0.1
Pregnenolone	<0.1

In serum, the relative level of DHEA are typically between 1 and 0.1% of the DHEA-S concentration. The cross reactivity to DHEA with the assay will contribute to an increase in measured DHEA-S concentrations of less than 2%.

### Sensitivity

The analytical sensitivity of dehydroepiandrosterone sulfate (DHEA-S) is 90.9 pg/mL. This was determined by adding two standard deviations to the mean O.D. obtained when the zero standard was assayed 18 times, and calculating the corresponding concentration.

## Limited product warranty

Life Technologies Corporation and/or its affiliate(s) warrant their products as set forth in the Life Technologies' General Terms and Conditions of Sale found on Life Technologies' website at [www.thermofisher.com/us/en/home/global/terms-and-conditions.html](http://www.thermofisher.com/us/en/home/global/terms-and-conditions.html). If you have any questions, please contact Life Technologies at [www.thermofisher.com/support](http://www.thermofisher.com/support).



**Manufacturer's address:** Life Technologies Corporation | 7335 Executive Way | Frederick, MD 21704 | USA

The information in this guide is subject to change without notice.

### DISCLAIMER

TO THE EXTENT ALLOWED BY LAW, LIFE TECHNOLOGIES AND/OR ITS AFFILIATE(S) WILL NOT BE LIABLE FOR SPECIAL, INCIDENTAL, INDIRECT, PUNITIVE, MULTIPLE, OR CONSEQUENTIAL DAMAGES IN CONNECTION WITH OR ARISING FROM THIS DOCUMENT, INCLUDING YOUR USE OF IT.

**Important Licensing Information:** These products may be covered by one or more Limited Use Label Licenses. By use of these products, you accept the terms and conditions of all applicable Limited Use Label Licenses.

**Corporate entity:** Life Technologies Corporation | Carlsbad, CA 92008 USA | Toll Free in USA 1 800 955 6288

© 2017 Thermo Fisher Scientific Inc. All rights reserved. All trademarks are the property of Thermo Fisher Scientific and its subsidiaries unless otherwise specified.

For support visit [thermofisher.com/support](http://thermofisher.com/support) or email [techsupport@lifetech.com](mailto:techsupport@lifetech.com).

thermofisher.com

17 January 2017

**ThermoFisher**  
SCIENTIFIC