

EZ-Link[®] Iodoacetyl-LC-Biotin EZ-Link Iodoacetyl-PEG₂-Biotin

21333 21334 _{0254.5}

Number

Description

21333

EZ-Link Iodoacetyl-LC-Biotin (N-Iodoacetyl-N-biotinylhexylenediamine), 50mg

Formula: $C_{18}H_{31}IN_4O_3S$

Molecular Weight: 510.43

Spacer Arm: 27.1Å

Net Mass Addition: 382.53

Solubility: > 2mg/mL in DMF for subsequent dilution in water or buffer

21334

EZ-Link Iodoacetyl-PEG₂-Biotin {(+)-Biotinyl-iodoacetamidyl-3,6-dioxaoctanediamine}, 50mg

Formula: C₁₈H₃₁IN₄O₅S

Molecular Weight: 542.43

Spacer Arm: 24.7Å

Net Mass Addition: 414.19

Solubility: ≥ 25mg/mL in water or buffer

Storage: Upon receipt store product at 4°C protected from light and moisture. Product is shipped at ambient temperature.

Introduction

The Thermo Scientific EZ-Link Iodoacetyl-LC-Biotin and Iodoacetyl-PEG₂-Biotin are long-chain, sulfhydryl-reactive biotinylation reagents. Iodoacetyl-LC-Biotin is not soluble in water and must be dissolved in an organic solvent such as DMF before further dilution in aqueous solutions. Iodoacetyl-PEG₂-Biotin has a hydrophilic polyethylene glycol (PEG) spacer arm that imparts high water-solubility to the reagent and confers added water-solubility to modified molecules.

For both reagents, reaction of the iodoacetyl group with a sulfhydryl (-SH) group is rapid and specific, especially when only a slight reagent-to-sulfhydryl molar excess is used and the reaction is performed at pH 8.3 (7.5-8.5). The reaction occurs by nucleophilic substitution of iodine with a thiol (sulfhydryl) group, resulting in a stable thioether bond (Figure 1). If thiols are unavailable on the molecule, reaction can occur with histidyl side chains at pH 6.9-7.0, but the reaction must be allowed to proceed for at least one week. Iodoacetyl reagents also will react with amino groups at pH > 10.



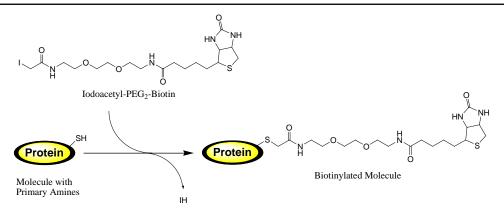


Figure 1. Biotinylation through thioether bond formation using EZ-Link Iodoacetyl-PEG₂-Biotin.

Important Product Information

- Iodoacetyl Biotin Reagents are moisture-sensitive. To avoid moisture condensation in the container, equilibrate vial to room temperature before opening. Store Iodoacetyl Biotin Reagent protected from light at 4-8°C and desiccated.
- Perform reactions in buffers that are free of thiols (sulfhydryl groups).
- Molecules to be reacted with the iodoacetyl moiety must have free (reduced) sulfhydryls. Reduce peptide disulfide bonds with Thermo Scientific Immobilized TCEP Disulfide Reducing Gel (Product No. 77712). Reduce disulfide bonds in high molecular weight proteins using 5mM TCEP (1:100 dilution of Thermo Scientific Bond-Breaker TCEP Solution, Product No. 77720) for 30 minutes at room temperature, followed by TCEP removal using a desalting column (e.g., Thermo Scientific Zeba Spin Desalting Columns). Proteins (e.g., antibodies) can be inactivated by complete reduction of their disulfide bonds. Selective reduction of hinge-region disulfide bonds in IgG can be accomplished with 2-Mercaptoethylamine•HCl (2-MEA, Product No. 20408). Sulfhydryls can be added to molecules using *N*-succinimidyl *S*-acetylthioacetate (SATA, Product No. 26102 or SAT(PEG)₄, Product No. 26099) or 2-iminothiolane•HCl (Traut's Reagent, Product No. 26101), which modify primary amines.

Reduction of IgG and Biotinylation with Iodoacetyl Biotin Reagent

The following method uses 2-mercaptoethylamine•HCl (2-MEA) as a selective and mild disulfide-cleaving reagent for reducing whole IgG in preparation for biotinylation (see Important Product Information). The protocol can be modified for other proteins, peptides and other molecules. The protein concentration during the mild reduction is not as critical as the absolute concentration of 2-MEA, which is 50mM; 1-10mg IgG/mL can be effectively reduced at this 2-MEA concentration. Generally, a 3- to 5-fold molar excess of iodoacetyl biotin reagent to sulfhydryl groups is sufficient to obtain efficient modification. Specific applications will require optimization of reducing or sulfhydryl addition steps and amount of biotinylation reagent.

Materials Required

- Sample Preparation Buffer: 0.1M sodium phosphate, 5mM EDTA, pH 6.0
- 1mL of 4mg/mL (27μM) IgG in Sample Preparation Buffer
- 2-Mercaptoethylamine•HCl (2-MEA), Product No. 20408
- Reaction Buffer: 50mM Tris•HCl, 5mM EDTA, pH 8.0-8.3
- Desalting column: e.g., Dextran Desalting Columns (Product No. 43230)

A. Prepare Reduced IgG

- 1. Add 1mL of the IgG solution to the vial containing the 6mg 2-MEA (results in 50mM 2-MEA).
- 2. Mix and incubate the solution for 90 minutes at 37°C.
- 3. Allow the solution to cool to room temperature. Remove the excess 2-MEA from the reduced IgG using a desalting column equilibrated with Reaction Buffer.



B. Biotinylate Reduced IgG With Iodoacetyl Biotin Reagent

- 1. Immediately before use, prepare 4mM solution of Iodoacetyl Biotin Reagent:
 - Dissolve 2mg Iodoacetyl-LC-Biotin in 1mL DMF.
 - Dissolve 2.2mg Iodoacetyl-PEG₂-Biotin in 1mL of Reaction Buffer.
- 2. Add 50μL of the Iodoacetyl Biotin solution per milliliter of the reduced IgG. (This results in 200μM Iodoacetyl Biotin per 50μM reduced hinge-region sulfhydryl groups, corresponding to a 4-fold excess of Iodoacetyl Biotin Reagent.)
- 3. Mix and incubate reaction in the dark for 90 minutes at room temperature.

Note: Performing the reaction in the dark limits conversion of liberated iodide ions to molecular iodine, which can react with tyrosine residues.⁵

4. Remove non-reacted Biotin Reagent by applying mixture to a desalting column that has been equilibrated with Reaction Buffer. Collect 0.5mL fractions and monitor for the presence of protein by measuring the absorbance at 280nm. The first absorption peak emerging from the column corresponds to fractions containing the biotinylated IgG. Alternatively, the non-reacted Biotin Reagent may be removed by dialysis.

Related Thermo Scientific Products

20357 High Capacity Streptavidin Agarose Resin, 2mL

20290 DTT, 5g

20408 2-Mercaptoethylamine•HCl (2-MEA), 6×6 mg

20409 TCEP•HCl, 1g

77712 Immobilized TCEP Disulfide Reducing Gel, 5mL

26101 Traut's Reagent, 500mg

26102 SATA, 50mg

Cited References

- 1. Hermanson, G.T. (1996). Bioconjugate Techniques, Academic Press. (Available from Pierce as Product No. 20002).
- 2. Savage, D.M. et.al. (1992). Avidin-Biotin Chemistry: A Handbook. Pierce Chemical Co., Rockford, IL.
- 3. Gurd, F.R.N. (1967). Carboxymethylation. Meth Enzmol XI, 532-541.
- 4. Yoshitake, S., et al. (1979). Conjugation of glucose oxidase from Aspergillus niger and rabbit antibodies using N-hydroxysuccinimide ester of N-(4-carboxycyclohexyl-methyl)-maleimide. Eur J Biochem 101:395-9.
- 5. Crestfield, A.M., et al. (1963). The preparation and enzymatic hydrolysis of reduced and S-carboxymethylated proteins. J Biol Chem 238(2):622-7.

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