INSTRUCTIONS



Protein A UltraLink® Resin

53139 53142 0508.4

Number Description

Protein A UltraLink Resin, 5mL settled resin (10mL total volume)

Binding Capacity: ≥ 16mg human IgG/mL resin

Supplied: 50% aqueous slurry containing 0.02% sodium azide

53142 Protein A Plus UltraLink Resin, 5mL settled resin (10mL total volume)

Binding Capacity: ≥ 30mg human IgG/mL resin

Supplied: 50% aqueous slurry containing 0.02% sodium azide

Storage: Upon receipt store products at 4°C. Products are shipped at ambient temperature.

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Introduction

The Thermo Scientific Protein A UltraLink Resin is used for the purification of IgG by gravity-flow, spin-column or medium-pressure chromatography methods. To purify IgG, the sample is added to the immobilized Protein A in a buffer that facilitates binding. After non-IgG components of the sample are washed from the resin, the bound IgG is recovered by elution.

Protein A is a cell wall component produced by several strains of *Staphylococcus aureus* that consists of a single polypeptide chain (MW ~ 46,700; MW by SDS-PAGE ~ 42,000) and contains little or no carbohydrate. Protein A has four high affinity $(K_a=10^8/M)$ binding sites capable of binding specifically to the Fc region of immunoglobulin molecules from several species. The Protein A molecule is heat-stable and retains its native conformation when exposed to denaturing reagents.

Covalently immobilized Protein A matrices have been extensively used to purify IgG from several species of mammals. ⁵⁻⁷ However, the interaction between Protein A and IgG is not equivalent for all species. ^{8,9} Even within a species, Protein A interacts with some subgroups of IgG and not others. For instance, human IgG_1 , IgG_2 and IgG_4 bind strongly, whereas IgG_3 does not bind and mouse IgG_1 binds poorly to Protein A. ¹⁰ There are also many instances in which monoclonal antibodies do not bind to Protein A, such as the majority of rat immunoglobulins. When using samples from species not well referenced in the literature, preliminary experiments should be conducted to determine if a particular immunoglobulin binds.

The UltraLink Biosupport is an azlactone-activated support that is hydrophilic, charge-free, high capacity, highly crosslinked, rigid, copolymeric and porous. This support is especially useful for medium pressure techniques when using large sample volumes requiring fast-flow techniques (FPLC) and large-scale applications. Agarose supports are useful for gravity-flow procedures; however, more rigid UltraLink Biosupport is required if flow rates require pressures greater than 25psi. More specific information regarding this support is detailed in the Additional Information Section.



Important Product Information

- Thermo Scientific Pierce Buffers provide the highest efficiency of IgG binding and elution for most species. Using other buffer formulations may significantly alter the binding capacity and the wash volumes required for efficient purification. Therefore, optimization may be necessary when using other buffers.
- For optimal recovery, use a sample size such that the expected IgG load on the column is less than 80% of the binding capacity (see Product Description on page 1 for binding capacities). The total IgG content of serum is approximately 10-15mg/mL. The concentration of antibody in tissue culture supernatant varies considerably among hybridoma clones.
- Antibodies from fetal bovine serum (FBS) culture media supplement will be purified along with the antibody of interest.

Procedure for Antibody Purification Using a Gravity-flow Column

Note: The following protocol is for using a gravity-flow column packed with 2mL of settled resin (i.e., 2mL of the 50% slurry). When using columns containing other resin volumes, reagent amounts must be adjusted accordingly.

A. Additional Materials Required

- Column capable of containing at least 2mL resin bed volume such as the Disposable Polypropylene Columns (Product No. 29922) or the Column Trial Pack (Product No. 29925), which contains two each of three column sizes
- Binding Buffer: Protein A IgG Binding Buffer (Product No. 21001 or 21007)
- Elution Buffer: IgG Elution Buffer (Product No. 21004 and 21009) or 0.1M glycine, pH 2-3
- Neutralization Buffer: Prepare 1mL of high-ionic strength alkaline buffer such as 1M phosphate or 1M Tris, (pH 7.5-9)

(Optional): Thermo Scientific Slide-A-Lyzer Dialysis Cassette or ZebaTM Spin Desalting Columns (Product No. 89893) for buffer exchange

B. Immunoglobulin Purification Procedure

- 1. Equilibrate all components to room temperature before use.
- 2. Carefully pack the column with 2mL of Protein A UltraLink Resin slurry according to the packing instructions provided with the columns.
- 3. Equilibrate column by adding 5mL of the Binding Buffer and allowing the solution to drain through.
 - **Note:** To avoid air bubbles being drawn into the resin, remove the top cap before the bottom cap when opening column.
- 4. To maintain optimal ionic strength for binding, dilute sample at least 1:1 with Binding Buffer.
 - **Note:** Plasma will become hazy upon dilution with the Binding Buffer because of lipoprotein precipitation. Centrifuge the diluted sample at $10,000 \times g$ for 20 minutes and apply the supernatant to the equilibrated column.
- 5. Apply diluted sample to the column and allow it to flow completely into the resin. Do not allow the resin bed to run dry. Any volume may be applied provided the total amount of antibody is less than 80% of column capacity.
 - **Note:** If the sample contains more IgG than can bind to the Protein A column (or is an antibody type that does not bind to Protein A), the flow-through will contain excess antibody. By saving the flow-through, non-bound antibody can be recovered and examined by antibody-specific assays.
- 6. Wash the column with 10-15mL of Binding Buffer.
 - **Not**e: If desired, verify that all non-bound proteins are removed from the column by collecting separate 2mL fractions and measuring their absorbance at 280nm. The last fractions should have absorbances similar to Binding Buffer alone.
- Elute antibodies with 5mL of Elution Buffer and collect 0.5-1mL fractions. Immediately adjust eluted fractions to
 physiologic pH by adding 100μL of the Neutralization Buffer to 1mL of eluate. Monitor the elution by measuring the
 absorbance at 280nm or by protein assay such as the Thermo Scientific Pierce BCA Protein Assay Kit (Product No.
 23225).
- 8. Pool the eluted antibody fractions that have the highest absorbance. The purified antibodies may be used directly for SDS-PAGE, or the buffer may be exchanged by dialysis or desalting column to one that is compatible with the specific downstream application (see Related Thermo Scientific Products).



- 9. Regenerate column by washing with 8-15mL of Elution Buffer. Columns can be regenerated at least 10 times without significant loss of binding capacity.
- 10. For storage, wash column with 5mL of water containing 0.02% sodium azide. When approximately 3mL of solution remains above the top disc, replace the bottom cap followed by the top cap on the column. Store column upright at 4°C.

Example Immunoprecipitation (IP) Procedure using Protein A Resin

A. Additional Materials Required

- 1.5-2mL microcentrifuge tube
- IP Buffer: 25mM Tris, 150mM NaCl; pH 7.2 (Thermo Scientific BupH Tris Buffered Saline Pack, Product No. 28379)
- Antigen Sample: Antigen-containing lysate or sample prepared in IP Buffer or other buffer that is compatible with both
 the desired antibody binding interaction and the binding of antibody to Protein A
- Elution Buffer: IgG Elution Buffer (Product No. 21004) or 0.1-0.2M glycine•HCl buffer, pH 2.5-3.0
- Electrophoresis Loading Buffer: Lane Marker Reducing Sample Buffer (5X), (Product No. 39000)
- Neutralization Buffer (optional): 1mL of strong alkaline buffer, such as 1M phosphate or 1M Tris, (pH 7.5-9)

B. Immunoprecipitation Procedure

Note: This procedure uses 50µL of settled Protein A resin (100µL resin slurry). This amount of resin is sufficient to bind 25-250µg of antibody. Depending on the amount of antibody needed to immunoprecipitate the desired amount of antigen, scale the amount of resin and suggested wash and elution volumes accordingly. To allow for proper mixing, make sure the total reaction volume does not completely fill the microcentrifuge tube.

- 1. In a microcentrifuge tube, combine $50\text{-}1000\mu\text{L}$ of the Antigen Sample and the optimized amount of antibody. Incubate the reaction overnight at 4°C .
- 2. Add 100µL of Immobilized Protein A resin slurry to the antigen-antibody complex.
- 3. Incubate reaction with gentle mixing for 2 hours at room temperature.
- 4. Add 0.5mL of IP Buffer, centrifuge for 2-3 minutes at $2500 \times g$ and discard supernatant. Repeat this step several times.
- 5. To elute the immune complex, add 50μ L of Elution Buffer and incubate for 5 minutes. Centrifuge tube for 1-3 minutes at $2500 \times g$ and collect the supernatant. Repeat this step and combine the two supernatant fractions.
 - Alternatively, wash the complex-bound resin with 0.5mL water, centrifuge for 2-3 minutes at $2500 \times g$, and discard supernatant. Add Electrophoresis Loading Buffer to the complex-bound resin and incubate for 5 minutes at 95° C. Centrifuge the resin mixture at $2500 \times g$, collect the supernatant and evaluate by SDS-PAGE.
- 6. Adjust eluate to physiological pH by adding ~10μL of the Neutralization Buffer per 100μL of eluate. The IP products may be used directly for SDS-PAGE, or the buffer may be exchanged by dialysis or desalting column to one that is compatible with the specific downstream application (see Related Thermo Scientific Products).



Troubleshooting

Problem	Possible Cause	Solution
Column flow is exceedingly slow (i.e., < 0.5mL/minute)	Outgassing of buffers or sample on the column resulted in blockage of resin pores with microscopic air bubbles	Degas buffers and remove air bubbles from column (See the Additional Information section for suggested Tech Tip protocol)
Considerable antibody purified, but no specific antibody of interest detected	Antibody of interest was at a very low concentration	Use serum-free medium for cell supernatant samples
		Affinity purify the antibody using the specific antigen coupled to an a affinity support such as Thermo Scientific AminoLink Plus Immobilization Kit (Product No. 44894)
Antibody of interest is degraded (as determined by lack of function in downstream assay)	Antibody was sensitive to low-pH Elution Buffer	Use Thermo Scientific Gentle Ag/Ab Elution Buffer (See Related Thermo Scientific Products)
	Downstream application was sensitive to neutralized Elution Buffer	Desalt or dialyze eluted sample into suitable buffer
No antibody detected in any elution fraction	Sample was devoid of antibody species or subclass that binds to Protein A	Refer to the Binding Characteristics Table for Protein A (See Additional Information section)

Additional Information

A. Specific Physical Characteristics of the UltraLink Biosupport

The UltraLink Biosupport is an azlactone-activated support that is hydrophilic, charge-free, high capacity, highly crosslinked, rigid, copolymeric and porous (Table 1). The support characteristics are important considerations when using large sample volumes requiring fast-flow techniques and large-scale applications. The UltraLink Biosupport is useful for medium pressure techniques such as FPLC.

Table 1. Characteristics of the UltraLink Biosupport.

Support pH Stability: 1-13

Average Particle Size: 50-80 microns

Exclusion Limit: > 2,000,000 daltons **Average Surface Area:** $> 250\text{m}^2/\text{g}$ of beads

Average Pore Volume: > 1.2mL/g of beads (> 60% of bead volume)

Pore Size: 1000Å

Maximum Pressure: 100psi (6.9bar)* **Maximum Linear Velocity:** 3000cm/hour

B. Calculating the Linear Flow Rate for Medium Pressure Chromatography

An important factor for success when performing medium pressure chromatographic (MPC) applications is limiting the pressure drop across the column, which is critical when attempting to increase scale by using a larger column. The indicated gauge pressure of an MPC apparatus may not actually measure the pressure drop across the column. Therefore, a more reliable criterion for MPC applications is to measure the linear flow rate of buffers through the column, which is a pressure-independent measurement. The linear flow rate is defined as the velocity of the buffer front passing through the resin bed and is usually expressed in cm/hour. UltraLink Biosupport has a maximum linear flow rate of approximately 3000cm/hour.

^{*}This value refers to the maximum pressure drop across a column that the resin can withstand. The indicated gauge pressure of a liquid chromatography apparatus may not be measuring the pressure drop across the column.



The linear flow rate through a cylindrical column can be calculated if the height of the resin bed and the inside diameter (or inside radius) of the column is known, and if column effluent is collected and measured for a given time. The calculations for determining linear velocity are shown below.

Calculations:

- r = Radius (cm)
- πr^2 = Column cross-sectional area
- $1 \text{cm}^3 = 1 \text{mL of buffer}$
- cm³/minute = Measured flow rate per minute (i.e., milliliter of effluent collected in 1 minute)

$$Linear\ velocity/minute = \frac{cm^3/minute}{\pi r^2}$$

Linear velocity/hour = (linear velocity/minute)(60 min/hr)

Therefore,

Linear velocity (cm/hr) =
$$\frac{\text{(cm}^3/\text{min)}(60 \text{ min/hr})}{\pi \text{r}^2}$$

C. Information Available from Our Website

- Tech Tip #34: Binding characteristics of Protein A, Protein G, Protein A/G and Protein L
- Tech Tip #4: Batch and spin cup methods for affinity purification of proteins
- Tech Tip #13: Pack beaded affinity resin into columns
- Tech Tip #7: Remove air bubbles from columns to restore flow rate
- Tech Tip #29: Degas buffers for use in affinity and gel filtration columns
- Tech Tip #43: Protein stability and storage

Related Thermo Scientific Products

66382	Slide-A-Lyzer® Dialysis Cassette Kit
69576	Slide-A-Lyzer MINI Dialysis Device Kit, 10K MWCO, 0.1mL
89893	Zeba Spin Desalting Columns, 7K MWCO, 10mL, 5/pkg
89882	Zeba Spin Desalting Columns, 7K MWCO, 0.5mL, 25/pkg
21027	Gentle Ag/Ab Elution Buffer, pH 6.6, 500mL
44894	AminoLink® Plus Immobilization Kit
37503	Pierce® Rapid ELISA Mouse mAb Isotyping Kit
32400	Recombinant Protein A, Peroxidase Conjugated, 1mg
26147	Pierce Crosslink IP Kit

Cited References

- 1. Bjork, L., et al. (1972). Some physicochemical properties of protein A from Staphylococcus aureus. Eur J Biochem 29:579-84.
- 2. Sjoquist, J., et al. (1972). Protein A isolated from Staphylococcus aureus after digestion with lysostaphin. Eur J Biochem 29:572-8.
- 3. Hjelm, H., et al. (1975). Immunologically active and structurally similar fragments of protein A from Staphylococcus aureus. Eur J Biochem 57:395-403.
- 4. Sjoholm, I. (1975). Protein A from Staphylococcus aureus: spectropolarimetric and spectrophotometric studies. Eur J Biochem 51:55-61.



- 5. Goding, J.W. (1978). Use of Staphylococcal protein A as an immunological reagent. J Immunol Methods 20:241-53.
- 6. Lindmark, R., et al. (1983). Binding of Immunoglobulins to protein A and immunoglobulin levels in mammalian sera. J Immunol Methods 62:1-13.
- 7. Kronvall, G. (1970). Phylogenetic insight into evolution of mammalian Fc fragment of gamma G globulin using staphylococcal protein A. *J Immunol* **104:**140-7.
- 8. Kronvall, G. and Williams, R.C. (1969). Differences in anti-protein A activity among IgG subgroups J Immunol 103(4):828-33.
- 9. Surolia, A., et al. (1982). Protein A: Nature's universal anti-body. TIBS 7:74-6.
- 10. Kronvall, G., et al. (1970). Protein A reactivity with mouse immunoglobulins. J Immunol 105:1116-23.
- 11. Ey, P.L., et al. (1978). Isolation of pure IgG₁, IgG_{2a}, and IgG_{2b} immunoglobulins from mouse serum using protein A-Sepharose. *Immunochem* **15**:429-36.

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