QUICK REFERENCE CARD

ChargeSwitch® gDNA Micro Tissue Kit

Catalog no. CS11203

25-0818 Version B; 18 July 2008

Follow the steps below to purify up to **5 µg** of genomic DNA from micro-dissected samples. For more detailed protocols and additional information, refer to the manual.

1. Before Starting

- ❑ 1. Set water bath at 55°C.
- ❑ 2. Prepare Lysis Mix – 1 ml Lysis Buffer (L15) and 10 µl Proteinase K per sample.

2. Preparing the Tissue Lysate

- ❑ 1. Micro dissect 3-5 mg tissue sample or mouse ear clip.
- ❑ 2. Add 1 ml Lysis Mix from Step 2 above.
- ❑ 3. Incubate at 55°C overnight or 1-2 hours with vortexing.*
- ❑ 4. Remove from incubation and add 5 µl of RNase A.
- ❑ 5. Pipet gently up and down until homogenous.
- ❑ 6. Incubate at room temperature for 5 minutes.

3. Binding the DNA

- ❑ 1. Vortex the ChargeSwitch® Magnetic Beads to resuspend.
- ❑ 2. Add 200 µl of Purification Buffer (N5) to sample and pipet gently up and down twice.
- ❑ 3. Add 40 µl of Magnetic Beads and pipet gently up and down five times.
- ❑ 4. Incubate at room temperature for 1 minute.

Binding the DNA, continued

- ❑ 5. Place the tube in the MagnaRack™ for 1 minute.
- ❑ 6. Carefully remove supernatant and discard.

4. Washing the Beads

- ❑ 1. Remove the tube from the MagnaRack™.
- ❑ 2. Add 1 ml of Wash Buffer (W12) and pipet gently up and down twice to resuspend beads.
- ❑ 3. Place the tube in the MagnaRack™ for 1 minute.
- ❑ 4. Remove the supernatant with a pipette and discard.
- ❑ 5. Repeat steps 1–4.

5. Eluting the DNA

- ❑ 1. Remove the tube from the MagnaRack™, and add 150 µl of Elution Buffer (E5).
- ❑ 2. Pipet gently up and down 10 times to resuspend beads.
- ❑ 3. Incubate for 5 minutes at room temperature.
- ❑ 4. Place the tube in the MagnaRack™ for 1 minute.
- ❑ 5. Remove the eluate containing purified DNA.

*Depending on tissue type. See detailed protocol in the manual.

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QUICK REFERENCE CARD

ChargeSwitch® gDNA Mini Tissue Kit

Catalog no. CS11204

25-0818 Version B; 18 July 2008

Follow the steps below to purify up to **30 µg** of genomic DNA from small amounts of tissue. For more detailed protocols and additional information, refer to the manual.

1. Preparing the Sample

- ❑ 1. Set water bath at 55°C.
- ❑ 2. Dissect 0.5 cm tail tip or 20-25 mg tissue sample. For tail tips, place sample in a 1 ml homogenizer.
- ❑ 3. Add 1 ml Lysis Buffer (L13). For tail tips, thoroughly homogenize and transfer to 1.5 ml microcentrifuge tube.
- ❑ 4. Add 20 µl of Proteinase K and vortex or invert briefly to mix.
- ❑ 5. Incubate at 55°C for 1.5-3 hours or until digested.
- ❑ 6. Remove from water bath, and centrifuge if necessary.
- ❑ 7. Add 10 µl of RNase A, and invert tube to mix.
- ❑ 8. Incubate at room temperature for 2 minutes.

3. Binding the DNA

- ❑ 1. Vortex the ChargeSwitch® Magnetic Beads to resuspend.
- ❑ 2. Add 120 µl of Magnetic Beads and pipet gently up and down five times.
- ❑ 3. Add 100 µl of Purification Buffer (N5).
- ❑ 4. Pipet gently up and down 10 times.
- ❑ 5. Place tubes in the MagnaRack™ for 2 minutes.

Binding the DNA, continued

- ❑ 6. Carefully remove supernatant and discard.

4. Washing the Beads

- ❑ 1. Remove the tube from the MagnaRack™.
- ❑ 2. Add 1 ml of Wash Buffer (W12) and pipet gently up and down twice to resuspend beads.
- ❑ 3. Place the tube in the MagnaRack™ for 1 minute or until clear.
- ❑ 4. Remove the supernatant with a pipette and discard.
- ❑ 5. Repeat steps 1–4 one more time.

5. Eluting the DNA

- ❑ 1. Remove the tube from the MagnaRack™.
- ❑ 2. Add 250 µl of Elution Buffer (E5).
- ❑ 3. Pipet gently up and down twice to resuspend beads.
- ❑ 4. Incubate for 5 minutes at 55°C.
- ❑ 5. Place the tube in the MagnaRack™ for 2 minutes.
- ❑ 6. Remove the eluate containing purified DNA.

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