

# MagMAX™ Microbiome Ultra Nucleic Acid Isolation Kit

High throughput isolation of Nucleic Acid (RNA and DNA) from soil, biofluids, and other samples

**Catalog Numbers** A42357 (with plate), A42358 (with tubes)

**Pub. No.** MAN0018070 **Rev.** C.0

**WARNING!** Read the Safety Data Sheets (SDSs) and follow the handling instructions. Wear appropriate protective eyewear, clothing, and gloves. Safety Data Sheets (SDSs) are available from [thermofisher.com/support](http://thermofisher.com/support).

## Product description

The Applied Biosystems™ MagMAX™ Microbiome Ultra Nucleic Acid Isolation Kit is developed for scalable, rapid purification of high-quality Total Nucleic Acid (RNA and DNA) from soil, swabs, and liquid samples (e.g. biofluids). You can use the nucleic acid purified with this kit in a broad range of molecular biology downstream applications, such as sequencing and real-time PCR. This protocol guides users through automated isolation of RNA and DNA from soil and liquid samples using the KingFisher™ Flex and the KingFisher™ Duo Prime.

## Contents and storage

Reagents that are provided in the kit are sufficient for 100 reactions (Cat. No. A42357 and A42358).

**Table 1** Components of MagMAX™ Microbiome Ultra Nucleic Acid Isolation Kit

Component	Amount	Storage
Lysis Buffer	80 mL	15°C to 25°C
Binding Solution	50 mL	
Wash Buffer	200 mL	
Elution Solution	20 mL	
Proteinase K	4 mL	
Total Nucleic Acid Binding Beads	2 mL	
Bead Tubes OR 96DW Bead Plate	100 tubes OR 1 plate	

For bulk reagents, use Cat. No. A42361 (Lysis Solution), A42359 (Binding Solution), A42360 (Wash Solution), A42364 (Elution Solution), A42363 (Proteinase K), and A42362 (Binding Beads).

For bead tubes and plate sold separately, use Cat. No. A42351 (Bead tubes, 100), and A42331 (Bead plate, 1).

## Required materials not supplied

Unless otherwise indicated, all materials are available through [thermofisher.com](http://thermofisher.com). MLS: Fisher Scientific ([fisherscientific.com](http://fisherscientific.com)) or other major laboratory supplier.

Item	Source
<b>Magnetic particle processor (one of the following, depending on quantity/volume of sample to be processed):</b>	
KingFisher™ Flex Magnetic Particle Processor with 96 Deep-Well Head	5400630
KingFisher™ Duo Prime Magnetic Particle Processor	5400110
<b>Equipment</b>	
Adjustable micropipettors	MLS
Multi-channel micropipettors	MLS
Vortex with Vortex adaptor capable of holding 2.0 mL Bead-beating tubes	MLS
Bead-beating instrument for 96-well plates (e.g. OMNI bead-ruptor)	MLS
Tube Centrifuge capable of reaching 14,000 × <i>g</i> and holding 2 mL tubes (e.g. Sorvall Legend Micro 21 Centrifuge, Ventilated; Thermofisher.com 75002435)	MLS
Plate Centrifuge capable of reaching 2,250 × <i>g</i> and holding 96DW plates (e.g. Thermo Scientific™ Sorvall™ Legend™ XT/XF with M20 rotor)	MLS
<b>Consumables</b>	
<b>Deep-well plates:</b>	
KingFisher™ Deepwell 96 Plate	95040450
<b>96-well standard plates (for use with KingFisher™ Flex only; tip comb placement and elution plate and/or eluate storage):</b>	
KingFisher™ 96 KF microplate	97002540
<b>Tip comb, compatible with the magnetic particle processor used:</b>	
KingFisher™ Duo Prime 12-tip comb, for use with KingFisher™ deep-well 96 plate	97003500
KingFisher™ 96 tip comb for DW magnets, for Flex protocol only	97002534
Elution Strip, for Duo protocol only	97003520
KingFisher™ Duo Cap for Elution Strip, for Duo protocol only	97003540

Item	Source
<b>Materials</b>	
MicroAmp™ Clear Adhesive Film	4306311
Conical Tubes (15 mL)	AM12500
Conical Tubes (50 mL)	AM12501
Reagent reservoirs	MLS
Foil Seals	14-222-3 42
Nonstick, RNase-Free Microfuge Tubes, 1.5 mL	AM12450
Nonstick, RNase-Free Microfuge Tubes, 2.0 mL	AM12475
<b>Reagents</b>	
Ethanol, 100% (molecular biology grade)	MLS
Nuclease-free Water	AM9932

## General guidelines

- Perform all steps at room temperature (20–25°C), unless otherwise noted.
- Clean the work surfaces with RNA Zap to remove nucleases, then wipe the surfaces with 70% to 100% molecular biology grade ethanol to remove additional contaminants.
- Precipitates can occur if the Lysis Buffer or Binding Solution is stored when room temperature is too cold. If there are precipitates, warm the reagents at 37°C and gently mix to dissolve the precipitates. Avoid creating bubbles.
- *For soil samples only:* In rare instances, if some inhibitors still remain within the purified DNA/RNA, the sample can be diluted 20x with nuclease-free water prior to downstream PCR or other reactions.

## Perform Nucleic Acid purification using Bead tubes and the KingFisher™ Flex

### 1 Set up the instrument

- a. Ensure that the instrument is set up with the proper magnetic head and the proper heat block, as indicated in the following table.

Component	Type
Magnetic head	96 deep-well magnetic head
Heat block	96 well deep-well heat block

**IMPORTANT!** Failure to use the proper magnetic head and heat block results in lower yields and potential harm to the instrument.

- b. Ensure that the proper program has been downloaded from the product page and loaded onto the instrument:
- Soil Samples: **MagMAX\_Microbiome\_Soil\_Flex**
  - Biofluid and other sample types: **MagMAX\_Microbiome\_Liquid\_Buccal\_Flex**

### 2 Lyse sample

- a. Set up the Vortex with the vortex adaptor.
- b. Add 800 µL of Lysis Buffer to the bead tubes.

## Guidelines for Binding Bead Mix

- Vortex Binding Beads thoroughly before each use.
- Ensure that the beads stay fully mixed within the solution during pipetting.
- Avoid creating bubbles during mixing and aliquoting.
- Binding/Bead Mix is very viscous so pipet with care to ensure that the correct volume is added to the sample.

## Before first use of the kit

- Prepare 80% Ethanol from 100% absolute Ethanol and Nuclease-Free Water.  
Prepare enough solution for a minimum volume of 2 mL per sample.

## Before each use of the kit

1. Vortex Beads vigorously to ensure they are homogenous.
2. Prepare Binding/Bead Mix according to the following table:

Component	Volume per well <sup>[1]</sup>
Total Nucleic Acid Binding Buffer	500 µL
Total Nucleic Acid Magnetic Beads	20 µL
<b>Total volume</b>	<b>520 µL</b>

<sup>[1]</sup> Use 10% Overage calculation when making a master mix for use with multiple samples.

3. Mix well by inversion, then store at room temperature.

## 2 Lyse sample (continued)

- c. Prepare samples according to the following table:

Sample type	Description
Soil Samples	Weigh out 200–250 mg, then place into prepared bead tube
Biofluid or Liquid sample	Remove 400–500 µL, then place into prepared bead tube
Skin/Buccal Swab	Remove the plastic stick, then place the swab into prepared bead tube
Bacterial/Yeast Culture	Remove 500 µL of the culture, then place into prepared bead tube

**Note:** For bacterial and Yeast cultures, and urine, user can centrifuge 1 ml then re-suspend the cell pellet in 800 µL of Lysis Buffer and transfer to the bead tube to start the extraction.

- d. Vortex the bead tube upside down for 10 seconds to mix the sample with the buffer.  
e. Set the speed to 2,500 rpm, then place the tubes onto the vortex adaptor.

**Note:** We recommend the VWR 24 tube adaptor and vortex for maximum nucleic acid yield. If an alternative instrument or adaptor is used, ensure that the instrument can meet the speed listed in the protocol.

- f. Lyse samples on the vortexer for 10 minutes.

STOPPING POINT Bead tube can be stored at 4°C overnight after lysis.

## 3 Set up the processing plates

Set up the Wash, Elution, and Tip Comb Plates outside the instrument according to the following table.

Plate ID	Plate position	Plate type	Reagent	Volume per well
Wash 1 Plate	2	Deep-well	Wash Buffer	1,000 µL
Wash 2 Plate	3	Deep-well	Wash Buffer	1,000 µL
Wash 3 Plate	4	Deep-well	80% Ethanol	1,000 µL
Wash 4 Plate	5	Deep-well	80% Ethanol	1,000 µL
Elution Plate	6	Deep-well	Elution Solution	50 µL
Tip Comb	7	Place a 96 Deep-well Tip Comb in a Standard deep-well Plate		

**Note:** To prevent evaporation and contamination, cover the prepared processing plates with paraffin film or MicroAmp™ Clear Adhesive Film until they are loaded into the instrument.

## 4 Digest with Proteinase K

- a. Remove tubes from vortexer, then centrifuge for 2 minutes at 14,000 × g.  
b. Transfer 400–500 µL of the Sample to the appropriate wells of a new deep-well plate. This plate is the Sample Plate.  
**Note:** For soil samples, transfer as much supernatant as possible without any particle carryover.  
c. Add 40 µL of Proteinase K to each sample.  
d. Select the correct program on the instrument:
  - Soil Samples: **MagMAX\_Microbiome\_Soil\_Flex**
  - Biofluid and other sample types: **MagMAX\_Microbiome\_Liquid\_Buccal\_Flex**
e. Start the run, then load the prepared plates into position when prompted by the instrument.

- 5 Bind, wash, then elute total nucleic acid**
- When prompted (approximately 20 minutes after the start of the protocol), remove the Sample plate from the instrument.
  - Invert Binding Bead Mix to mix, then add 520 µL to each sample in the Sample Plate.  
**Note:** Remix the Binding Bead Mix by inversion frequently during pipetting to ensure even distribution of beads to all samples or wells. The mixture containing the Binding Beads is viscous. Therefore, pipet slowly to ensure that the correct amount is added. DO NOT use a repeat pipet to add to the samples as the high viscosity will cause variations in volume added.
  - Place the Sample Plate back onto the instrument, then start the run.
  - After the protocol is complete (~35 minutes after adding Binding Bead Mix), immediately remove the elution plate from the instrument and cover the plate or transfer the eluate to the final tube or plate of choice for final storage.

The purified nucleic acid is ready for immediate use. Alternatively, store the plate at -20°C for up to 6 months and -80°C for greater than 6 months.

## Perform Nucleic Acid purification using Bead plates and the KingFisher™ Flex

- 1 Set up the instrument**
- Ensure that the instrument is set up with the proper magnetic head and the proper heat block, as indicated in the following table.

Component	Type
Magnetic head	96 deep-well magnetic head
Heat block	96 well deep-well heat block

**IMPORTANT!** Failure to use the proper magnetic head and heat block results in lower yields and potential harm to the instrument.

- Ensure that the proper program has been downloaded from the product page and loaded onto the instrument:
  - Soil Samples: **MagMAX\_Microbiome\_Soil\_Flex**
  - Biofluid and other sample types: **MagMAX\_Microbiome\_Liquid\_Buccal\_Flex**

- 2 Set up the processing plates**

Set up the Wash, Elution, and Tip Comb Plates outside the instrument according to the following table.

Plate ID	Plate position	Plate type	Reagent	Volume per well
Wash 1 Plate	2	Deep-well	Wash Buffer	1,000 µL
Wash 2 Plate	3	Deep-well	Wash Buffer	1,000 µL
Wash 3 Plate	4	Deep-well	80% Ethanol	1,000 µL
Wash 4 Plate	5	Deep-well	80% Ethanol	1,000 µL
Elution Plate	6	Deep-well	Elution Solution	50 µL
Tip Comb	7	Place a 96 Deep-well Tip Comb in a Standard deep-well Plate		

**Note:** To prevent evaporation and contamination, cover the prepared processing plates with paraffin film or MicroAmp™ Clear Adhesive Film until they are loaded into the instrument.

- 3 Lyse sample**
- Centrifuge the 96-well bead plate at 3,700–4,000 rpm for 10 seconds to collect the beads on the bottom of the plate before opening the seal.
  - Open the seal of the bead plate, then add 800 µL of Lysis Buffer to each well.

### 3 Lyse sample (continued)

- c. Prepare samples according to the following table:

Sample type	Description
Soil Samples	Weigh out 200–250 mg, then place into prepared bead tube
Biofluid or Liquid sample	Remove 400–500 µL, then place into prepared bead tube
Skin/Buccal Swab	Remove the plastic stick, then place the swab into prepared bead tube
Bacterial/Yeast Culture	Remove 500 µL of the culture, then place into prepared bead tube

**Note:** For bacterial and Yeast cultures, and urine, user can centrifuge 1 ml then re-suspend the cell pellet in 800 µL of Lysis Buffer and transfer to the bead tube to start the extraction.

**Note:** Use a Kim wipe to wipe down the top of the wells and edges to remove any sample or bead material and ensure proper sealing and prevent leaks.

- d. Seal the bead plate with MicroAmp™ Clear Adhesive Film by pressing down around each well and the edges of the plate using your thumb or adhesive film applicator.

**Note:** For detailed instructions on plate sealing, see *MagMAX™ Bead beating plate guidelines Quick Reference* (MAN0018558).

- e. Repeat step 3d twice with foil seals.

**Note:** Ensure the edges and all the wells of the plate are sealed properly.

- f. Set the bead beater for 2 minutes, then clamp the plate into place.

- Omni Bead Ruptor 96: Set at 30 Hz for 2 minutes
- Mini Bead Beater 96: Set for 2 minutes
- Vortex with plate adaptor: Set at 2,000 rpm for 5 minutes

- g. Lyse the samples on the bead beater for 2 minutes.

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STOPPING POINT Bead Plate can be stored at 4°C overnight after lysis.

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### 4 Digest with Proteinase K

- a. Remove the bead plate from the instrument, then centrifuge for 5 minutes at 3,700 rpm.

**Note:** Do not remove the seal until the plate has been centrifuged after the bead beating.

- b. Transfer 400–500 µL of the Sample to the appropriate wells of a new deep-well plate.

This plate is the Sample Plate.

**Note:** For soil samples, transfer as much supernatant as possible without any particle carryover.

- c. Add 40 µL of Proteinase K to each sample.

- d. Select the correct program on the instrument:

- Soil Samples: **MagMAX\_Microbiome\_Soil\_Flex**
- Biofluid and other sample types: **MagMAX\_Microbiome\_Liquid\_Buccal\_Flex**

- e. Start the run, then load the prepared plates into position when prompted by the instrument.

### 5 Bind, wash, then elute total nucleic acid

- a. When prompted (approximately 20 minutes after the start of the protocol), remove the Sample plate from the instrument.

- b. Invert Binding Bead Mix to mix, then add 520 µL to each sample in the Sample Plate.

**Note:** Remix the Binding Bead Mix by inversion frequently during pipetting to ensure even distribution of beads to all samples or wells. The mixture containing the Binding Beads is viscous. Therefore, pipet slowly to ensure that the correct amount is added. DO NOT use a repeat pipet to add to the samples as the high viscosity will cause variations in volume added.

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**5** Bind, wash, then elute total nucleic acid *(continued)*

- c. Place the Sample Plate back onto the instrument, then start the run.
- d. After the protocol is complete (~35 minutes after adding Binding Bead Mix), immediately remove the elution plate from the instrument and cover the plate or transfer the eluate to the final tube or plate of choice for final storage.

The purified nucleic acid is ready for immediate use. Alternatively, store the plate at  $-20^{\circ}\text{C}$  for up to 6 months and  $-80^{\circ}\text{C}$  for greater than 6 months.

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## Perform Nucleic Acid purification using Bead tubes and the KingFisher™ Duo Prime

### 1 Set up the instrument

- a. Ensure that the instrument is set up with the proper magnetic head and the proper heat block, as indicated in the following table.

Component	Type
Magnetic head	12-tip magnetic head
Heat block	12 well deep-well heat block

**IMPORTANT!** Failure to use the proper magnetic head and heat block results in lower yields and potential harm to the instrument.

- b. Ensure that the proper program has been downloaded from the product page and loaded onto the instrument:
- Soil Samples: **MagMAX\_Microbiome\_Soil\_Duo**
  - Biofluid and other sample types: **MagMAX\_Microbiome\_Liquid\_Buccal\_Duo**

### 2 Lyse sample

- a. Set up the Vortex with the vortex adaptor.
- b. Add 800 µL of Lysis Buffer to the bead tubes.
- c. Prepare samples according to the following table:

Sample type	Description
Soil Samples	Weigh out 200–250 mg, then place into prepared bead tube
Biofluid or Liquid sample	Remove 400–500 µL, then place into prepared bead tube
Skin/Buccal Swab	Remove the plastic stick, then place the swab into prepared bead tube
Bacterial/Yeast Culture	Remove 500 µL of the culture, then place into prepared bead tube

**Note:** For bacterial and Yeast cultures, and urine, user can centrifuge 1 ml then re-suspend the cell pellet in 800 µL of Lysis Buffer and transfer to the bead tube to start the extraction.

- d. Vortex the bead tube upside down for 10 seconds to mix the sample with the buffer.
- e. Set the speed to 2,500 rpm, then place the tubes onto the vortex adaptor.

**Note:** We recommend the VWR 24 tube adapter and vortex for maximum nucleic acid yield. If an alternative instrument or adaptor is used, ensure that the instrument can meet the speed listed in the protocol.

- f. Lyse samples on the vortexer for 10 minutes.

**STOPPING POINT** Bead tube can be stored at 4°C overnight after lysis.

**3 Set up the Sample Plate and Elution Strip** Set up the Sample Plate and Elution Strip according to the following tables, respectively.

**Table 2** Sample plate

Row ID	Plate Row	Reagent	Volume per well
Sample row	A	Sample	Varies
Tip Comb	B	Tip Comb	Empty
—	C	Empty	
80% Ethanol	D	80% Ethanol	1,000 µL
80% Ethanol	E	80% Ethanol	1,000 µL
Wash Buffer	F	Wash Buffer	1,000 µL
Wash Buffer	G	Wash Buffer	1,000 µL
—	H	Empty	

**Table 3** Elution strip

Row ID	Plate Row	Reagent	Volume per well
Elution Solution	A	Elution Solution	50 µL

**Note:** To prevent evaporation and contamination, cover the prepared processing plate and elution strip with paraffin film or MicroAmp™ Clear Adhesive Film until they are loaded into the instrument.

- 4 Digest with Proteinase K**
- Remove tubes from vortexer, then centrifuge for 2 minutes at 14,000 × g.
  - Transfer 400–500 µL of the Sample to the appropriate wells of Row A in the Sample Plate.  
**Note:** For soil samples, transfer as much supernatant as possible without any particle carryover.
  - Add 40 µL of Proteinase K to each sample.
  - Select the correct program on the instrument:
    - Soil Samples: **MagMAX\_Microbiome\_Soil\_Duo**
    - Biofluid and other sample types: **MagMAX\_Microbiome\_Liquid\_Buccal\_Duo**
  - Start the run, then load the prepared plate and elution strip into position when prompted by the instrument.

- 5 Bind, wash, then elute total nucleic acid**
- When prompted (approximately 20 minutes after the start of the protocol), remove the Sample plate from the instrument.
  - Invert Binding Bead Mix to mix, then add 520 µL to each sample in Row A of the Sample Plate.  
**Note:** Remix the Binding Bead Mix by inversion frequently during pipetting to ensure even distribution of beads to all samples or wells. The mixture containing the Binding Beads is viscous. Therefore, pipet slowly to ensure that the correct amount is added. DO NOT use a repeat pipet to add to the samples as the high viscosity will cause variations in volume added.
  - Place the Sample Plate back onto the instrument, then start the run.
  - After the protocol is complete (~35 minutes after adding Binding Bead Mix), immediately remove the elution strip from the instrument and transfer the eluate to the final tube or plate of choice for final storage.

The purified nucleic acid is ready for immediate use. Alternatively, store the plate at –20°C for up to 6 months and –80°C for greater than 6 months.



# Perform Nucleic Acid purification using Bead plates and the KingFisher™ Duo Prime

## 1 Set up the instrument

- a. Ensure that the instrument is set up with the proper magnetic head and the proper heat block, as indicated in the following table.

Component	Type
Magnetic head	12-tip magnetic head
Heat block	12 well deep-well heat block

**IMPORTANT!** Failure to use the proper magnetic head and heat block results in lower yields and potential harm to the instrument.

- b. Ensure that the proper program has been downloaded from the product page and loaded onto the instrument:

- Soil Samples: **MagMAX\_Microbiome\_Soil\_Duo**
- Biofluid and other sample types: **MagMAX\_Microbiome\_Liquid\_Buccal\_Duo**

## 2 Set up the Sample Plate and Elution Strip

Set up the Sample Plate and Elution Strip according to the following tables, respectively.

**Table 4** Sample plate

Row ID	Plate Row	Reagent	Volume per well
Sample row	A	Sample	Varies
Tip Comb	B	Tip Comb	Empty
—	C	Empty	
80% Ethanol	D	80% Ethanol	1,000 µL
80% Ethanol	E	80% Ethanol	1,000 µL
Wash Buffer	F	Wash Buffer	1,000 µL
Wash Buffer	G	Wash Buffer	1,000 µL
—	H	Empty	

**Table 5** Elution strip

Row ID	Plate Row	Reagent	Volume per well
Elution Solution	A	Elution Solution	50 µL

**Note:** To prevent evaporation and contamination, cover the prepared processing plate and elution strip with paraffin film or MicroAmp™ Clear Adhesive Film until they are loaded into the instrument.

## 3 Lyse sample

- a. Centrifuge the 96-well bead plate at 3,700–4,000 rpm for 10 seconds to collect the beads on the bottom of the plate before opening the seal.
- b. Open the seal of the bead plate, then add 800 µL of Lysis Buffer to each well.

### 3 Lyse sample (continued)

- c. Prepare samples according to the following table:

Sample type	Description
Soil Samples	Weigh out 200–250 mg, then place into prepared bead tube
Biofluid or Liquid sample	Remove 400–500 µL, then place into prepared bead tube
Skin/Buccal Swab	Remove the plastic stick, then place the swab into prepared bead tube
Bacterial/Yeast Culture	Remove 500 µL of the culture, then place into prepared bead tube

**Note:** For bacterial and Yeast cultures, and urine, user can centrifuge 1 ml then re-suspend the cell pellet in 800 µL of Lysis Buffer and transfer to the bead tube to start the extraction.

**Note:** Use a Kim wipe to wipe down the top of the wells and edges to remove any sample or bead material and ensure proper sealing and prevent leaks.

- d. Seal the bead plate with MicroAmp™ Clear Adhesive Film by pressing down around each well and the edges of the plate using your thumb or adhesive film applicator.
- Note:** For detailed instructions on plate sealing, see *MagMAX™ Bead beating plate guidelines Quick Reference* (MAN0018558).
- e. Repeat step 3d twice with foil seals.
- Note:** Ensure the edges and all the wells of the plate are sealed properly.
- f. Set the bead beater for 2 minutes, then clamp the plate into place.
- Omni Bead Ruptor 96: Set at 30 Hz for 2 minutes
  - Mini Bead Beater 96: Set for 2 minutes
  - Vortex with plate adaptor: Set at 2,000 rpm for 5 minutes
- g. Lyse the samples on the bead beater for 2 minutes.

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STOPPING POINT Bead Plate can be stored at 4°C overnight after lysis.

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### 4 Digest with Proteinase K

- a. Remove the bead plate from the instrument, then centrifuge for 5 minutes at 3,700 rpm.
- Note:** Do not remove the seal until the plate has been centrifuged after the bead beating.
- b. Transfer 400–500 µL of the Sample to the appropriate wells of Row A in the Sample Plate.
- Note:** For soil samples, transfer as much supernatant as possible without any particle carryover.
- c. Add 40 µL of Proteinase K to each sample.
- d. Select the correct program on the instrument:
- Soil Samples: **MagMAX\_Microbiome\_Soil\_Duo**
  - Biofluid and other sample types: **MagMAX\_Microbiome\_Liquid\_Buccal\_Duo**
- e. Start the run, then load the prepared plate and elution strip into position when prompted by the instrument.

### 5 Bind, wash, then elute total nucleic acid

- a. When prompted (approximately 20 minutes after the start of the protocol), remove the Sample plate from the instrument.
- b. Invert Binding Bead Mix to mix, then add 520 µL to each sample in Row A of the Sample Plate.
- Note:** Remix the Binding Bead Mix by inversion frequently during pipetting to ensure even distribution of beads to all samples or wells. The mixture containing the Binding Beads is viscous. Therefore, pipet slowly to ensure that the correct amount is added. DO NOT use a repeat pipet to add to the samples as the high viscosity will cause variations in volume added.

**5** Bind, wash, then elute total nucleic acid *(continued)*

- c. Place the Sample Plate back onto the instrument, then start the run.
- d. After the protocol is complete (~35 minutes after adding Binding Bead Mix), immediately remove the elution strip from the instrument and transfer the eluate to the final tube or plate of choice for final storage.

The purified nucleic acid is ready for immediate use. Alternatively, store the plate at  $-20^{\circ}\text{C}$  for up to 6 months and  $-80^{\circ}\text{C}$  for greater than 6 months.

## Limited product warranty

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For descriptions of symbols on product labels or product documents, go to [thermofisher.com/symbols-definition](http://thermofisher.com/symbols-definition).

The information in this guide is subject to change without notice.

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**Revision history:** Pub. No. MAN0018070

Revision	Date	Description
C.0	27 July 2019	In contents and storage, changed For 1,000 reaction volume to For bulk reagents.
B.0	22 April 2019	Updated sku for bead plate in Contents and storage from A423331 to A42331.
A.0	17 March 2019	New document.

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