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**Package Contents**

Catalog Numbers:	Amount:
A35895	10 preps
A36228	20 preps

**Storage Conditions**

- Upon receipt, store Resuspension Buffer at 4°C, and all the other components at room temperature

**Required Materials**

- Thermo Scientific™ FastVac™ Vacuum Manifold with vacuum source capable 400 mm Hg pressure at the vacuum manifold
- Microcentrifuge capable of reaching  $\geq 10,000 \times g$  at room temperature
- Tubes with a minimum volume of 50 mL
- 1.5 mL centrifuge tubes
- Pipette for 200 to 400  $\mu\text{L}$
- 95–100% Ethanol

**Timing**

Bacterial culture: overnight  
Purification: 30 minutes

**Selection Guide**

Go online to view related products:  
**PureLink™ Nucleic Acid Purification Kit**

**Product Description**

- The PureLink™ Fast Low Endotoxin Maxi Plasmid Purification Kit enables isolation of high quality, low endotoxin (<1 EU/ $\mu\text{g}$ ), plasmid DNA ready for immediate use, avoiding the need for subsequent precipitation steps
- Isolate up to 1.5 mg of high quality, ultrapure plasmid DNA from 150 mL of bacterial culture
- DNA is free of RNA, salt, and protein, making it ideal for transfection, restriction endonuclease digestion, *in-vitro* transcription, PCR amplification, and DNA sequencing
- Colored buffers that permit error-free visualization of complete bacterial cell lysis and neutralization

**Important Guidelines**





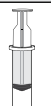


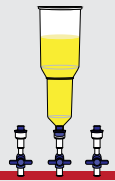
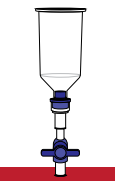
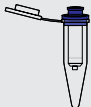
- The Lysis Buffer and Binding Buffer may have precipitant. If this occurs, dissolve the precipitate by incubating the bottles at 30–37°C for 10–20 minutes and mix by inversion
- Elution Buffer contains 10 mM Tris-HCl, pH 8.5, 0.1 mM EDTA. If required, pure water can also be used to elute the plasmid DNA
- DNA yield can be increased by pre-warming the Elution Buffer to 50°C and/or increasing the incubation period up to 5 minutes prior to centrifugation
- For low copy number plasmids or if higher concentration is desired, plasmid DNA can be eluted in as little as 200  $\mu\text{L}$

**Online Resources**

Visit our product page for additional information and protocols. For support, visit [thermofisher.com/support](http://thermofisher.com/support).

## Maxiprep plasmid isolation protocol

Before first use of the kit, add 88 mL of 95–100% ethanol to the 23 mL of Wash Buffer 2. Mark the label to indicate that ethanol is added.

Steps	Procedure Details
<b>1</b> Pellet the cells 	Sediment cells by centrifugation for <b>10 minutes</b> at $\geq 3,500 \times g$ , then discard the supernatant.
<b>2</b> Add Resuspension Buffer 	Add <b>14 mL</b> of Resuspension Buffer (red) to the cell pellet and resuspend by vortexing or pipetting.
<b>3</b> Add Lysis Buffer 	Add <b>14 mL</b> of Lysis Buffer (blue) and mix by inverting 6 times. Do not vortex. Incubate at room temperature for <b>3 minutes</b> . Lysis is complete when the mixture turns dark purple and viscous.
<b>4</b> Add Precipitation Buffer 	Add <b>14 mL</b> of Precipitation Buffer (yellow) and mix by inverting 6 times. Do not vortex. The sample will turn yellow when neutralization is complete.
<b>5</b> Load the lysate 	Load the lysate into the syringe filter and wait <b>5 minutes</b> until the precipitate has floated to the top.
<b>6</b> Filter the lysate 	Remove the lock and filter the lysate into a fresh 50 mL tube. Do not use excess pressure. Save this clarified lysate.
<b>7</b> Add Binding Buffer 	Add <b>14 mL</b> of Binding Buffer to the clarified lysate and mix by inverting 10 times.
<b>8</b> Bind DNA to the column 	Add the mixture into the column assembly and turn on the vacuum until all the liquid has passed through the column.
<b>9</b> Wash DNA 	Remove the 50 mL reservoir and wash with <b>5 mL</b> of Wash Buffer 1 (once), and then with <b>5 mL</b> of Wash Buffer 2 (twice), using the vacuum manifold. Turn off the vacuum between washes.
<b>10</b> Elute DNA 	Remove and discard the 15 mL conical reservoir. Transfer the column to a 1.5 mL collection tube, then centrifuge at $\geq 10,000 \times g$ for 1 minute to remove any residual Wash Buffer. Transfer the column to a fresh 1.5 mL tube. Add <b>200 <math>\mu</math>L</b> of Elution Buffer twice, allowing the first 200 $\mu$ L to absorb in the membrane before dispensing the last 200 $\mu$ L, and incubate for 2 minutes, then centrifuge at $\geq 10,000 \times g$ for 1 minute to elute the DNA.

The information in this guide is subject to change without notice.

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