

AmpFℓSTR™ Yfiler™ PCR Amplification Kit

Catalog Number 4359513

Pub. No. 4359563 Rev. K

Note: For safety and biohazard guidelines, see the “Safety” appendix in the *AmpFℓSTR™ Yfiler™ PCR Amplification Kit User Guide* (Pub. No. 4358101). Read the Safety Data Sheets (SDSs) and follow the handling instructions. Wear appropriate protective eyewear, clothing, and gloves.

IMPORTANT! Reagents used for amplification must be isolated from any source of contaminating DNA, especially from previously amplified AmpFℓSTR™ PCR products and the AmpFℓSTR™ Allelic Ladders.

Thaw reaction mix, primer set, control DNA, and allelic ladder completely before first use. To avoid freeze/thaw cycles, store these components at 2 to 8°C after initial use. Store enzyme at –15 to –25°C.

The AmpFℓSTR™ Yfiler™ Primer Set and Allelic Ladder contain primers that are labeled with light-sensitive dyes. Protect the primers and allelic ladder from light; do not leave them exposed on the laboratory bench.

To obtain optimal amplification and typing results, follow the procedures in the *AmpFℓSTR™ Yfiler™ PCR Amplification Kit User Guide* (Pub. No. 4358101).

Contents and storage

Cat. No.	Contents	Quantity	Storage conditions
4359517	AmpFℓSTR™ Yfiler™ PCR Reaction Mix Contains MgCl ₂ , dATP, dGTP, dCTP, and dTTP, bovine serum albumin, and 0.05% sodium azide in buffer and salt.	1 tube, 1.1 mL	–15 to –25°C upon receipt. 2 to 8°C after initial use.
4359515	AmpFℓSTR™ Control DNA 9947A, Contains 10 ng/μL human female cell line DNA in 0.05% sodium azide and buffer.	1 tube, 0.025 mL	
4307984	AmpFℓSTR™ Control DNA 007, Contains 0.10 ng/μL human male genomic DNA in 0.05% sodium azide and buffer.	1 tube, 0.3 mL	
4359519	AmpFℓSTR™ Yfiler™ Primer Set Contains locus-specific 6-FAM™, VIC™, NED™, and PET™ dye-labeled and unlabeled primers in buffer. The primers amplify the Y-STR loci DYS19, DYS385 a/b, DYS389 I/II, DYS390, DYS391, DYS392, DYS393, DYS437, DYS438, DYS439, DYS448, DYS456, DYS458, DYS635 (Y GATA C4), and YGATA H4.	1 tube, 0.55 mL	–15 to –25°C upon receipt. 2 to 8°C after initial use. Store protected from light.
4373675	AmpFℓSTR™ Yfiler™ Allelic Ladder Contains the following amplified alleles: <ul style="list-style-type: none"> 6-FAM™ dye (blue): DYS389I 10–15; DYS389II 24–34; DYS390 18–27; DYS456 13–18. VIC™ dye (green): DYS19 10–19; DYS385 7–25, DYS458 14–20. NED™ dye (yellow): DYS391 7–13; DYS392 7–18; DYS393 8–16; DYS439 8–15; DYS635 (Y GATA C4) 20–26. PET™ dye (red): DYS437 13–17; DYS438 8–13; DYS448 17–24; Y GATA H4 8–13. 	1 tube, 0.05 mL	–15 to –25°C upon receipt. 2 to 8°C after initial use. Store protected from light. IMPORTANT! The allelic ladder contains PCR products. Do not amplify. To avoid contamination, store the allelic ladder separate from the other kit components and unamplified DNA.
4385214	AmpliAq Gold™ DNA Polymerase, enzyme activity 5 U/μL	2 tubes, 0.05 mL/tube	–15 to –25°C.

Materials required but not provided

Cat. No.	Component
4322682	GeneScan™ 500 LIZ™ Size Standard, 2 tubes, 200 µL/tube
or	or
4408399	GeneScan™ 600 LIZ™ Size Standard v2.0, 2 tubes, 200 µL/tube
4358101	<i>AmpFℓSTR™ Yfiler™ PCR Amplification Kit User Guide</i>

Performance characteristics

The AmpFℓSTR™ Kits are developed and manufactured by Thermo Fisher Scientific. We test each lot of kits to ensure that the kits perform according to specifications and are free from interfering contaminants.

Each AmpFℓSTR™ Yfiler™ PCR Amplification Kit contains reagents optimized to amplify and type 0.5 ng of AmpFℓSTR™ Control DNA 007.

The recommended range of input sample DNA is approximately 0.5 to 1.0 ng of human DNA. However, we have successfully typed samples containing less than 0.5 ng of human DNA.

Follow the protocol in the *AmpFℓSTR™ Yfiler™ PCR Amplification Kit User Guide* to collect and analyze data on the instruments and software supported for use with this kit.

Limited product warranty

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Life Technologies Ltd | 7 Kingsland Grange | Woolston, Warrington WA1 4SR | United Kingdom

For descriptions of symbols on product labels or product documents, go to thermofisher.com/symbols-definition.

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Revision history: Pub. No. 4359563

Revision	Date	Description
K	22 October 2018	Updated branding and trademarks, no technical changes.
J	August 2012	Basis for this revision.

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